Award Number: W81XWH-06-1-0292

TITLE: Role of Obesity in Prostate Cancer Development

PRINCIPAL INVESTIGATOR: Margot P. Cleary, Ph.D.

CONTRACTING ORGANIZATION: University of Minnesota Hormel Institute
Austin, MN 55912

REPORT DATE: March 2009

TYPE OF REPORT: Final

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.
Role of Obesity in Prostate Cancer Development

Obesity was induced by injections with gold-thioglucose (GTG) at either 6, 16 or 26 weeks of age. Body weight was monitored and the mice are followed until 46 weeks of age. Unfortunately there was an unexpectedly high mortality rate in TRAMP mice in response to the GTG injections. This resulted in a limited number of mice available to follow. However, the general results indicated little effect of obesity on prostate cancer in this model. A second experiment was conducted using a prostate cancer cell line developed from a TRAMP mouse tumor, TRAMP-C2. Since this cell line was developed from a mouse on the C57BL6 background it can be inoculated into wild-type mice and tumor formation can be monitored. Mice were fed a high fat diet and then divided by body weight into Obesity-Prone, Overweight and Obesity-Resistant groups with an additional group fed a low-fat diet. Mice were inoculated with the cell line and tumor growth monitored. There was some indication that the high-fat diet per se affected tumor weight and size. Interestingly, genital-urinary and prostate weights were highest in the Obesity-Prone mice. Results of these two studies are being integrated with an additional study of dietary-induced obesity in TRAMP mice (funded by another agency) to complete assessment of the effect of body weight and/or a high-fat diet on prostate cancer in this animal model.
<table>
<thead>
<tr>
<th>Section</th>
<th>Page</th>
</tr>
</thead>
<tbody>
<tr>
<td>Introduction</td>
<td>4</td>
</tr>
<tr>
<td>Body</td>
<td>4</td>
</tr>
<tr>
<td>Key Research Accomplishments</td>
<td>16</td>
</tr>
<tr>
<td>Reportable Outcomes</td>
<td>16</td>
</tr>
<tr>
<td>Conclusion</td>
<td>17</td>
</tr>
<tr>
<td>References</td>
<td>18</td>
</tr>
<tr>
<td>Appendices</td>
<td>21</td>
</tr>
</tbody>
</table>
INTRODUCTION:

A number of epidemiological studies indicate that increased body weight plays a role in the development of prostate cancer [1-9]. Although, not all studies have found obesity to be associated with increased risk of prostate cancer, Bergström et al concluded that based on obtained relative risk values 5,000 new cases of prostate cancer per year in Europe could be attributed to obesity [10]. In addition, mortality from prostate cancer is increased with elevated body weights [11], and obesity was recently reported to be associated with higher prostate cancer grade at diagnosis, as well as with higher recurrence rates [12]. The potential role of body weight in the development and progression of prostate cancer is of interest given that the incidence of overweight/obesity is increasing throughout the world, and the potential for lifestyle changes to alter body weight status. Interestingly, since we originally submitted this proposal an increasing number of publications have addressed the issue of obesity and its association with the development and prognosis of prostate cancer [13-17].

The initial goal of the present proposal was to evaluate the effects of obesity initiated at different ages on prostate cancer development using the TRAMP mouse model. Obesity was to be induced at different ages (6, 16 and 26 weeks of age) using gold-thioglucose (GTG) injections. Results of that study will be presented. However, there were a number of problems with this protocol primarily a very high mortality rate of TRAMP mice receiving GTG which was not anticipated based on the literature as well as preliminary studies we conducted using wild-type C57BL6 mice. Due to this aspect of the study was suspended and a different approach was implemented using a cell line developed from a TRAMP mouse tumor, TRAMP-C2. These cells were implanted into mice with different body weights following consumption of a high fat diet. Results of this study will also be presented.

