

AD _____

Award Number: W81XWH-~~€~~ ~~€~~ €H

TITLE: V@ÁQ] æ&f Á-æÓ [{ { [] } Á ÖT GÁPÚÁ } Á@Á^} • äãã Á -Ái^æ cÓæ &^!Á Ái^æf ^}c

PRINCIPAL INVESTIGATOR: Sã Ápá • @a|á

CONTRACTING ORGANIZATION: UT ÖPREÜ [à^!cÁ [[áÁ @ • [] Á ^áæÁ&@ [[]
ÁJã &ææ æ ÊPÁE | | |

REPORT DATE: June 20FF
Á

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for public release; distribution unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE			<i>Form Approved</i> <i>OMB No. 0704-0188</i>		
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.					
1. REPORT DATE (DD-MM-YYYY) 01-06-2011		2. REPORT TYPE Annual		3. DATES COVERED (From - To) 7 MAY 2010 - 6 MAY 2011	
4. TITLE AND SUBTITLE The Impact of a Common MDM2 SNP on the Sensitivity of Breast Cancer to Treatment			5a. CONTRACT NUMBER		
			5b. GRANT NUMBER W81XWH-07-1-0403		
			5c. PROGRAM ELEMENT NUMBER		
6. AUTHOR(S) Kim Hirshfield E-Mail: hirshfie@umdnj.edu			5d. PROJECT NUMBER		
			5e. TASK NUMBER		
			5f. WORK UNIT NUMBER		
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) UMDNJ-Robert Wood Johnson Medical School Piscataway, NJ 08854			8. PERFORMING ORGANIZATION REPORT NUMBER		
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSOR/MONITOR'S ACRONYM(S)		
			11. SPONSOR/MONITOR'S REPORT NUMBER(S)		
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT The discovery of a single nucleotide polymorphism (SNP) in the mdm2 promoter uncovered a previously unknown role of this SNP in predicting early onset of breast and the possibility that this germ line variation could decrease the effectiveness of treatment. These outcomes are likely due to the increased expression of mdm2 protein in SNP309 individuals, which blunts the p53-mediated apoptotic response to DNA damage. The objective of this proposal is to test the hypothesis that SNP309 decreases the effectiveness of radiation and chemotherapy in breast cancer and that this negative impact can be overcome by targeted down-regulation of mdm2. There appears to be a trend toward excess contralateral events with the variant and enrichment of the variant in ER+ breast cancer recurrences. We observed that anti-estrogen agent, fulvestrant, causes a decrease in mdm2 protein half-life, leading to a reduction in mdm2 following treatment with this agent. We demonstrate that combined use of fulvestrant with chemotherapeutic drugs doxorubicin, etoposide and paclitaxel can enhance the sensitivity of breast cancer cells to these cytotoxic agents. We observed that mdm2 expression is differentially modulated by estrogen, the anti-estrogen tamoxifen, and genistein in a genotype-specific manner. The largest effects on reduction in mdm2 expression at the protein level occur in the mdm2 SNP309 cell line. We will continue to explore mechanistic studies in vitro while evaluating the clinical outcome associations of SNP309 to chemotherapy, hormonal therapy and radiation therapy.					
15. SUBJECT TERMS mdm2, breast cancer, polymorphisms					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON USAMRMC
a. REPORT U	b. ABSTRACT U	c. THIS PAGE U			19b. TELEPHONE NUMBER (include area code)
			UU	30	

Table of Contents

	<u>Page</u>
Introduction.....	4
Body.....	4
Key Research Accomplishments.....	12
Reportable Outcomes.....	11
Conclusion.....	12
References.....	13
Appendices.....	16
Supplementary Data.....	17

INTRODUCTION

The recent discovery of a single nucleotide polymorphism (SNP) in the mdm2 promoter uncovered a previously unknown role of this SNP in predicting early onset of breast and the possibility that this germ line variation could decrease the effectiveness of treatment. These outcomes are likely due to the increased expression of mdm2 protein in SNP309 individuals, which blunts the p53-mediated apoptotic response to DNA damage. The objective of this proposal is to test the hypothesis that SNP309 decreases the effectiveness of radiation and chemotherapy in breast cancer patients and that this negative impact can be overcome by targeted down-regulation of mdm2. The rationale in support of these objectives are molecular epidemiological data showing that individuals harboring SNP309 are at increased risk for early onset breast cancer, and laboratory studies showing that SNP309 decreases the activity of DNA damaging agents. If we are to achieve better results of treatment for patients with breast cancer, the choice of treatment must eventually benefit from a more precise understanding of the genetic abnormalities that are present in each individual's tumor. Using the same dose of drug or amount of radiation for each breast cancer patient cannot possibly be consistent with our understanding of modern molecular medicine. For example, subtle variations in our genetic code (called single nucleotide polymorphisms, [SNPs "snips"]) exist in the human population and make us susceptible to certain diseases and resistant to others. Similarly, these polymorphisms can make us more or less sensitive to treatment. Since these polymorphisms exist both in breast cancer and in normal tissues, understanding their impact on both the patient and the tumor will eventually guide the choice and dose of drug and amount of irradiation. Therefore, our objective is to improve the ways in which patients with breast cancer are evaluated and treated through an understanding of subtle variations in the human genome. The proposal brings together a team of molecular biologists/epidemiologists, pharmacologists, radiation and medical oncologists, and statisticians to focus on this novel approach to breast cancer treatment.

BODY

Task 1. Determine the impact of mdm2 SNP309 on the results of breast irradiation

Updating and assuring complete clinical data has been ongoing. Paperwork for IRB in accordance with recommendations from the IRB at CINJ and the human investigations committees of the DOD was completed and IRB-approval obtained. Patient accrual was initiated through the Radiation Oncology Clinics.

We have completed analysis of mdm2 on the cohort of patients whom we have long term follow-up. We confirmed an association of SNP309 with young patient age in the population of over 250 patients previously treated with long-term follow-up. While all patients in the previously treated database were in a younger age group, a larger percentage of patients of the GG genotype were under age 40 compared to the TT/TG genotypes (65% vs 35%, $p < 0.01$). We also found a correlation with race, with few African American patients having the GG homozygous genotype at SNP309. There were no other strong correlations between the SNP309 status and clinical-pathologic variables such as histology, ER status, Her2 status, nodal status, T-stage, family

history. There did not appear to be strong correlations with local-regional outcome in this dataset. There appears to be a trend toward excess contralateral events with a 10-year event rate of 9% in the TT/TG subset compared to over 20% in GG carriers. In addition, in this data set there was a difference in distant metastasis in the GG subtypes, with the 10 year rate of distant metastasis-free survival 89% in the TT/TG subset compared to 76% in the GG subtype ($p=0.04$). This will be further explored in multivariate analysis. Although there were no clear differences in local control, further exploratory subset analysis will be performed to determine if there are subsets within this cohort with higher local relapse rates.

In the prospective cohort, we continue to recruit patients in the radiation therapy clinic as well as in CINJ breast clinic. In the radiation therapy clinic we continue to actively accrue patients and continue accrual in the CINJ clinics such that we will have reached our accrual goals of patients treated with breast conserving surgery and radiation by years end. We will then analyze this larger cohort for SNP309 and evaluate outcomes and clinical-pathologic correlations over the next year.

Immunohistochemical Analysis of MDM2

Cells suffering DNA damage ultimately progress through apoptotic pathways, in which p53 plays a central role. Murine double minute 2 (MDM2) and its related ortholog MDM4, are involved in the negative regulation of p53 including ubiquitin-mediated targeting of p53 for proteolytic degradation and transcriptional activity. Moreover, MDM2 is induced by p53 thus creating a negative feedback loop (Wu, 1993; Barak, 1993; Kussie, 1996; Lin, 1994; Freedman, 1999). Apart from its p53 ubiquitination function, MDM2 has other functions including nuclear-cytoplasmic shuttling of p53 and prominent interactions with various ribosomal proteins (Roth, 1998; Marechal, 1994). MDM4 has similar structure and function to MDM2; however, it can be degraded by MDM2 (Shvarts, 1997; Shvarts, 1996; Okamoto, 2005). Appropriate expression of p53 propels cells down apoptotic pathways, but this progression may be counteracted by overexpression of MDM2/MDM4 and subsequent degradation of p53. It follows that dysregulation of p53 by MDM2/MDM4 could potentially cause radiation resistance through inability of the cell to undergo p53-mediated apoptosis.

Several studies have reported the significance of MDM2 in cancers of the prostate, breast and ovary (Marchetti, 1995; Marchetti, 1995; McCann, 1995; Khor, 2005). In breast cancer, MDM2 has been extensively studied as prognostic marker for overall and disease specific survival. Over expression of MDM2 has found to be associated with worse breast cancer specific survival and has been demonstrated to have a role in enhancing estrogen receptor alpha ($ER\alpha$) mediated gene expression and altering $ER\alpha$ stability (Kim, 2011; Turbin, 2006; Marchetti, 1995; Duong, 2007). However, the significance of MDM2 on local recurrence of breast cancer has not been adequately explored with most reports focusing on either recurrence-free survival or overall survival (Khor, 2005). Specifically, there have been no reports assessing the significance of MDM2 on local recurrence in early stage breast cancer treated with BCS + RT. Additionally, relatively little is known regarding the effects of over/under expression of MDM4 and p53 on local recurrence.

