Malaria is a major world health problem with hundreds of millions of cases and millions of deaths per year. Research on malaria concentrates on *Plasmodium falciparum*: this parasite is the major cause of the fatal disease. This review highlights research in the areas of vaccine development, pathophysiology of disease, drug resistance, and drug therapy.

**Vaccine development**

Vaccine research in malaria targets three stages of the parasite life cycle: 1) the preerythrocytic stages, encompassing sporozoite and liver stage parasites; 2) the blood-stage cycle of clinical infection; and 3) the sexual stages of the parasite in the mosquito. The goal of the preerythrocytic vaccines is sterile immunity against the early stages of infection to prevent the symptomatic blood stage disease. Vaccines aimed at the malaria blood stages could seek sterile immunity as well, but might also be effective if they contained the infection and prevented the most severe sequelae. Vaccines directed at the sexual stages prevent development in the mosquito. They do this by inducing human antibodies against parasite antigens expressed in the insect vector. These antibodies are ingested by the mosquito with the infected blood meal and interfere with parasite development.

**Preerythrocytic vaccines**

The most studied antigen from the preerythrocytic phases of the life cycle is the circumsporozoite protein, which covers the surface of the sporozoite. The circumsporozoite antigen repeat motif is the predominant epitope recognized by antibodies in the serum of sporozoite immunized animals. Transfer of monoclonal antibodies directed at this repeat epitope can protect mice against sporozoite challenge. However, polyclonal serum antibodies are much less protective than the monoclonal antibodies. Do Rosano et al. [1] has now shown that immune serum may actually aid the parasite during its development in the mosquito. They infected mosquitoes with *P. falciparum* and let them feed on blood containing antismerozoite antibodies. The number of sporozoites per mosquito more than doubled. Furthermore, *in vitro* liver cell invasion by these sporozoites was now unaffected by immune serum that otherwise neutralized sporozoites. The authors concluded that the effects on transmission of malaria should be considered when assessing any candidate vaccine.

Recent work has focused on the responses of T cells to the circumsporozoite protein because it now appears that immune T cells can kill pre-erythrocytic forms of malaria. Hoffman et al. (Science 1987, 237:639) had previously reported results from a prospective study of adults in Kenya that showed no correlation of anticircumsporozoite serum antibody with protection from malaria. Now Hoffman et al. [2] report that in this group, there was a correlation of protection with T-cell proliferative responses to a short segment of the *P. falciparum* circumsporozoite protein. Although this study was small, it is perhaps significant that this same segment contains an epitope recognized by cytotoxic T cells, which are important immune effector cells in the animal models of sporozoite immunity.

Romero et al. [3] showed that T cells reactive with the circumsporozoite molecule can protect mice against sporozoite infection. They first identified a cytotoxic T-cell epitope on the *Plasmodium berghei* circumsporozoite protein and then raised CD8+ T-cell clones against this short peptide. Transfer of several of these clones protected mice against sporozoite challenge. Very large numbers of cells were needed to protect these animals, but it is encouraging that the circumsporozoite protein contains potentially protective epitopes for T-cell immunity.

Hoffman et al. [4], in another paper, explored the role of CD8+ T cells in the immune response to liver stages of malaria. They reported that immune mice develop cellular infiltrates in their livers after sporozoite chal-
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<td>Weiss WR, Hoffman SL</td>
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challenge. They transferred immune spleen cells, and were able to find similar infiltrates in recipient animals after challenge. Depleting immune spleens of CD8+ cells before transfer abolished the infiltrates. Immune spleen cells could also inhibit parasite growth in cultured liver cells. These papers by Hoffman et al. [3] and Romero et al. [4] emphasized the importance of T-cell responses in protective immunity to the early stages of malaria infection. However, much remains to be learned about antigens and immune mechanisms before a vaccine based on T-cell immunity can be a reality in humans.

Blood-stage vaccine antigens

A large research effort aims at vaccine against the blood stages of malaria. The symptoms and lethal complications of malaria begin when the parasite ruptures from the liver and starts its cycles of erythrocyte infection. A vaccine that would mute or completely prevent this stage of the infection is thus very attractive. Furthermore, there is evidence from epidemiologic and clinical studies that some natural immunity to blood stage infections is acquired by persons living in malaria endemic areas. Several papers published this year dissected the immune responses to candidate malaria blood stage antigens.