BODY:

We previously used diet-induced obesity to determine effects of body weight on the development of mammary tumors in several transgenic mouse strains [18;19]. However due to the younger age and aggressive nature of the disease in the TRAMP mice as well as the variable response of mice to this intervention, i.e., a range of body weights obtained, we decided to use an alternative approach to increase body weight and to determine the effect of obesity initiated at specific ages on prostate cancer. Our rationale was that due to the short time for development of prostate cancer in TRAMP mice and the transition of the disease through different stages, a more uniformly developing obesity model would make it easier to interpret results and fewer experimental animals would be needed. An alternative to diet-induced obesity is to damage the hypothalamus [20]. This can be done by physical/mechanical means (knife cuts and electrical lesions) or less invasively by chemical damage. Specifically, a single injection with GTG results in the majority of the mice gaining weight and becoming obese [21]. It is unclear why not all the treated animals respond, because when they are re-injected they then develop obesity. This is an important observation as it indicates that the initial lack of response is not due to resistance to GTG. Shortly after leptin was identified, GTG-induced obesity was reported to increase plasma leptin as identified by immunoblot; and leptin mRNA expression in adipose tissue was elevated compared to lean animals [22]. More recently when leptin levels were assessed by commercially available radioimmunoassay kits, GTG obese mice were found to have serum leptin levels two-fold higher than in control mice [23;24]. Body weight gain without consumption of a high-fat diet is obtained, although food intake is initially increased in GTG-treated mice [23;25]. Body weights eventually
plateau and caloric intakes are appropriate for body weights. There is no age-sensitive time-point at which GTG needs to be administered in order to produce the effect on body weight. For example, reported results include mice injected as young as 3-4 wks of age [26-29] In other studies mice were anywhere from 8 to 20 weeks of age [21;23;25;30;31]. Also different GTG doses over a range from 0.3-2.0 mg/g body weight have been used. In our own study in female nude mice we found that a dose of 0.5 mg/g resulted in a high mortality rate therefore we decided to undertake a preliminary study in male mice prior to injecting the TRAMP mice with GTG.

**Pilot study 1:**

Gold thioglucose (GTG) was injected into 14 male wild-type mice at a dose of 0.5 mg/g body weight. A control group of six male wild-type mice was injected with the same volume of PBS. One mouse became ill after receiving GTG and euthanization was necessary. Mice ranged in age from 8 to 23 weeks and were followed for ten weeks to monitor body weight changes and general body condition. Fifty-four percent (7 out of 13) of the mice became obese. The GTG obese cohort gained significantly more weight than either the GTG non-obese or the control mice (Figure 1). There was no significant difference in weight gain between the non-obese and the PBS injected mice. In comparison to our earlier study in female mice the male mice tolerated the 0.5 dose well.

![Pilot study #1](image)

**Pilot study 2:**

Since the 0.5 dose was well tolerated and in an effort to increase the percentage of mice that become obese a dose of 0.8 mg/g body weight of GTG was injected into 12 male wild-type mice. A control group was made up of five male wild-type mice that were injected with PBS. Four mice became ill after receiving GTG and euthanization was necessary. The mice, ranging in age from 11 to 13 weeks, were then followed for 10 weeks to monitor body weight changes and general body condition. Sixty-three percent (5 out of 8) became obese. The GTG obese cohort gained significantly more weight than either the GTG non-obese or the PBS-injected mice (Figure 2). There was no significant difference in the 10-week weight gain between the non-obese and the control mice.

**Figure 1:** 10-week weight gain during pilot study 1. ANOVA P = 0.001; GTG obese versus GTG lean P < 0.01; GTG obese versus PBS P < 0.01; GTG lean versus PBS P > 0.05.