Because MDM2 SNP309 correlates with expression of *mdm2* (Bond, 2005), we also explored whether *mdm2* expression correlated with in-breast, regional (lymph node), and distant

recurrence of breast cancer in women treated for early stage breast cancer. MDM4, another negative regulator of p53, and p53 itself were also evaluated for prognostic value in this cohort. This analysis was limited as germline DNA from peripheral blood was not available for genotyping. Genotyping was not performed on tumor tissue as patient tissue did not have consent for such analysis. Breast cancer tissue cores were compiled into a tissue microarray (TMA), n=514, that was evaluated for MDM2, MDM4, p53. All patients had histological evidence of invasive breast carcinoma with early stage (I/II) disease and were treated with breast conserving surgery and radiation. The size of the primary tumor was considered to be the largest tumor diameter reported by the pathologist after surgical excision. Margin status was defined as positive if tumor cells were present on the most peripheral slide of the tumor. Following surgery, patients received standard whole breast irradiation to a total median dose to the breast of 48 Gy and a total tumor bed dose of 64 Gy; regional nodes were treated to a median dose of 46 Gy, as clinically indicated. Adjuvant systemic chemotherapy and/or adjuvant hormone therapy was administered as clinically indicated in accordance with standard practices during this time interval. Local recurrence was defined as clinically and biopsy-proven relapse in the ipsilateral breast. Ipsilateral breast recurrence free time was defined from the time of initial diagnosis to ipsilateral breast tumor relapse; nodal relapse free time was defined as the time from initial diagnosis to the time of biopsy proven nodal relapse; locoregional recurrence free time was defined as time from initial diagnosis to either ipsilateral breast recurrence or nodal recurrence. Descriptive statistics comparing MDM2 expression with conventional markers of tumor aggressiveness were analyzed by standard chi-squared tests, or, when appropriate, Fisher's exact test. Estimates of disease-free survival were calculated by the Kaplan–Meier product-limit method and the differences were assessed by the log-rank test. Probabilities of survival were calculated from the date of breast cancer diagnosis to either the date at which relapse was clinically identified or the date of last contact. Univariate and multivariate survival analysis was carried out using Cox's proportional hazard regression model. Multivariate analysis was used to assess the independent contribution of each variable to survival, statistically significant. A computer program package SAS (Version 9.2, SAS Institute, Cary, NC) was used for all statistical testing and management of the database.

Results of Immunohistochemical Analysis

Descriptive statistics and correlations with clinicopathologic markers

The description of the entire patient cohort is shown in Table 1. The median age at diagnosis of the entire patient cohort was 55 years (range 25-88 years) with 40% of patients being younger than 50 at the time of diagnosis. 63% (n= 183), 29% (n= 86) and 10% (n= 40) of the patient population were ER, PR and HER2/*neu* positive respectively. Twenty-five percent (n= 72) of the patient population was negative for all three markers. 46% (n= 140) of patient population received adjuvant hormonal therapy and 36% (n= 108) of the patient population received adjuvant chemotherapy. Fourteen percent (n= 72) received both adjuvant hormone and chemotherapy. As of September 2009, median follow-up on this cohort was 7.23 years during which 17.5% (n=90) of patients died. Fifty patients (9.7%) experienced ipsilateral breast recurrence, 2.3% (n=12) experienced nodal relapse and 11.3% (n=58) experienced locoregional recurrence.

Immunohistochemical staining results

The predominant intracellular staining of MDM2 was cytoplasmic. Immunoreactivity was completely absent in some tumor cores while in others, the number of immunoreactive cells

ranged from very few to the majority of cells. Samples of both positive and negative cores and slides are shown in Figure 1. Of 26 cores, 8.6% were scored as positive for MDM2 staining while 91.4% were scored as negative. Staining for MDM4 was less specific with 56.7% ($n=174$) of tumor cores being scored as positive (data not shown). Immunoreactivity for MDM4 encompassed a range from very few cells stained positive to almost all cells being stained positive. Staining for MDM4 was predominantly nuclear. Thirty-one percent ($n=140$) of patients stained positive of p53 with the staining being predominantly nuclear (data not shown).

Association between MDM2 and patient outcomes

Ten-year survival analysis was performed for ipsilateral breast recurrence free survival (IBRFS), nodal recurrence free survival (NRFS), and locoregional recurrence free (LRFS) as a function of MDM2, MDM4 and p53 expression. Only MDM2 was found to be a significant predictor of IBRFS ($p=0.0319$) and LRFS ($p=0.0165$) by log rank test. The corresponding survival curves are shown in Figure 2.

Univariate analysis was performed using MDM2, age, race, ER status, PR status, HER2/*neu*, systemic therapy, triple negative status, nodal status, tumor size and margin status. Relevant results are displayed in Table 2. When assessing for IBRFS, only tumor size, margin status, and MDM2 positivity were found to be significant ($p=0.0002$; $p=0.0137$; $p=0.0416$, respectively). When assessing for LRFS, tumor size, margin status and MDM2 were again found to be significant ($p=0.0001$; $p=0.0367$; $p=0.0229$, respectively). It should be noted that while nodal status approached significance for NRFS ($p=0.0686$), no variable was significant for NRFS possibly due to the small number of events. Considering that MDM2 was the only marker that was significant on univariate and log-rank tests, it was the only IHC marker used for multivariate analysis. Multivariate analysis was done for IBRFS and LRFS using the three variables that were significant in univariate analysis: margin status, tumor size and MDM2 expression. Nodal status was also included as it approached significance for significance for nodal recurrence on univariate analysis; which in turn may have an impact on LRFS (table 3). MDM2 was again found to be a significant predictor of IBRFS and LRFS ($p=0.0009$ and 0.0003 respectively). Additionally tumor size was once again found to be independently predictive of IBRFS and LRFS ($p=0.0007$ and $p=0.0010$). As MDM2 is an estrogen responsive, prosurvival gene, a subset analysis of ER+ and ER- tumors was done. MDM2 was found to be predictive of IBRFS and LRFS on univariate analysis only in the ER+ subset ($p=0.0003$ and $p=0.0011$ respectively, data not shown). The results were similarly validated in multivariate studies ($p=0.0037$ and $p=0.0037$ respectively). The prognostic value of MDM2 was however not similarly observed in the ER- subset. MDM2 expressors were found to be 9.1 times more likely to experience locoregional recurrence free survival. The multivariate results are displayed in table 4.

Through ubiquitination, MDM2 marks p53 for degradation and hence diminishes its cellular capacity to carry out p53-mediated apoptosis. In vivo and in vitro studies have shown MDM2 to be a key negative regulator of p53 and its apoptotic pathways (Barak, 1993; Kussie, 1996; Lin, 1994; Wu, 1993). While many studies have explored the significance of MDM2 in prostate and breast cancers largely with a focus on overall recurrence, there is a lack of data reporting the significance of MDM2 protein expression on local and regional outcomes following breast radiation (Khor, 2005; Marchetti, 1995; McCann, 1995). In addition, much evidence has recently been shed on the structurally similar protein MDM4, but few have explored its significance in

breast cancer and no studies have explored its significance in relation to local and regional outcomes in early stage breast cancer treated with BCS+RT. Lastly, although p53 has been well studied *in vivo* and *in vitro*, associations between expression and outcomes have not yielded the predicted results. As such we hoped to study the prognostic potential of other protein markers for assessing response to radiation in early stage breast cancer.

In this study, we showed that MDM2 overexpression is associated with significantly worse local recurrence in stage I and stage II invasive breast cancer treated with BCS+RT as defined by ipsilateral breast recurrence free survival (IBRFS) and loco-regional recurrence free survival (LRFS). When examining MDM4 and p53 expression however, we were not able to appreciate a similar prognostic value. Lastly, as MDM2 expression has been linked to active ER α signaling, (Kim, 2011; Marchetti, 1995) we sought to determine if the prognostic significance of MDM2 was associated with this subset. The fact that MDM2 was only found to have predictive value in the ER+ subset may be explained by higher biologic activity of MDM2 in estrogen responsive tumors. Numerous studies have identified associations between MDM2 and ER α expression in breast tissue and breast cancer cell lines [Hori, 2002; Sheikh, 1993; Marchetti, 1995]. *In vitro* data have demonstrated that MDM2 is an estrogen-responsive gene through action of activated ER α on the estrogen response element in the first intron of MDM2 (Hu, 2007; Brekman, 2011; Okumura, 2002; Phelps, 2003). Furthermore, data support two separate interactions between MDM2 and estrogen receptor signaling. Duong *et al.* (Duong, 2007) demonstrate that MDM2 plays a role in ER α turnover through its ubiquitin-ligase activity and targeted ER α degradation and downregulation. In contrast to these findings, Kim *et al.* (Kim, 2011) demonstrated MDM2-enhanced ER α -mediated transactivation in the presence of wildtype p53. Both studies however emphasize protein-protein interactions between MDM2 and ER α leading to these functional responses.