Troy-Blomberg et al. [5] studied T- and B-cell responses to Pf155:RESA. This antigen is deposited on the erythrocyte surface by invading Plasmodium falciparum merozoites, and antibodies to it will block invasion in vitro. Fourteen synthetic peptides, corresponding to predicted T-cell epitopes, were tested for reactivity with cells and serum from Gambians living in a malarious area. Two peptides stimulated T-cell responses from 61% and 60% of the 46 donors, while 9% responded to crude whole Pf155 antigen. Serum antibodies likewise reacted with many peptides, and for individual donors, there appeared to be an inverse correlation between B- and T-cell responses to the Pf155:RESA repeat regions. T-cell reactivity was maintained even after the season of malaria transmission, while antibody titers waned. This work provides the immunologic groundwork for designing a synthetic vaccine based on the Pf155:RESA sequences.

Burns et al. [6] studied another merozoite antigen in the Plasmodium yoelii rodent model. The major merozoite surface antigen coats the merozoite and passive transfer of monoclonal antibodies to this antigen protect mice against P. yoelii blood infection. Using an Escherichia coli expression system, the epitope of this protective monoclonal was mapped to a cysteine-rich fragment of the precursor major merozoite surface antigen. The epitope was destroyed by reduction and presumably is conformationally dependent on cross-linking of the cysteines. This region is highly conserved between malaria species, including P. falciparum, and is another promising candidate antigen for a blood stage malaria vaccine.

Marsh et al. [7] studied antibodies to blood stage antigens of P. falciparum in relation to parasitemia and disease. They followed 134 Gambian children and found that only the response to parasite-dependent red blood cell neoantigens, as measured by erythrocyte agglutination, was associated with lower incidence of clinical malaria. Interestingly, children reacting to these antigens had less illness but the same parasite levels as their nonreactive peers. This finding underlines the dichotomy between infection and clinical disease in malaria and hints that a vaccine to prevent illness may not have to produce sterile immunity.

Transmission-blocking vaccine antigens

Transmission-blocking vaccines are designed to kill malaria in the mosquito. Human antibodies directed against the sexual stages of the parasite are ingested by the mosquito and react with the developing stages in the mosquito gut. Kaslow et al. [8] studied one such target antigen, Pfs25. He sequenced the Pfs25 gene from eight geographically distinct isolates of P. falciparum and found that they were identical except for one example of a single conservative amino acid change. This is encouraging as variation of malaria antigens is notorious. Protective transmission-blocking antibodies against this antigen should be effective against all strains.

Pathophysiology of malaria

Adhesion molecules

In Plasmodium, the mature stages of infected erythrocytes do not circulate freely but bind to capillary and venular epithelium. This prevents the infected cells from being filtered by the spleen, and also may contribute to the vascular plugging that is the hallmark of cerebral malaria. Thus, the molecular basis of the adhesion of parasitized cells is of great interest.

Erythrocytes infected by P. falciparum have structures called "knobs" at their surface, and it was thought that these knobs were the site of binding to endothelium. Udomsangpetch et al. [9] discovered a knobless P. falciparum that retains its adherent properties in vitro assays. Thus, the knob is merely a correlate of the true adherence molecules on the surface of the parasitized erythrocyte. The nature of the true adhesion molecule remains unknown.

Two naturally occurring molecules on endothelium, thrombospondin and CD36, have been proposed as host ligands for the parasite adhesion molecules. Shepherd et al. [10] found that purified thrombospondin could bind parasitized erythrocytes, but that binding to melanoma cells in vitro did not correlate with thrombospondin on their surface. Oquendo et al. [11] cloned the human CD36 gene and showed that transfected cells were able to bind parasitized erythrocytes. Ochonhouse et al. [12] purified CD36 from platelets and found that plastic coated with this product bound parasitized erythrocytes. Most significantly, addition of sol-
uble CD36 was then able to reverse the binding. Treatment for cerebral malaria using analogues of CD36 may thus be a possibility in the future.

Cytokines in severe malaria

Encephalopathy due to *P. falciparum* is a major cause of mortality and is thought to be due to vascular damage in the brain. As mentioned earlier, mature parasitized erythrocytes adhere to and clog small vessels in the brain, and may be the cause of the vascular lesion. An intriguing alternative hypothesis is that the vascular lesions are not due solely to erythrocyte plugging, but involve the toxic effects of a circulating cytokine, tumor necrosis factor (TNF). Some strains of mice infected with rodent malarial develop cerebral malaria and vascular lesions without plugging by infected erythrocytes. Instead leukocytes pack the damaged vessels. This lesion in mice can be prevented by *in vivo* treatment with antibody to the lymphocyte CD4 antigen or by antibody to TNF.