**Figure 2:** 10-week weight gain during pilot study 2. ANOVA P = 0.001; GTG obese versus GTG lean P < 0.01; GTG obese versus PBS P < 0.01; GTG lean versus PBS P > 0.05.
Study of TRAMP mice injected with GTG at three different ages

**General Methods:** All mice had *ad libitum* access to purified AIN-93M diet and water. Following the pilot studies we determined that the 0.8 dose would be used and we started injection. Body weights were recorded weekly and at that time mice were palpated for tumors. Mice that received GTG were categorized as obese or non-obese, based on weight gain relative to the PBS control mice. Serum samples were collected from the retro-orbital sinus at baseline and every 5 weeks until a tumor was palpated. Following tumor palpation, serum was collected every 3 weeks until study termination. Data are presented as mean ± SE.

**Survival:** To our surprise the TRAMP mice did not tolerate the GTG as well as anticipated and several adjustments were made to the protocol in attempts to improve survival. The dose of 0.8 mg/g was best tolerated in the mice injected at 26 weeks of age; this cohort had the highest survival at 42%. Mice in the 6- and 16-week cohorts had a much lower rate of survival at this dose, 0 and 13%, respectively. Lowering the dose to 0.5 mg/g increased survival at 6-weeks of age to 23%, but only 9% survived at this dose when injected at 16 weeks of age. A summary of the survival data is presented in Table 1.

<table>
<thead>
<tr>
<th>Table 1. Percent Surviving after GTG Injection</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>0.8 mg/g</strong></td>
</tr>
<tr>
<td>6-week</td>
</tr>
<tr>
<td>16-week</td>
</tr>
<tr>
<td>26-week</td>
</tr>
</tbody>
</table>

**Obesity rate, weight gain and fat pad weights:** After receiving GTG injections, as expected, some mice rapidly gained weight, GTG-obese, while others did not, GTG-lean. The percent that became obese was identical for the 6- and 26-week cohort, 57%. In the 16-week cohort, only 33% became obese. Body weight curves for the three cohorts are shown in Figure 3.
The GTG-lean mice and PBS control mice had similar body weights over the course of the study. The actual weight gains are provided in Figure 4 where it can be seen that the GTG-lean mice gained a similar amount of weight as did control mice given saline injections only. As expected GTG-obese mice gained significantly more weight than did the two lean groups. Figure 5 presents fat pad weights which were similar in the GTG-lean and control mice which were significantly lower compared to the GTG obese mice in each age cohort.
**Figure 4.** Average weight gain of TRAMP mice in the 6 week (Panel A), 16 week (Panel B), and 26 week (Panel C) cohorts. For both the 6 and 26 week cohorts ANOVA p<0.0001 and columns with different superscripts significantly different from each other. For the 16 week cohort t test between the two lean groups was not significantly.

**Figure 5.** Average fat pad weights (sum of epididymal and retroperitoneal) for TRAMP mice in the 6 week (Panel A), 16 week (Panel B) and 26 week (Panel C) cohorts. For 6 and 26 week cohorts ANOVA p<0.001, columns with different superscripts are significantly different. For 16 week cohort t test not significant between the two lean groups.
### Table 2. End point comparisons for TRAMP male mice