These findings suggest that while both MDM4 and MDM2 are involved in the negative regulation of p53 and subsequent arrest of apoptosis, only MDM2 protein expression may have prognostic value in determining local outcomes in early stage breast cancer treated with BCS+RT. These results add to a growing body of evidence demonstrating that increased expression of MDM2 has negative prognostic value for various endpoints in multiple tumor types (Bueso-Ramos, 1996; Khor, 2005; Kim, 2011; Marchetti, 1995; Marchetti, 1995; McCann, 1995; Turbin, 2006; Lukas, 2001). The prognostic value of MDM2 found to be independent of MDM4 and p53 status of the tumor cores. Additionally, it should be noted that MDM2 was found to be an independent predictor for local outcomes in early stage breast cancer regardless of patients having received chemotherapy or hormone therapy.

Interestingly, there is a correlation between MDM2 expression and Her2 phenotype, *i.e.* higher expression of MDM2 was more common in Her2 overexpressors (Table 1). Nearly an equal number of tumors stained positive for Her2 as were positive for p53. P53 expression, normally low in the absence of cell stress, is thought to increase in the presence of p53 mutation due to resultant stabilization of the dysfunctional protein. P53 mutations are more common in Her2 overexpressing breast tumors. At least one study has identified a relationship between Her2 expression with MDM2 expression (Casalini, 2001). However, in that study, MDM2 is downregulated in the presence of wild type p53. Therefore, the association observed in this

dataset may, in part, reflect the p53-Her2 pathway interaction. This dataset though, does not have sufficient information to validate this hypothesis.

To our knowledge this is the first study assessing the significance of protein expression of MDM2, MDM4 and p53 for local recurrence in conservatively treated, early stage breast cancer. This cohort demonstrated that increased expression of MDM2 correlated with reduced ipsilateral breast recurrence free survival, and worse locoregional relapse free survival in early stage breast cancer treated with breast conserving surgery and radiotherapy. Moreover, on subset analysis, it was found that MDM2 was only found to have prognostic value in the ER + subset alluding to the importance of this protein in ER+ breast cancer. These results add to the growing body of evidence assessing the prognostic value of MDM2 expression, and its potential as a therapeutic target in combination with radiation therapy. If confirmed in larger studies, these results can have significant clinical implications. However, further studies are needed to assess its importance in regional recurrence, and of MDM4 and combinations of other markers in prognosis.

Task 2 Determine the impact of mdm2 SNP309 on the results of adjuvant chemotherapy.

A total of 2453 women have been consented for participation in the parent study protocol as of May 12, 2010 (CINJ Protocol #040406, IRB# 0220044862). Of these, genomic DNA has been isolated from 1,720 patients. The information contained in Table 5 reflects data available from chart review for study participants (this chart review was completed as of February 15, 2010).

The timing of recurrence is an important variable in this dataset since the median follow-up time is 7.2 years. Of 160 recurrences, however, 71% occur by the end of 5 years (Table 6). The majority of recurrences beyond five years reflect estrogen receptor positive disease.

The nature of recurrence reflects the initial stage, molecular features, and type of therapy given adjuvantly. Table 7 depicts the distribution of adjuvant therapies delivered in this cohort of breast cancer patients. The majority of patients received radiation, chemotherapy, and/or hormonal therapy. Only about 12% of patients received trastuzumab.

We will be using this cohort to determine the genotype-specific recurrence free survival for the following: 1) hormone receptor positive and hormone receptor negative breast cancers; 2) hormone receptor positive breast cancer patients receiving hormonal therapy alone; 2) breast cancer patients receiving chemotherapy only (hormone receptor positive and negative disease); 3) breast cancer patients receiving chemotherapy followed by hormonal therapy (hormone receptor positive only).

Breast Cancer Recurrence as a Function of Receptor Status, MDM2 SNP309 Genotype, and Adjuvant Therapy. Of 157 recurrences with known genotypes, more than 50% were in estrogen receptor negative (ER-) breast cancers, as expected. In estrogen receptor negative breast cancer, the recurrence rate was 29% as compared to 16% in estrogen receptor positive (ER+) disease. There is no significant difference in risk of recurrence by genotype for either estrogen receptor positive or estrogen receptor negative breast cancers (Table 8). For ER- disease risk of GG vs. TT genotype, OR 1.132 CI [0.594-2.158], p=0.707. For ER+ disease, OR for recurrence for GG as compared with TT was 1.329 CI [0.837-2.11], p=0.227. Although the frequency of recurrence for GG ER- is 20% and for ER+ is 13%, this is not statistically significant (p=0.41).

Because of the lack of targeted therapy for hormone receptor negative disease, its more aggressive behavior and propensity to recur, the majority of patients with hormone receptor negative disease received chemotherapy (Tables 9, 10). Those patients with ER- disease receiving chemotherapy demonstrate an enrichment of GG genotype in those that recur as compared to other genotypes. This was not significant: OR 1.566 CI [0.608-4.036], $p=0.352$. However, in ER+ patients receiving chemotherapy, heterozygotes are enriched in those recurring but this did not reach statistical significance.

Association of MDM2 SNP309 with Recurrence of Early Stage Breast Cancer

Because stage III disease has the highest risk of recurrence due to its advanced nature, early stage disease was then analyzed separately. This included stage 0 through stage IIB disease. Again, there is an insignificant enrichment of the heterozygotes recurring in those that were ER+ and received hormone therapy. Overall, recurrence rates were similar between ER- disease and ER+ disease by genotype (Table 11). This finding is significant because hormone receptor positive disease has a better prognosis than hormone receptor negative disease in general.

Site-Specific Recurrence as a Function of MDM2 SNP309

We analyzed the site of recurrence for stages 0-III breast cancer as a function of MDM2 genotype. There were few cases where recurrences were regional or multiple sites including local, regional, and distant loci ($n=11$). Therefore, most recurrences were either local ($n=49$) or distant only ($n=49$). While genotype did not associate with risk of local recurrence, G carriers had a higher risk of recurrence: OR 2.188 CI [1.070-4.477], $p=0.028$. Pattern of recurrence in G carriers also favored distant over local recurrences: OR 3.263 CI [1.262-2.456], $p=0.013$. The lack of association with local recurrence rate, but association with distant recurrence confirms the finding in Aim 1.

Combinatorial Analysis of MDM2 SNP309 with MDM4 Genotypes

Because we had previously shown that the variant G allele of MDM2 SNP309 associates with earlier age of diagnosis of ductal breast cancers (1) and more recently demonstrated in the same population that the variant T allele of MDM4 also results in earlier age of diagnosis of ductal breast cancers (2), we asked whether the combination of each risk allele would further modify the age at diagnosis of ductal breast cancers. The combination of the risk genotypes of MDM4 with MDM2 results in the earliest onset of estrogen receptor negative breast cancer. The mean age of diagnosis for MDM4/MDM2 combinations were 41.9 and 50.8 for TT/TG and CC/TG, respectively ($\Delta=8.9$ years; $p=0.0099$). There were insufficient numbers to compare homozygous variants for both MDM4 and MDM2 with the combination wildtype. There was only one TT/GG combination, diagnosed at age 42. In contrast, in estrogen receptor positive breast cancer, the MDM4 risk allele appears to negate the previously-observed earlier onset of the MDM2 SNP309 G allele. For example, when MDM4 was homozygous wildtype, there was a 1.8 year difference in age of onset where the GG combination was diagnosed earlier. When the MDM4 homozygous variant TT was combined with MDM2 SNP309, the age of diagnosis was 54.2 years and 51.9 years for the TT/TT and TT/TG combined genotypes. Although the combined TT/GG variants showed an age of diagnosis of 64 years, there were only 3 cases, underpowering this comparison.

Task 3 Determine the ability of anti-estrogens to restore drug and irradiation sensitivity by decreasing mdm2 expression

In this grant period, we have investigated the effects of anti-estrogen agent, fulvestrant, on mdm2 expression and sensitivity of human breast cancer cells to chemotherapeutic drugs. We found that in both MCF7 (T/G) and T47D (G/G) human breast cancer cell lines, fulvestrant decreases mdm2 expression to similar extents (Figure 3). Further, fulvestrant not only abolished the effect of estradiol (E₂), but also was also able to suppress mdm2 protein levels below the control (no E₂) level (Figure 4). Mdm2 depletion by fulvestrant did not correlate with an increase in p53 activation (slight decrease) and no change in p21 levels was observed (Figure 5). Fulvestrant did not cause a reduction in mdm2 mRNA, but reduces mdm2 protein half-life (Figure 6). The combination of fulvestrant and chemotherapeutic drugs doxorubicin, etoposide or paclitaxel showed synergism in MCF7 and T47D cells (Figure 7).

Epidemiologic evidence suggests that genistein intake is inversely related to the risk of several tumors including breast cancer but its mechanism of action is not completely understood. However, conflicting data exists on the effect of genistein on the expression of the estrogen-dependent mdm2 gene. We hypothesized that if genistein acted like an anti-estrogen, it could bind estrogen receptor (ER), preventing binding to the ERE at the mdm2 promoter and lead to down-regulation of mdm2 expression. For those cells in which SNP309 is present, we anticipated even stronger effects. To explore this, we grew breast cancer cells under conditions of no estrogen (PF), normal media (N), with estradiol (E₂), with Tamoxifen (T), and with genistein (G). We selected three ER+ breast cancer cell lines representing the three MDM2 SNP309 genotypes: ZR75-1 (TT), MCF-7 (TG), and T47D (GG). Protein was isolated from the cells grown in the various conditions and Western blot analysis was performed (Figure 8).