Grau *et al.* have published two new papers on TNF in malaria. In the first [13], cerebral malaria in the mouse was prevented by treatment of animals with antibodies to interferon-γ, which inhibited TNF production by macrophages. His model of immunopathology in mice is now that CD4+ lymphocytes release lymphokines, including interferon-γ. These lymphokines induce TNF production from macrophages, and TNF is responsible for the lesions in the cerebral vessels.

In the second paper Grau *et al.* [14] related TNF levels to cerebral malaria in humans. They reported on a study of 65 severely ill Malawian children, and showed that serum TNF levels on hospital admission were high and correlated with young age, high parasite counts, hypoglycemia, and death. Thus, TNF may be involved in human as well as rodent cerebral malaria. This opens the exciting prospect of anti-lymphokine therapy for cerebral malaria, an otherwise poorly treatable condition.

Mechanisms of drug resistance

Resistance of *P. falciparum* to chloroquine has become a major therapeutic problem. It appears that the parasite has evolved a mechanism to excrete the drug, which renders it insensitive to normal therapeutic drug levels. Calcium channel blocking drugs inhibit this excretion, and the parasite once again becomes sensitive to chloroquine. This reversal by calcium channel blockers is similar to multidrug resistance in mammalian tumors, in which *P. falciparum* pumps chemotherapeutic agents out of cells making the tumor drug resistant. Calcium channel blockers inhibit the pump and restore drug sensitivity.

Two groups reported finding a gene in *P. falciparum* that is an analogue of the mammalian multidrug resistance gene [15,16]. This gene, Pfmdr, exists in all parasite strains but has a larger number of copies in some chloroquine-resistant parasites. Some resistant strains also have an increased amount of translated messenger RNA from the Pfmdr gene. This is all consistent with Pfmdr being the mechanism of chloroquine resistance in at least some instances.

Progress has also been made in understanding the mechanism of drug resistance to pyrimethamine [17]. This drug is an inhibitor of dihydrofolate reductase, an important enzyme in nucleic acid synthesis. Using genetic crosses between resistant and susceptible strains of *P. falciparum*, resistance to pyrimethamine was localized to chromosome 4 where the dihydrofolate reductase gene is situated. Resistance correlated with a single point mutation producing an asparagine at position 108, which the authors believe inhibits the binding of pyrimethamine. This knowledge of the mechanisms of drug resistance may lead to modifications that will increase the effectiveness of these antimalarials.

Treatment of severe malaria

Although mild cases of malaria respond to oral therapy, treatment of severe malaria can be complex because the side effects of therapies may exacerbate the hypoglycemia and circulatory collapse that is caused by malaria itself. Three papers addressed the pharmacology of quinine, chloroquine, and quinidine with an eye to minimizing their side-effects.

Molyneux *et al.* [18] studied dose rates of intravenous quinine in children with cerebral malaria. They found that quinine-induced insulin release and hypoglycemia could be avoided if the drug was infused over 3 hours instead of 1.

White *et al.* [19] reported on various routes and dosages of chloroquine in children with severe malaria. He studied intravenous, subcutaneous, intramuscular, and nasogastric methods of drug delivery, and found adequate serum drug levels achieved by all routes. Hypotension occurred most commonly in the higher dose parenteral regimens, although resolution of parasitemia and fever was similar in all groups. The authors recommended that when possible, chloroquine should be given by intravenous infusion but may also be given safely by multiple frequent injections.

Miller *et al.* [20] reviewed the US experience treating patients with severe malaria, using intravenous quinidine and exchange transfusion. Continuous intravenous quinidine was effective in decreasing parasite levels, and cardiac abnormalities, tinnitus, and vomiting were associated with elevated serum drug levels in a few cases. Exchange transfusion of up to 10 units of blood dropped parasite counts quickly. They recommend a protocol using both modalities where adequate hospital resources are available.
Annotated references and recommended reading

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Resistance to pyrimethamine is found to correlate with a point mutation resulting in an amino acid change at the putative binding site for the drug. This may lead to the design of more effective compounds.


Hypoglycemia due to quinine could be avoided using slower rates of infusion.


The definitive study of the pharmacology of chloroquine use for the treatment of malaria in children. They find that the drug is rapidly absorbed by a variety of routes and can be safely given if dosing is not done too rapidly.


A review of recent cases of severe malaria in the United States known to the Centers for Disease Control. Intravenous quinidine with or without exchange transfusion was effective therapy for severe malaria.