<table>
<thead>
<tr>
<th></th>
<th>GTG-6</th>
<th></th>
<th></th>
<th></th>
<th>Tumor differentiation</th>
<th>Percent with metastasis</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Final body weight (g)</td>
<td>Age at tumor palpation (weeks)</td>
<td>Age at death (weeks)</td>
<td>GU weight (g)</td>
<td>Well</td>
<td>Moderate</td>
</tr>
<tr>
<td>Obese (N=4)</td>
<td>51.78 ± 5.30&lt;sup&gt;a&lt;/sup&gt;</td>
<td>27.67 ± 4.91 (N=3)</td>
<td>35.75 ± 4.59</td>
<td>6.23 ± 1.40</td>
<td>50%</td>
<td>25%</td>
</tr>
<tr>
<td>Lean (N=3)</td>
<td>36.07 ± 3.75&lt;sup&gt;b&lt;/sup&gt;</td>
<td>22.0 ± 3.51</td>
<td>28.33 ± 8.84</td>
<td>6.30 ± 1.96</td>
<td>33%</td>
<td>0%</td>
</tr>
<tr>
<td>PBS (N=15)</td>
<td>36.92 ± 1.33&lt;sup&gt;b&lt;/sup&gt;</td>
<td>29.36 ± 1.61 (n=14)</td>
<td>38.33 ± 1.92</td>
<td>7.84 ± 0.84</td>
<td>60%</td>
<td>7%</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>GTG-16</th>
<th></th>
<th></th>
<th></th>
<th>Tumor differentiation</th>
<th>Percent with metastasis</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Final body weight (g)</td>
<td>Age at tumor palpation (weeks)</td>
<td>Age at death (weeks)</td>
<td>GU weight (g)</td>
<td>Well</td>
<td>Moderate</td>
</tr>
<tr>
<td>Obese (N=1)</td>
<td>51.9</td>
<td>30</td>
<td>40</td>
<td>9.42</td>
<td>100%</td>
<td>0%</td>
</tr>
<tr>
<td>Lean (N=2)</td>
<td>33.2 ± 1.0</td>
<td>25 (N=1)</td>
<td>24 ± 4</td>
<td>3.87 ± 2.4</td>
<td>100%</td>
<td>100%</td>
</tr>
<tr>
<td>PBS (N=13)</td>
<td>35.5 ± 1.7</td>
<td>26.5 ± 1.4 (N=12)</td>
<td>35.6 ± 2.5</td>
<td>7.3 ± 0.8</td>
<td>54%</td>
<td>8%</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>GTG-26</th>
<th></th>
<th></th>
<th></th>
<th>Tumor differentiation</th>
<th>Percent with metastasis</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Final body weight (g)</td>
<td>Age at tumor palpation (weeks)</td>
<td>Age at death (weeks)</td>
<td>GU weight (g)</td>
<td>Well</td>
<td>Moderate</td>
</tr>
<tr>
<td>Obese (N=8)</td>
<td>48.61 ± 2.60&lt;sup&gt;a&lt;/sup&gt;</td>
<td>33.38 ± 0.94</td>
<td>42.13 ± 1.22</td>
<td>9.10 ± 1.54 (N=6)</td>
<td>50%</td>
<td>50%</td>
</tr>
<tr>
<td>*Lean (N=6)</td>
<td>39.08 ± 2.46&lt;sup&gt;b&lt;/sup&gt;</td>
<td>33.5 ± 1.09</td>
<td>43.0 ± 1.67</td>
<td>8.90 ± 1.45 (N=5)</td>
<td>67%</td>
<td>17%</td>
</tr>
<tr>
<td>PBS (N=12)</td>
<td>36.65 ± 1.63&lt;sup&gt;b&lt;/sup&gt;</td>
<td>32.3 ± 1.1</td>
<td>41.3 ± 1.4</td>
<td>8.16 ± 1.0 (N=11)</td>
<td>58%</td>
<td>8%</td>
</tr>
</tbody>
</table>

<sup>a</sup> columns with different letters indicate a significant difference among the groups
<sup>b</sup> pathology report not received for one mouse in this group
Prostate cancer results: A summary of the results for the cohorts is presented in Table 2. In all three cohorts final body weights were significantly higher for the GTG-obese mice compared to the GTG-lean and PBS control mice. For the other determinations there were no significant differences partially attributable to the small sample sizes in GTG-obese and GTG-lean groups. However, a few interesting observations were made. For example, the GTG-obese mice injected at 6 weeks of age had a delay in tumor detection compared to the GTG-lean mice and a delayed age at death. Genital-urinary tract (GUT) weight was not impacted by body weight. Tumor differentiation was improved and metastases rate was reduced in GTG-obese mice compared to the PBS control mice. Due to the poor survival rate for the mice injected at 16 weeks of age it was not possible to make conclusions for this group. For the mice injected at 26 weeks of age, age of tumor detection, age at death and GUT weights were similar in all three groups. There was however, a trend for GTG-obese mice to have an improved tumor differentiation profile compared to both lean groups and to have a reduced metastasis rate compared to the control saline injected mice. As indicated above serum samples were obtained from the mice. We attempted to evaluate the samples for testosterone and estradiol levels in relationship to obesity status and in relationship to age of tumor detection. However, in general these results did not provide any consistent findings.