In MCF-7 cells (TG), mdm2 protein is reduced when cells are grown in the absence of estrogen media as compared with normal media or with estradiol. With Tamoxifen or genistein, relative to estradiol, mdm2 was reduced, but remained at levels higher than that in the absence of estrogen. In T47D (GG genotype), the response in the absence of estrogen, normal media, and with estradiol treatment is similar to that of MCF-7 cells (TG genotype). However, by comparison, mdm2 levels are reduced to levels nearly equivalent to those in the absence of estrogen when treated with Tamoxifen and genistein. Of interest, the ~50kDa isoform of mdm2 is reduced further with genistein as compared with Tamoxifen, suggesting an effect on alternative splicing. In ZR75-1 cells (TT), no 50kDa isoform is expressed. In contrast to the MCF7 and T47D cells, genistein and Tamoxifen treatment resulted in **increased** mdm2. Increased expression may be the result of increased transcription or posttranslational changes leading to reduced degradation and longer half-life. These results suggest a genotype-specific effect of genistein and may explain contradictory effects observed in studies.

The P2 promoter of mdm2 has an ERE and we previously demonstrated that mdm2 levels are estradiol dose-dependent and genotype dependent (preliminary data for proposal). Therefore, we had hypothesized that Tamoxifen, an anti-estrogen that binds ER, would result in decreased mdm2 as well as decreased binding at the promoter as determined by chromatin immunoprecipitation (figure 9). While this was true in ZR75-1 cells and to a much lesser degree in MCF7 cells, binding occurred in the presence of Tamoxifen in T47D. As genistein is thought of as an anti-estrogen, we hypothesized that genistein treatment would result in decreased

binding to the ERE. With genistein treatment, ER still bound the P2 promoter region but transcription was reduced in MCF7 and T47D. Interestingly, binding appeared to be reduced in ZR75-1 for treatment with estradiol, Tamoxifen, and genistein. Since protein levels were increased in ZR75-1 with Tamoxifen and genistein, this suggests that post-translational modification leading to longer half-life may play a role in increased mdm2 levels with these treatments. It is not clear if this is truly a genotype-specific effect or if this is related to this particular cell line.

KEY RESEARCH ACCOMPLISHMENTS

- We observed that anti-estrogen agent, fulvestrant, causes a decrease in mdm2 protein half-life, leading to a reduction in mdm2 following treatment with this agent.
- We demonstrate that combined use of fulvestrant with chemotherapeutic drugs doxorubicin, etoposide and paclitaxel can enhance the sensitivity of breast cancer cells to these cytotoxic agents.
- We observed that mdm2 expression is differentially modulated by estrogen, the anti-estrogen tamoxifen, and genistein in a genotype-specific manner. The largest effects on reduction in mdm2 expression at the protein level occur in the mdm2 SNP309 cell line.
- We observed that binding of estrogen receptor alpha to the mdm2 promoter is less efficient in the wildtype mdm2 breast cell line in the presence of estrogen, tamoxifen, and genistein as compared with cell lines carrying at least one variant allele.
- We have accrued the patients needed to evaluate the role of SNP309 in mdm2 on outcomes associated with chemotherapy and hormonal therapy.
- We have analyzed associations between MDM2 SNP309 and breast cancer phenotypes.
- We have observed that mdm2 tissue expression in primary breast tumors correlates with local and locoregional recurrence of breast cancer in women with stage I or stage II tumors undergoing breast conserving surgery and radiation.

REPORTABLE OUTCOMES

Manuscript

Neboori H, Wu H, Kulkarni D, Goyal S, Schiff D, Moran MS, Yang JM, Hirshfield KM, Haffty BG. The Prognostic Value of MDM2 Expression in Early Stage Breast Cancer Treated with Breast Conserving Surgery and Radiotherapy (BCS+RT), Submitted to Cancer 2011.

Abstracts

Nayak M, Hait WN, Hirshfield KM, Haffty B, Yang, JM. A Single Nucleotide Polymorphism in the MDM2 Promoter (SNP 309) Alters the Sensitivity to Topoisomerase II-Targeting Drugs, Era of Hope Meeting 2008, Washington D.C., poster.

Jager A, Hirshfield KM, Hait WN, Haffty B, Yang, JM. The selective estrogen receptor down-regulator, fulvestrant, decreases MDM2 expression and enhances sensitivity of human breast carcinoma cells to chemotherapeutic drugs. AACR 100th Annual Meeting Abstracts 2009, p. 280, poster.

Jäger A, Hait WN, Toppmeyer D, Haffty B, Hirshfield KM, Yang JM. Fulvestrant Decreases MDM2 Expression and Enhances Sensitivity of Human Breast Carcinoma Cells to Chemotherapeutic Drugs. New Jersey Annual Retreat on Cancer Research 2009, poster.

Neboori H, Wu H, Kulkarni D, Goyal S, Schiff D, Moran MS, Yang JM, Hirshfield KM, Haffty BG. The Prognostic Value of MDM2 Expression in Early Stage Breast Cancer Treated with Breast Conserving Surgery and Radiotherapy (BCS+RT), Era of Hope Meeting, Orlando, Florida, August 2011, poster.

Degree obtained that are supported by this award
None

CONCLUSIONS

1. Selective estrogen receptor down-regulator, fulvestrant, decreases MDM2 expression and enhances sensitivity of human breast carcinoma cells to chemotherapeutic drugs (such as doxorubicin, etoposide and paclitaxel).
2. The anti-estrogen tamoxifen decreases MDM2 expression in a genotype-specific manner.
3. MDM2 SNP309 G allele associates with increased risk of distant recurrence of breast cancer.
4. MDM2 SNP309 G allele associates with increased risk of contralateral breast cancer events.
5. Mdm2 tissue expression in primary breast tumors is prognostic for both local and locoregional recurrence of breast cancer in women with stage I or stage II tumors undergoing breast conserving surgery and radiation.

References:

Abdel-Fatah TM, Powe DG, Agboola J, Adamowicz-Brice M, Blamey RW, Lopez-Garcia MA, Green AR, Reis-Filho JS, Ellis IO: The biological, clinical and prognostic implications of p53 transcriptional pathways in breast cancers. J Pathol 2010, 220: 419-434.

Bankfalvi A, Tory K, Kemper M, Breukelmann D, Cubick C, Poremba C, Fuzesi L, Lelle RJ, Bocker W: Clinical relevance of immunohistochemical expression of p53-targeted gene products mdm-2, p21 and bcl-2 in breast carcinoma. Pathol Res Pract 2000, 196: 489-501.

Barak Y, Juven T, Haffner R, Oren M: mdm2 expression is induced by wild type p53 activity. *EMBO J* 1993, 12: 461-468.

Bond G, Hirshfield KM, Kirchhoff T, Alexe G, Bond EE, Robins H, Bartel F, Taubert H, Wuerl P, Hait W, Toppmeyer D, Offit K, and Levine A. MDM2 SNP309 accelerates tumor formation in a gender-specific and hormone-dependent manner. *Cancer Research*, 2006; 66: 5104-5110.

Bond GL, Hu W, Bond EE, Robins H, Lutzker SG, Arva NC, Bargonetti J, Bartel F, Taubert H, Wuerl P, Onel K, Yip L, Hwang SJ, Strong LC, Lozano G, Levine AJ. A single nucleotide polymorphism in the MDM2 promoter attenuates the p53 tumor suppressor pathway and accelerates tumor formation in humans. *Cell* 2005, 119:591-602.

Brekman A, Singh KE, Polotskaia A, Kundu N, Bargonetti J. A p53-independent role of Mdm2 in estrogen-mediated activation of breast cancer cell proliferation. *Breast Cancer Res*. 2011;13(1): R3.

Bueso-Ramos CE, Manshouri T, Haidar MA, Yang Y, McCown P, Ordonez N, Glassman A, Sneige N, Albitar M: Abnormal expression of MDM-2 in breast carcinomas. *Breast Cancer Res Treat* 1996, 37: 179-188.

Casalini P, Botta L, Menard S. Role of p53 in HER2-induced proliferation or apoptosis. *J Biol Chem*. 2001; 276: 12449-53.

Danovi D, Meulmeester E, Pasini D, Migliorini D, Capra M, Frenk R, de Graaf P, Francoz S, Gasparini P, Gobbi A et al: Amplification of Mdmx (or Mdm4) directly contributes to tumor formation by inhibiting p53 tumor suppressor activity. *Mol Cell Biol* 2004, 24: 5835-5843.

Duong V, Boulle N, Daujat S, Chauvet J, Bonnet S, Neel H, Cavailles V. Differential regulation of estrogen receptor alpha turnover and transactivation by Mdm2 and stress-inducing agents. *Cancer Res*. 2007; 67: 5513-21.

Freedman DA, Wu L, Levine AJ: Functions of the MDM2 oncoprotein. *Cell Mol Life Sci* 1999, 55: 96-107.

Hori M, Shimazaki J, Inagawa S, Itabashi M: Overexpression of MDM2 oncoprotein correlates with possession of estrogen receptor alpha and lack of MDM2 mRNA splice variants in human breast cancer. *Breast Cancer Res Treat* 2002, 71: 77-83.

Hu W, Feng Z, Ma L, Wagner J, Rice JJ, Stolovitzky G, Levine AJ. A single nucleotide polymorphism in the MDM2 gene disrupts the oscillation of p53 and MDM2 levels in cells. *Cancer Res*. 2007; 67: 2757-65.