Figure 6. Expression of Acrp30 and leptin receptors from TRAMP prostate tumor tissue. Immunohistochemistry of A) AdipoR1, B) AdipoR2, C) Ob-Rb and D) control with goat serum instead of primary antibody. The presence of reddish brown color indicates a positive reaction for the specific primary antibodies. Arrows indicate areas of positive staining for each antibody. Hematoxylin was utilized for the blue counter staining of the nuclei. E) Westerns of AdipoR1, AdipoR2 and Ob-R from TRAMP prostate tumor tissues from four different TRAMP animals. The individual animal numbers at the top of the blots. The antibodies used are shown along the left hand side.

Expression of AdipoR1, AdipoR2 and Ob-Rb in tumor tissue from mouse prostate cancer: Because of the increasing interest in body weight and how it might affect prostate cancer and due to the findings of in vitro studies with leptin and adiponectin [32-36] which are both produced in adipose tissue we investigated whether tumors from TRAMP mice express the two receptors for Acrp30, AdipoR1 and AdipoR2 and the signaling form of the leptin receptor, Ob-Rb. Tumors from four different mice were examined using immunohistochemistry and western analysis. Representative staining from the immunohistochemistry is shown in Figure 6. Expression of AdipoR1 was found in all prostate tumor tissue from TRAMP mice and was primarily located in epithelial cells on the apical region (Fig 6A). In samples from all mice AdipoR2 (Fig 6B) was present primarily in the same areas as AdipoR1 but the staining was less intense. Ob-Rb (Fig 6C) was expressed throughout the samples. Control staining with goat serum instead of primary antibody was negative (Fig 6D). Western blot analysis of frozen tissue from the same mice was also performed and expression of AdipoR1, AdipoR2 and leptin receptor was found in all prostate tumor tissues examined (Fig 6E).
Study using TRAMP-C2 Cells

Because of the difficulties with GTG injections described above we decided to take a different approach to evaluate the effect of obesity on prostate tumor development. We utilized a diet-induced obesity regimen for our studies which we had used successfully in studying mammary tumor development in transgenic mice as well as in a xenograft study [18;19;37;38]. C57BL6 mice are particularly susceptible to become obese when fed a high-fat diet. Furthermore, use of this strain provides the unique opportunity to evaluate prostate cancer cell ability to develop tumors in mice fed the same diet but with different body weights. Previous studies in our lab and in other labs have shown that although most C57BL6 mice fed a high-fat diet will gain weight and become overweight or obese, some will stay in the body weight range of low-fat fed mice [18;39]. This occurrence provides the opportunity to compare mice of the same body weight consuming diets of different composition as well as to compare mice fed the same diet but with different body weights. C57BL6 mice have a normal immune system and may be implanted with the syngeneic Tramp C2 tumor cells which will cause tumors to develop [40;41]. Using this model we investigated the influence of a high fat diet on TRAMP-C2 cell growth in vivo. We also determined the effects of weight by comparing obesity resistant vs obesity prone mice for TRAMP-C2 tumor growth. The use of this cell line provides a straightforward approach which is much simpler than using athymic mice in xenograft studies of human prostate cancer cells as the mice do not need to be maintained in an ultraclean environment. Also it is fortuitous to be able to use C57BL6 mice as these mice readily develop dietary-induced obesity.

*In vitro study:* Prior to the mouse study we did some in vitro studies to evaluate the effects of the addition of leptin and adiponectin to the TRAMP-C2 cells. These results are presented in Figure 7.

**Figure 7.** Proliferation of TRAMP-C2 cells 48 hours after treatment with Acrp30, leptin or both. Cell proliferation as a percent is shown along the y-axis. Cells in serum-free media were considered to be 100%. A) The concentrations of Acrp30 are shown along the x-axis. Bars represent standard error of the mean from the four different experiments and asterisks are significantly different from untreated. (ANOVA p=0.0393). The concentrations of Acrp30 and leptin are shown along the x-axis. Bars represent standard error of the mean from three different experiments. B) Cell proliferation of the TRAMP-C2 cells in response to increasing levels of leptin (ng/ml). The concentration of leptin is shown along the x-axis. Bars represent standard error of the mean from three different experiments.
It can be seen that when the cells were treated with Acrp30 in the physiological range of 2.5-20 μg/ml there was a dose-related reduction in proliferation of the TRAMP-C2 cells after 48 hours (Fig 7A). The difference in proliferation was statistically significant at the Acrp30 concentration of 20 μg/ml compared to the untreated cells. The cells had 70% proliferation compared to the untreated controls. In addition, cells were treated with a combination of Acrp30 and leptin. We found that the addition of leptin blocked the ability of Acrp30 to inhibit proliferation. However, figure 7B shows that treatment of the cells with leptin alone in its physiological range for 48 hours did not result in a statistically significant change in cell proliferation. This data strengthens the possibility that prostate cancer proliferation can be inhibited by Acrp30 and that leptin can block this protective effect. This would provide an explanation for how obesity could impact prostate cancer cell development and/or progression because in the obese state the high levels of serum leptin and low levels of adiponectin would be permissive for cell proliferation.

**In vivo experiments with TRAMP-C2 cells:** We obtained C57BL6 male mice (n=160) from Jackson Laboratory, Bar Harbor ME in groups of 40. From weaning at 4 weeks of age mice were maintained on AIN-93M diet [38;42]. At 6 weeks of age 120 mice were switched AIN-93M-High-Fat diet [38]. At 20 weeks of age mice were implanted with the TRAMP-C2 cells (3x10⁶) in the left flank. Mice were weighed and palpated for tumors over the next 10 weeks. At the termination of the study a blood sample was obtained and the tumors were removed, measured and weighed and then processed for histopathology. Epididymal and retroperitoneal fat pads were removed as well as genitourinary tracts and prostates.

**Weight gain and visceral fat pad weights in low-fat fed and high-fat fed mice:** Figure 8A shows the weights of the 4 different groups of mice at 25 weeks of age (this age was used to avoid potential problems as the tumors grew). As expected, the mice fed a high-fat diet could be divided into three weight groupings, obesity resistant, overweight and obesity-prone [18]. Five of the high-fat fed mice had to be removed from the study due to non-study related illnesses. All 40 of the low-fat fed mice finished the study. The ANOVA for the mouse weights was P<0.0001. All groups were significantly different from each other (P<0.001). The low-fat fed mice weighed on average 35.4 grams, the high-fat fed obesity resistant mice averaged 33.8 grams which was significantly different from the low-fat fed mice. The high-fat fed overweight mice averaged 37.6 grams and the high-fat fed obesity-prone mice weighed an average of 41.1 grams. We also weighed the visceral fat pads from the mice (Fig 8B) when they were sacrificed. Similar results were found as compared to the body weights. The ANOVA for the visceral fat pads was P<0.0001. The lightest visceral fat pads were from the low-fat fed mice (1.78 grams) and the high-fat fed obesity resistant mice (1.73 grams) and these were not significantly different from each other. The visceral fat pads of the high-fat fed overweight mice averaged 2.16 grams and the visceral fat pads from the high-fat fed obesity-prone mice weighed an average of 2.49 grams and were significantly different from each other.