Khor LY, Desilvio M, Al-Saleem T, Hammond ME, Grignon DJ, Sause W, Pilepich M, Okunieff P, Sandler H, Pollack A: MDM2 as a predictor of prostate carcinoma outcome: an analysis of Radiation Therapy Oncology Group Protocol 8610. *Cancer* 2005, 104: 962-967.

Kim K, Burghardt R, Barhoumi R, Lee SO, Liu X, Safe S: MDM2 regulates estrogen receptor {alpha} and estrogen responsiveness in breast cancer cells. *J Mol Endocrinol* 2011, 46: 67-79.

Kulkarni DA, Vazquez A, Haffty BG, Bandera EV, Hu W, Sun YY, Toppmeyer DL, Levine AJ, Hirshfield KM. A polymorphic variant in human MDM4 associates with accelerated age of onset of estrogen receptor negative breast cancer. *Carcinogenesis* 2009, 30: 1910-5.

Kussie PH, Gorina S, Marechal V, Elenbaas B, Moreau J, Levine AJ, Pavletich NP: Structure of the MDM2 oncoprotein bound to the p53 tumor suppressor transactivation domain. *Science* 1996, 274: 948-953.

Lin J, Chen J, Elenbaas B, Levine AJ: Several hydrophobic amino acids in the p53 amino-terminal domain are required for transcriptional activation, binding to mdm-2 and the adenovirus 5 E1B 55-kD protein. *Genes Dev* 1994, 8: 1235-1246.

Lukas J, Gao DQ, Keshmeshian M, Wen WH, Tsao-Wei D, Rosenberg S, Press MF. Alternative and aberrant messenger RNA splicing of the mdm2 oncogene in invasive breast cancer. *Cancer Res.* 2001; 61: 3212-9.

Marchetti A, Buttitta F, Girlando S, Dalla Palma P, Pellegrini S, Fina P, Doglioni C, Bevilacqua G, Barbareschi M: mdm2 gene alterations and mdm2 protein expression in breast carcinomas. *J Pathol* 1995, 175: 31-38.

Marchetti A, Buttitta F, Pellegrini S, Merlo G, Chella A, Angeletti CA, Bevilacqua G: mdm2 gene amplification and overexpression in non-small cell lung carcinomas with accumulation of the p53 protein in the absence of p53 gene mutations. *Diagn Mol Pathol* 1995, 4: 93-97.

Marechal V, Elenbaas B, Piette J, Nicolas JC, Levine AJ: The ribosomal L5 protein is associated with mdm-2 and mdm-2-p53 complexes. *Mol Cell Biol* 1994, 14: 7414-7420.

McCann AH, Kirley A, Carney DN, Corbally N, Magee HM, Keating G, Dervan PA: Amplification of the MDM2 gene in human breast cancer and its association with MDM2 and p53 protein status. *Br J Cancer* 1995, 71: 981-985.

Okamoto K, Kashima K, Pereg Y, Ishida M, Yamazaki S, Nota A, Teunisse A, Migliorini D, Kitabayashi I, Marine JC et al: DNA damage-induced phosphorylation of MdmX at serine 367 activates p53 by targeting MdmX for Mdm2-dependent degradation. *Mol Cell Biol* 2005, 25: 9608-9620.

Okumura N, Saji S, Eguchi H, Nakashima S, Saji S, Hayashi S. Distinct promoter usage of mdm2 gene in human breast cancer. *Oncol Rep.* 2002; 9: 557-63.

Phelps M, Darley M, Primrose JN, Blaydes JP. p53-independent activation of the hdm2-P2 promoter through multiple transcription factor response elements results in elevated hdm2 expression in estrogen receptor alpha-positive breast cancer cells. *Cancer Res.* 2003; 63: 2616-2.

Roth J, Dobbelstein M, Freedman DA, Shenk T, Levine AJ: Nucleo-cytoplasmic shuttling of the hdm2 oncoprotein regulates the levels of the p53 protein via a pathway used by the human immunodeficiency virus rev protein. *EMBO J* 1998, 17: 554-564.

Sheikh MS, Shao ZM, Hussain A, Fontana JA. The p53-binding protein MDM2 gene is differentially expressed in human breast carcinoma. *Cancer Res.* 1993; 53:3226-8.

Shvarts A, Bazuine M, Dekker P, Ramos YF, Steegenga WT, Merckx G, van Ham RC, van der Houven van Oordt W, van der Eb AJ, Jochemsen AG: Isolation and identification of the human homolog of a new p53-binding protein, Mdmx. *Genomics* 1997, 43: 34-42.

Shvarts A, Steegenga WT, Riteco N, van Laar T, Dekker P, Bazuine M, van Ham RC, van der Houven van Oordt W, Hateboer G, van der Eb AJ et al: MDMX: a novel p53-binding protein with some functional properties of MDM2. *EMBO J* 1996, 15: 5349-5357.

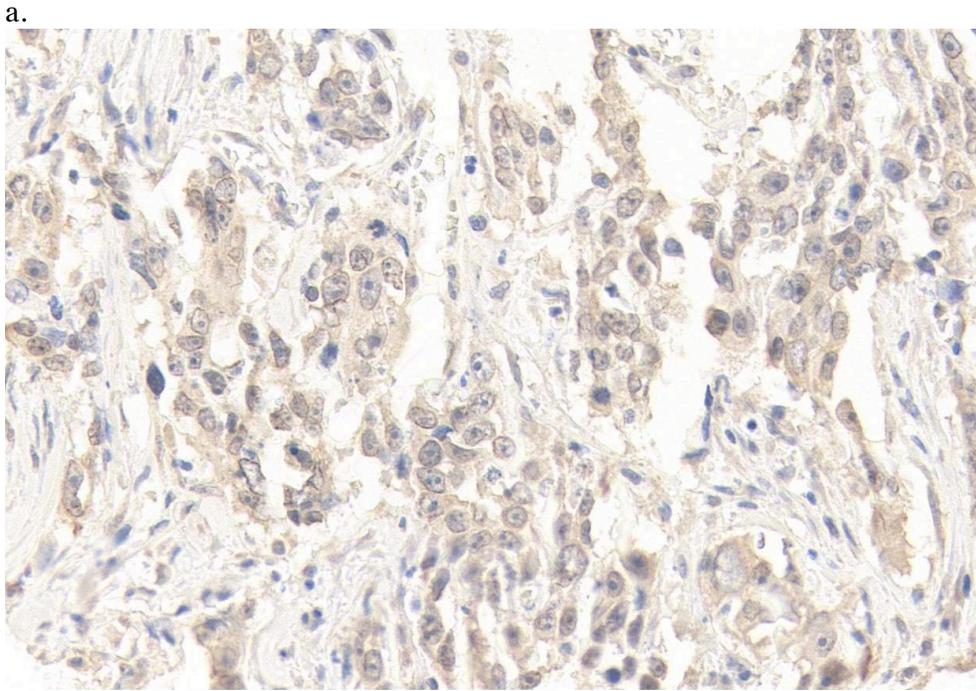
Turbin DA, Cheang MC, Bajdik CD, Gelmon KA, Yorida E, De Luca A, Nielsen TO, Huntsman DG, Gilks CB: MDM2 protein expression is a negative prognostic marker in breast carcinoma. *Mod Pathol* 2006, 19: 69-74.

Wu X, Bayle JH, Olson D, Levine AJ: The p53-mdm-2 autoregulatory feedback loop. *Genes Dev* 1993, 7: 1126-1132.

APPENDICES: none

SUPPORTING DATA

Figure1: Representative immunohistochemical staining of MDM2 in breast tumors (40x): a. MDM2 positive; b. MDM2 negative



b.