![Figure 8](image-url). **A.** Body weights at 25 weeks of age and **B.** Fat pad weights at termination. Low-fat fed (LF), high-fat fed obesity-resistant (HF Ob-Res), high-fat fed overweight (HF OverWt) and high-fat fed obesity-prone (HF Ob-Pr). Bars represent standard error and letters show significance differences.
Genitourinary tract and prostate cancer results: We found that the genitourinary tracts from the high-fat fed obesity-resistant mice were lowest (Table 3) and that the genitourinary tract weights from the high-fat fed obesity prone mice were highest. The low-fat fed mice and high-fat fed overweight mice did not have significantly different genitourinary tract weights. Because the TRAMP-C2 cells were injected subcutaneously in the flank we were able to harvest normal prostates from the mice to examine the effects of weight and a high fat diet on normal prostates. We found that the high-fat fed obesity-prone mice had significantly heavier prostates as compared to Low-Fat and Obesity-Resistant groups (Table 3). The prostate weight was also higher in Obesity-Prone compared to Overweight mice but the difference was not significantly different.

Table 3. Genital-Urinary Tract and Prostate Weights in Male C57BL6 Mice with Diet-Induced Obesity (mean ± sem)

<table>
<thead>
<tr>
<th></th>
<th>Low-Fat</th>
<th>Obesity-Resistant</th>
<th>Overweight</th>
<th>Obesity-Prone</th>
</tr>
</thead>
<tbody>
<tr>
<td>GU-Tract Weight (g)</td>
<td>0.51&lt;sup&gt;b&lt;/sup&gt; ± 0.01</td>
<td>0.47&lt;sup&gt;c&lt;/sup&gt; ± 0.01</td>
<td>0.51&lt;sup&gt;b&lt;/sup&gt; ± 0.01</td>
<td>0.55&lt;sup&gt;a&lt;/sup&gt; ± 0.01</td>
</tr>
<tr>
<td>ANOVA p&lt;0.0001</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Prostate Weight (g)</td>
<td>0.0668&lt;sup&gt;b&lt;/sup&gt; ± 0.0021 (n=19)</td>
<td>0.0624&lt;sup&gt;b&lt;/sup&gt; ± 0.0045 (n=20)</td>
<td>0.0721&lt;sup&gt;a,b&lt;/sup&gt; ± 0.0029 (n=19)</td>
<td>0.08126&lt;sup&gt;a&lt;/sup&gt; ± 0.0048 (n=20)</td>
</tr>
<tr>
<td>ANOVA p=0.005</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Columns with different superscripts are significantly different from each other.

We examined the TRAMP-C2 tumors harvested from the mice and compared the different groups based on weight and volume. Figure 9A shows that the TRAMP-C2 tumors from the low-fat fed and high-fat fed obesity-prone mice were the lightest. The tumors from the high-fat fed obesity-resistant and the high-fat fed overweight mice were heavier. However, the differences were not significant. When the tumor volumes were computed the low-fat fed mice had the smallest tumors followed by the high-fat fed obesity-prone mice with the high-fat fed obesity-resistant and overweight mice having the largest tumors by volume but there was not a significant difference. When mice were divided into low-fat fed verses high-fat fed mice we found that the high-fat fed mice had heavier tumors than the low-fat fed mice (Fig 10A) (P<0.01). We also found that the average tumor volume of the high-fat fed mice was higher as compared to the low-fat fed mice (Fig 9B) (P<0.0007).
Figure 8A. Tumor weight at sacrifice. The four groups of mice are; low-fat fed (LF), high-fat fed obesity-resistant (HF Ob-Res), high-fat fed overweight (HF OverWt) and high-fat fed obesity-prone (HF Ob-Pr). Bar represent standard error. ANOVA = NS

Figure 8B. Tumor volume at sacrifice. The four groups of mice are; low-fat fed (LF), high-fat fed obesity-resistant (HF Ob-Res), high-fat fed overweight (HF OverWt) and high-fat fed obesity-prone (HF Ob-Pr). Bar represent standard error. ANOVA = NS