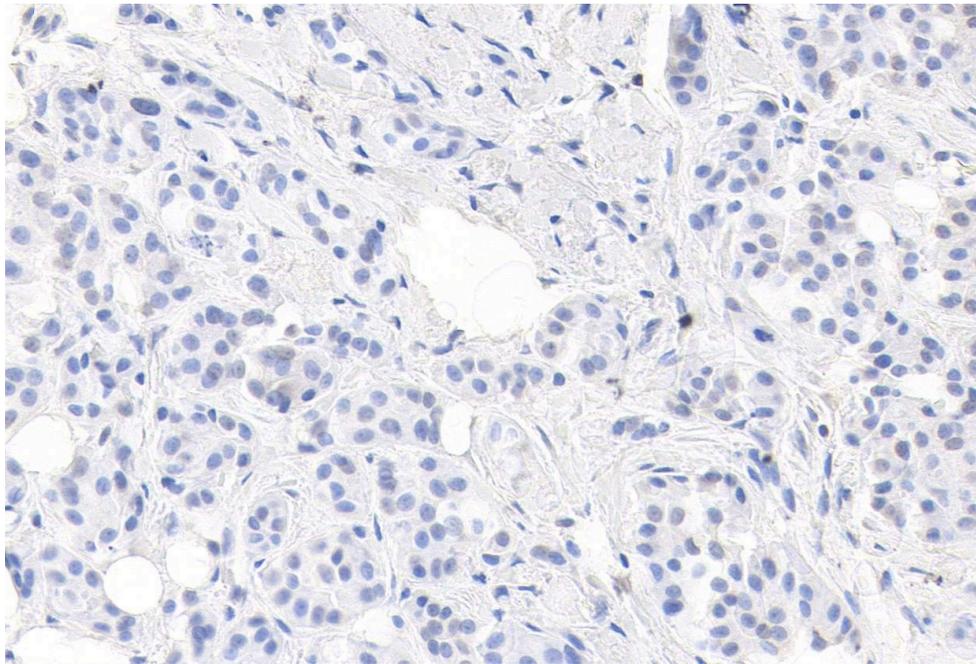
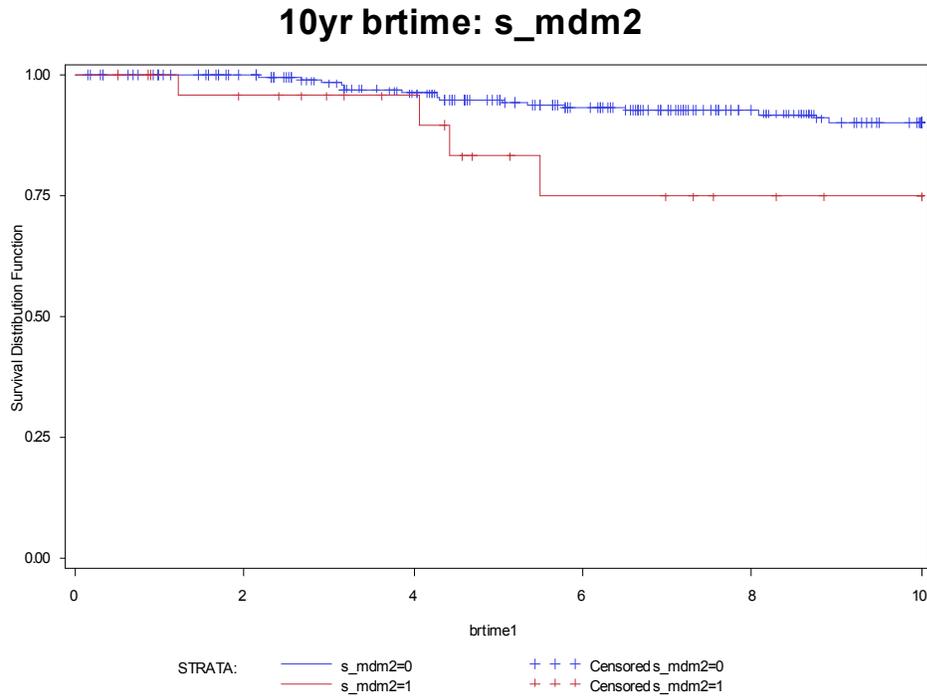


Figure2: Kaplan Meier survival curves of MDM2+ (red) v. MDM2- (blue) for: a. Ipsilateral breast recurrence b. Locoregional recurrence

a.



b.

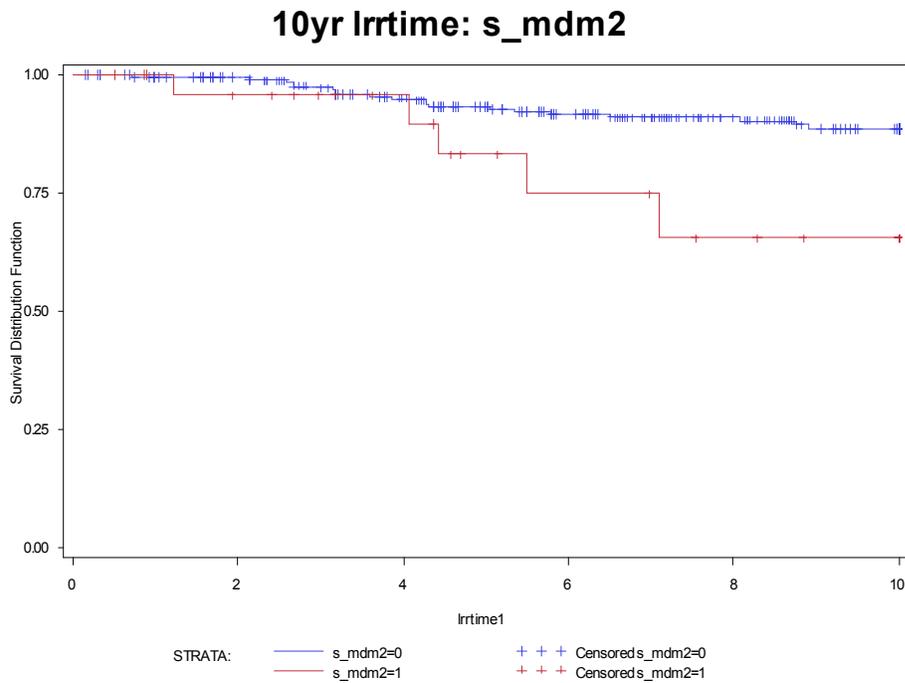


Table 1. Correlation of clinical and pathologic factors with MDM2 expression

Feature	MDM2 expression			p
	Total (%)	Negative (%)	Positive (%)	
Age (y)				0.90
≤ 50	120(40)	110(40)	10(38)	
> 50	183(60)	167(60)	16(62)	
Race				0.67
White	241(80)	221(80)	20(77)	
Black	49(16)	45(16)	4(15)	
Other	13(4)	11(4)	2(7)	
ER				0.58
Negative	108(37)	100(38)	8(32)	
Positive	183(63)	166(64)	17(68)	
PR				0.91
Negative	211(71)	193(71)	18(72)	
Positive	86(29)	79(29)	7(28)	
HER2				<0.01
Negative	250(86)	234(88)	16(67)	
Positive	40(14)	32(12)	8(33)	
Triple Negative Status				0.12
ER-/PR-/Her2-	72(25)	69(26)	3(12)	
≥ 1+ Marker	220(75)	198(74)	22(88)	
Adjuvant hormones				0.51
Not Received	162(54)	147(53)	15(60)	
Received	140(46)	130(47)	10(40)	
Adjuvant chemotherapy				0.37
Not Received	194(64)	180(65)	14(56)	
Received	108(36)	97(35)	11(44)	
Pathologic N stage				0.75
0	180(77)	164(77)	16(76)	
1	49(21)	44(21)	5(24)	
2	5(2)	5(2)	0	
Tumor size				0.38
T1(<2cm)	183(64)	170(67)	13(52)	
T2 (2-5cm)	101(36)	89(33)	12(48)	
Margin status				0.59
Positive	23(8)	21(8)	2(8)	
Negative	218(72)	197(71)	21(80)	
Unknown	62(20)	59(21)	3(12)	

* The totals for each marker are different as all clinicopathologic data was not available for every patient.

Table 2: Univariate analysis of prognostic factors and 10 year outcomes

Prognostic Factor	IBRFS HR (95% CI)	p-value	NRFS HR (95% CI)	p-value	LRFS HR (95% CI)	p-value
MDM2 positivity	3.077 (1.044 - 9.073)	0.0416	2.059 (0.247 - 17.134)	0.5042	3.084 (1.169 - 8.134)	0.0229
Age > 50 years	0.678 (0.375 - 1.226)	0.1988	0.616 (0.178 - 2.128)	0.4438	0.638 (0.368 - 1.105)	0.1086
Race	1.318 (0.780 - 2.230)	0.3025	1.500 (0.531 - 4.232)	0.4439	1.337 (0.824 - 2.171)	0.2393
ER status	0.853 (0.447 - 1.629)	0.6302	0.415 (0.099 - 1.739)	0.2292	0.797 (0.434 - 1.460)	0.4619
PR status	0.664 (0.292 - 1.512)	0.3295	N/A	0.9941	0.561 (0.249 - 1.264)	0.1632
HER2 status	0.716 (0.220 - 2.329)	0.5784	1.158 (0.142 - 9.415)	0.8907	0.847 (0.302 - 2.373)	0.7521
Systemic Therapy*	0.656 (0.363 - 1.18)	0.1631	0.850 (0.240 - 3.016)	0.8014	0.663 (0.382 - 1.151)	0.1440
Triple negative status	0.881 (0.431 - 1.800)	0.7285	0.385 (0.096 - 1.542)	0.1777	0.836 (0.431 - 1.621)	0.5954
Nodal Status	1.161 (0.463 - 2.910)	0.7496	5.280 (0.881 - 31.652)	0.0686	1.403 (0.621 - 3.170)	0.4154
Tumor Size	3.175 (1.738 - 5.802)	0.0002	2.538 (0.720 - 8.952)	0.1474	3.027 (1.727 - 5.303)	0.0001
Margin	3.174 (1.267 - 7.950)	0.0137	2.400 (0.268 - 21.487)	0.4337	2.607 (1.061 - 6.403)	0.0367

* Systemic Therapy is defined as receiving hormonal therapy, chemotherapy or both.

Table 3. Multivariate analysis of prognostic factors and 10 year outcomes

Prognostic Factor	IBRFS HR (95% CI)	p-value	LRFS HR (95% CI)	p-value
Tumor size	9.310 (2.570 - 33.725)	0.0007	5.829 (2.039 - 16.665)	0.0010
Margin status	1.099 (0.569 - 2.123)	0.7784	1.174 (0.663 - 2.079)	0.5822
Nodal Status	0.694 (0.175 - 2.762)	0.6048	1.029 (0.327 - 3.235)	0.9613
MDM2 expression	11.235 (2.710 - 46.577)	0.0009	9.274 (2.794 - 30.783)	0.0003

Abbreviations: IBRFS = Ipsilateral breast recurrence survival. LRFS = Locoregional recurrence-free survival.

Table 4. 10 year Multivariate analysis of MDM2 in ER+ and ER- subsets

Prognostic Factor	ER+ (n=183)				ER- (n=108)			
	IBRFS HR (95% CI)	p-value	LRFS HR (95% CI)	p-value	IBRFS HR (95% CI)	p-value	LRFS HR (95% CI)	p-value
Tumor size	1.676 (0.759 - 3.700)	0.2014	1.676 (0.759 - 3.700)	0.2014	1.228 (0.660 - 2.286)	0.5172	1.190 (0.706 - 2.006)	0.5141
Margin status	1.725 (0.270 - 11.032)	0.5649	1.725 (0.270 - 11.032)	0.5649	0.850 (0.051 - 14.072)	0.9093	0.783 (0.054 - 11.337)	0.8575
Nodal Status	0.861 (0.614 - 1.207)	0.3855	0.861 (0.614 - 1.207)	0.3855	1.326 (0.917 - 1.916)	0.1339	1.262 (0.904 - 1.761)	0.1716
MDM2 expression	9.144 (2.051 - 40.769)	0.0037	9.144 (2.051 - 40.769)	0.0037	0.000 (0.000 - N/A)	0.9955	1.978 (0.226 - 17.272)	0.5374

Abbreviations: IBRFS = Ipsilateral breast recurrence survival. NRFS = Nodal recurrence free survival.

LRFS = Locoregional recurrence free survival.

* Systemic Therapy is defined as receipt of hormonal therapy, chemotherapy or both.

Table 5- Demographics of Study Cohort at The Cancer Institute of New Jersey.

Race	Number of Patients	% of Patients
African American	57	5.7
Asian	41	4.1
Caucasian	771	77.3
Hispanic	61	6.1
Indian	25	2.5
Other	43	4.3

Tumor Type	Number of Patients	% of Patients
Colloid/Mucinous	12	1.3
DCIS	93	9.8
Invasive Ductal	705	74.5
Invasive Lobular	94	9.9
Medullary	6	0.6
Metaplastic	4	0.4
Other	32	3.4
Unknown	52	n/a

ER Status	Number of Patients	% of Patients
Positive	748	74.9
Negative	250	25.1

PR Status	Number of Patients	% of Patients
Positive	638	63.9
Negative	361	36.2

Her2/Neu Status	Number of Patients	% of Patients
Not amplified or 0-2+ IHC	553	79.9
Amplified or 3+ IHC (all 2+ by IHC were reflexed for FISH)	145	20.1

Stage	Number of Patients	% of patients
0	93	9.3
1	345	34.6
IIA	198	12.5
IIB	125	12.5
IIIA	70	7.0
IIIB	25	2.5
IIIC	20	2.0
IV	40	4
Unknown	156	15.6

Tumor	% of Patients
T0	5.2
T1	46.3
T2	24.7
T3	6.5
T4	4.6

Node status	
N0	47.6
N1	32.5
N2	3.7
N3	0.2

Metastatic Status	
M0	86
M1	4

Recurrence Status	% of patients
Yes	20.3
No	79.7

(excludes stage IV at diagnosis)

Table 6. Time to recurrence of breast cancer from date of initial biopsy-proved disease.

Year(s) to recurrence	n	% of all recurrences
1	13	0.081
2	39	0.243
3	23	0.144
4	19	0.119
5	21	0.131
6	10	0.063
7	5	0.031
8-10	8	0.050
>10	22	0.138

Table 7. Distribution of the adjuvant therapy received by breast cancer patients in this cohort.

Patients Receiving Each Treatment	No (%)	Yes (%)
Radiation	22.8	77.2
Chemotherapy	32.7	67.3
Hormonal therapy	27.7	72.3
Trastuzumab	87.7	12.3

Table 8. Rates of breast cancer recurrence as a function of hormone receptor status and use of adjuvant hormone therapy.				
	ER-/no hormone therapy		ER+/hormone therapy	
	No recurrence	Recurrence	No Recurrence	Recurrence
TT	50 (0.35)	19 (0.32)	190 (0.38)	31 (0.32)
TG	70 (0.49)	28 (0.47)	224 (0.45)	54 (0.55)
GG	23 (0.16)	12 (0.20)	85 (0.17)	13 (0.13)

Table 9. Rates of ER- breast cancer recurrence by MDM2 SNP309 genotype in those receiving adjuvant chemotherapy.				
	ER-/no chemo		ER-/chemo	
	No recurrence	Recurrence	No Recurrence	Recurrence
TT	8	1	41 (0.36)	16 (0.30)
TG	2	0	54 (0.48)	26 (0.49)
GG	0	0	18 (0.16)	11 (0.21)

Table 10. Rates of ER+ breast cancer recurrence by MDM2 SNP309 genotype in those receiving adjuvant chemotherapy.				
	ER+/no chemo		ER+/chemo	
	No recurrence	Recurrence	No Recurrence	Recurrence
TT	70 (0.38)	7	88 (0.37)	20 (0.29)
TG	84 (0.46)	5	106 (0.45)	40 (0.58)
GG	30 (0.16)	3	44 (0.18)	9 (0.13)

Table 11. Rate of breast cancer recurrence in ER+ and ER- disease by MDM2 SNP309 genotype and use of adjuvant hormone therapy.				
	ER-/no hormone therapy		ER+/hormone therapy	
	No recurrence	Recurrence	No Recurrence	Recurrence
TT	25 (0.31)	11 (0.37)	116 (0.36)	20 (0.29)
TG	42 (0.53)	15 (0.50)	148 (0.46)	37 (0.54)
GG	13 (0.16)	4 (0.13)	58 (0.18)	11 (0.16)

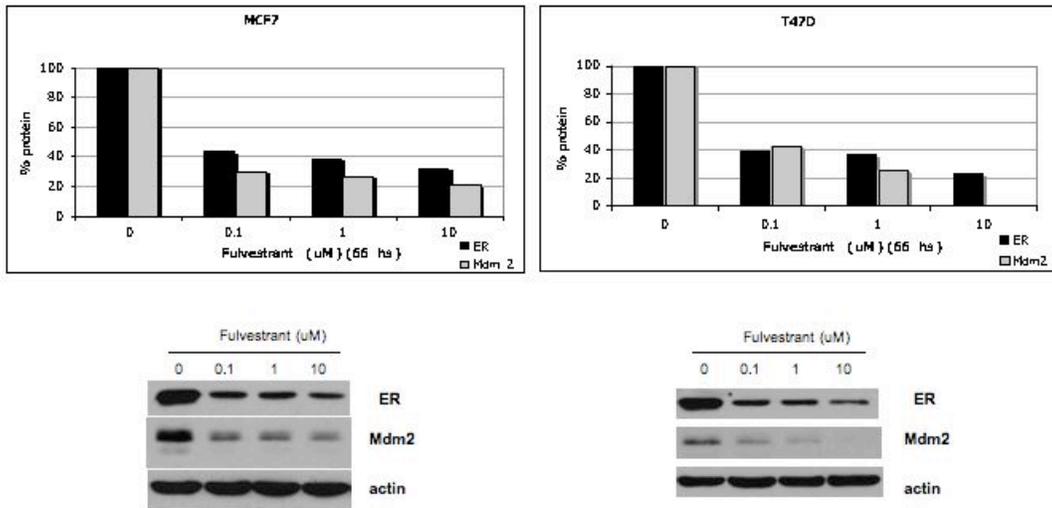


Figure 3. Effect of the antiestrogen fulvestrant on expression of estrogen receptor and mdm2 proteins. Two breast cancer cell lines MCF7 and T47D were grown at various concentrations (0-10 micromolar) of fulvestrant for 66 hours. Protein was then harvested and levels of estrogen receptor and mdm2 were assayed by Western blot. The upper plots demonstrate the dose-dependent reduction of both proteins in each cell line.

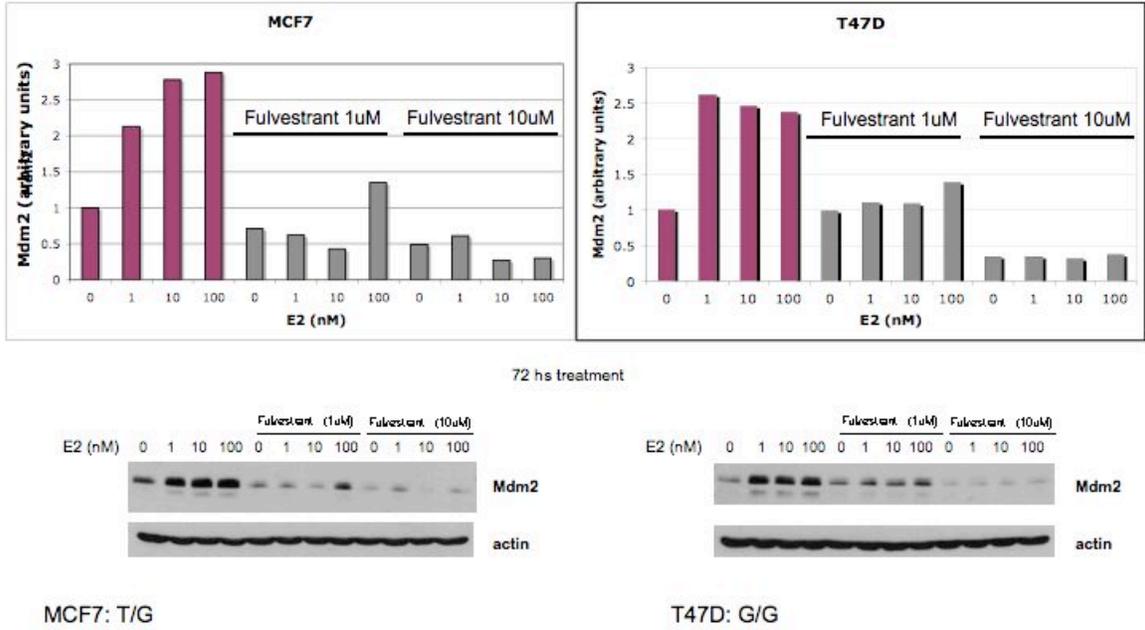


Figure 4. Effect of the antiestrogen fulvestrant on mdm2 levels in breast cancer cells grown in the presence of estradiol. Two breast cancer cell lines MCF7 and T47D were grown in the presence of estradiol, and estradiol with one of two concentrations of fulvestrant. The lower plots represent the Western blot analysis corresponding to the quantification in the upper graphs.

MCF7: T/G; p53 wt

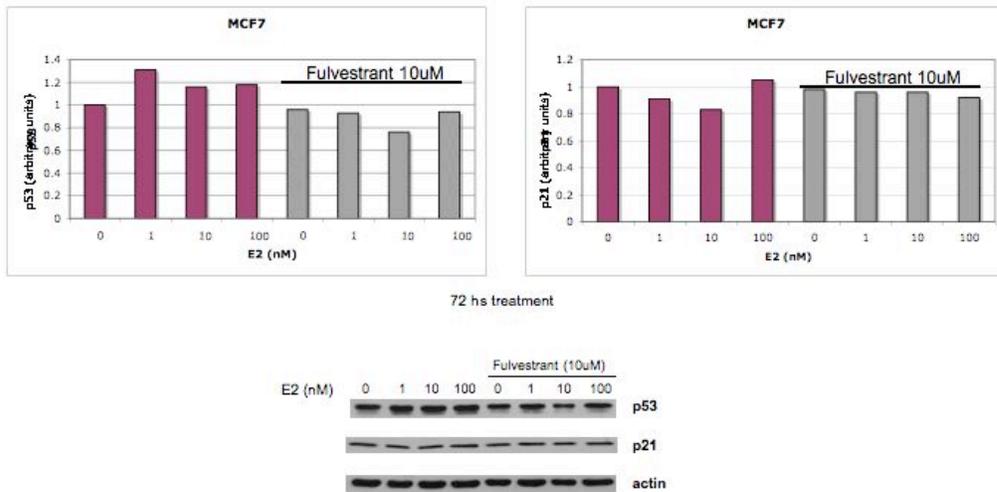


Figure 5. Effect of estradiol and the antiestrogen fulvestrant on p53 and p21 in breast cancer cell lines. The breast cancer cell lines MCF7 was grown in estradiol alone or with the presence of 10micromolar fulvestrant. Protein was harvested and Western blot analysis performed to detect p53 and p21. The lower plot depicts the Western blot for each protein using actin as a loading control. This plot was used to quantitate protein levels expressed in the upper curves.

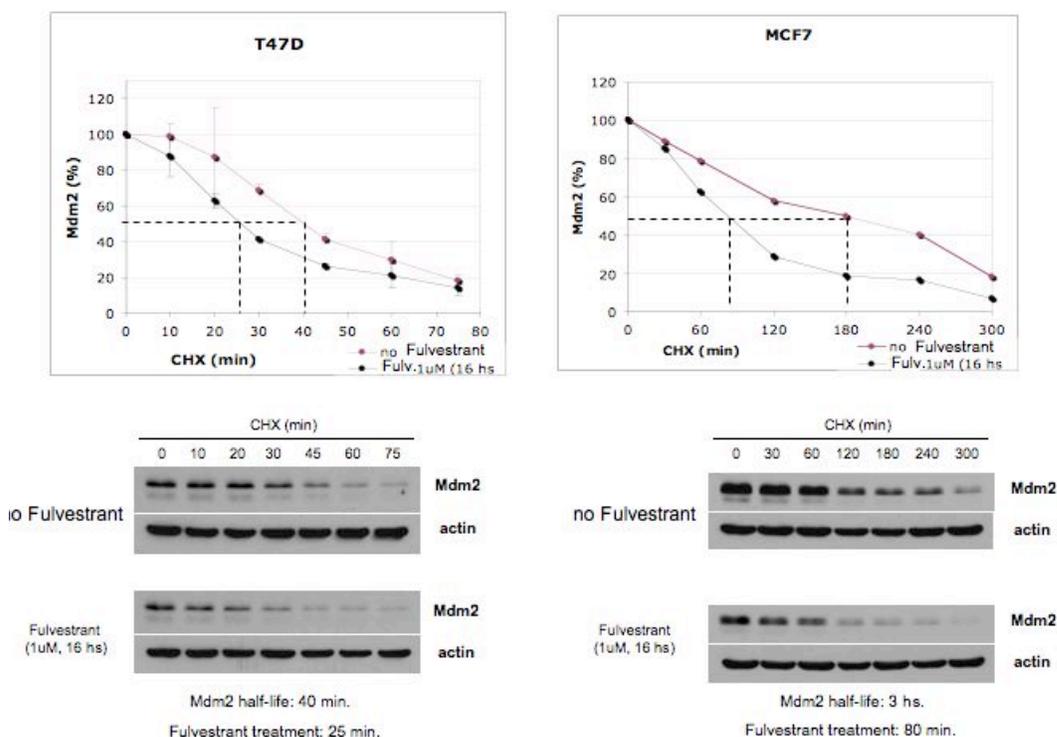


Figure 6. Effect of fulvestrant on the half-life of mdm2 protein. Two breast cancer cell lines T47D and MCF7 were grown in the absence and the presence of the antiestrogen fulvestrant. Cells were treated with cycloheximide (CHX) and mdm2 protein expression was determined at various time points. The lower curves show Western Blot analyses from each cell type using actin as a loading control and were used to quantitate mdm2 levels given in the corresponding curves above.

		<u>CompuSyn Analysis:</u>	
MCF7	Fulvestrant : Doxorubicin at constant ratio	1 : 0.5	Synergism
		1 : 0.015	Synergism
	Fulvestrant : Paclitaxel at constant ratio	1 : 0.025	Synergism / Additive
T47D	Fulvestrant : Etoposide at constant ratio	1 : 0.0005	Synergism
		1 : 1	Synergism
		1 : 5	Synergism
T47D	Fulvestrant : Doxorubicin at constant ratio	1 : 0.25	Synergism / Additive
		1 : 0.0035	Synergism / Additive
	Fulvestrant : Paclitaxel at constant ratio	1 : 0.0125	Synergism / Antagonism
T47D	Fulvestrant : Etoposide at constant ratio	1 : 0.0002	Synergism / Antagonism
		1 : 10	Synergism / Additive
		1 : 0.07	Synergism / Additive / Antagonism

Figure 7. Effect of combining the antiestrogen fulvestrant with doxorubicin, paclitaxel, or etoposide in two breast cancer cell lines. Analysis of cell response was determined using the CompuSyn program. Each combination was observed at two concentrations of chemotherapeutic agent while keeping the concentration of fulvestrant constant. The last column indicates the type observed effect of the combination for each drug and dose.

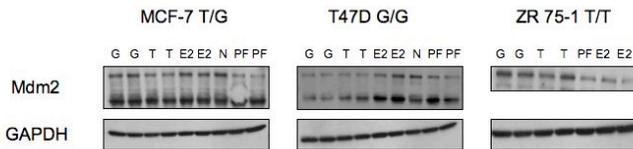


Figure 8. Western blot demonstrates mdm2 protein expression in three ER+ breast cancer cells lines representing the three SNP309 genotypes: ZR75-1 (TT), T47D (GG), MCF7 (TG). Cells were grown under different conditions: phenol-free, charcoal stripped media (PF), normal media (N), estradiol (E2), Tamoxifen (T), or genistein (G).

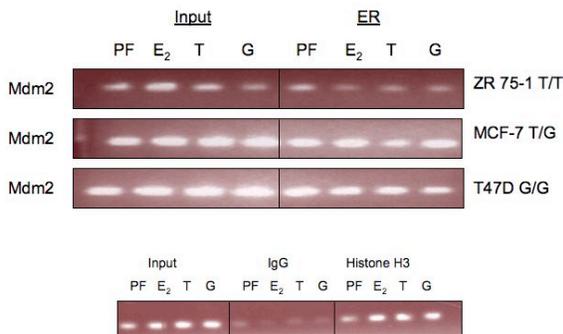


Figure 9. Chromatin immunoprecipitation using anti-ERalpha antibody with PCR of the mdm2 P2 promoter region was performed in the three ER+ breast cancer cell lines representing each of the three MDM2 genotypes.