Figure 9A. Tumor weights at sacrifice. The average weight of the tumors from the low-fat fed mice (LF) is shown as red. The average weight of tumors from all of the high-fat fed mice (HF) is shown as blue. Bars represent standard error and letters show significance differences. P<0.01

Figure 9B. Tumor volumes at sacrifice. The average tumor volume of the low-fat fed mice (LF) is shown as red. The average tumor volume of all of the high-fat fed mice (HF) is shown as blue. Bars represent standard error and letters show significance differences. P<0.0007
Overall these findings do not produce a clear picture of the effect of body weight on prostate tumor development. There seems to be some indication that a high fat does impact tumor TRAMP-C2 growth. Interestingly higher body weight was associated with an increase in GU-tract weight as well as prostate weight. How this would be related to prostate cancer development remains unclear. We are presently analyzing data from a separate study funded by a different agency whereby the diet-induced protocol was utilized in TRAMP mice. Interestingly we found little effect of body weight on age of tumor development and death but there did appear to be an increase in poorly differentiated tumors 19% in the Obesity-Prone TRAMP mice compared to all other groups. The Overweight and Obesity-Resistant mice had 6% and 8% respectively while there were none in the Low-Fat group. This finding suggests both a body weight and diet effect on tumor grade. Our task now is to integrate these three different studies and determine how the findings relate to tumor development in these models and assess whether results help to explain the involvement of obesity in men.
KEY RESEARCH ACCOMPLISHMENTS:
1) Performed experiments as described to induce obesity at specific ages.
2) Preliminary interpretation indicates that obesity at a young age may be protective with respect to the development of prostate cancer. This is consistent with some human epidemiological studies.
3) Found that toxicity and mortality associated with GTG-induced obesity makes it impractical for continued use.
4) Determined that AdipoR1, AdipoR2 and Ob-Rb are expressed by TRAMP prostate cancer cells.
5) Measured serum estradiol and testosterone levels at various times over the course of the experiment but results were not consistent with any relationship to when tumors were detected.
6) We used an alternative approach to address the issue of the effect of body weight on prostate cancer development by feeding high fat diets to induce obesity. Body weights were affected by the diet.
7) Mice with obesity had higher genital urinary tract weights as well as prostate weights. Although the high fat fed mice tended as a group to have higher tumor weights than the low-fat mice this was primarily attributable to the obesity-resistant and overweight groups.
8) Performed in vitro experiments that indicate that TRAMP-C2 cell proliferation is inhibited by Acrp30 and that this effect is blocked by high levels of leptin.

REPORTABLE OUTCOMES:

ROLE OF OBESITY AT DIFFERENT AGES IN PROSTATE CANCER DEVELOPMENT IN TRAMP MICE. Margot P. Cleary, Melissa J.L. Bonorden, Olga P. Rogozina and Nancy K. Mizuno
Presented at the IMPACT meeting September 2007, Atlanta, GA. (abstract included Appendix A)

CHARACTERIZATION OF ADIPONECTIN RECEPTOR EXPRESSION AND FUNCTION IN TRAMP PROSTATE TUMORS AND THE TRAMP-C2 CELL LINE. Michael E. Grossmann, Nancy K. Mizuno, Melissa J. L. Bonorden, Amitabha Ray and Margot P. Cleary
Presented at the Frontiers in Cancer Prevention Research meeting December 2007, Philadelphia, PA (abstract included Appendix B)

IMPACT OF TWO TYPES OF OBESITY ON PROSTATE CANCER IN THE TRAMP MOUSE
Melissa J.L. Bonorden, Michael E. Grossmann, Olga P. Rogozina, D.Joshua Liao, Joseph P. Grande and Margot P. Cleary
To be presented at the AACR meeting April 2009 in Denver, CO (abstract included Appendix C)

ROLE OF THE ADIPONECTIN LEPTIN RATIO IN PROSTATE CANCER
Michael E. Grossmann, Nancy K. Mizuno, Melissa J.L. Bonorden, Amitabha Ray, Irina Sokolchik, Meena L. Narasimhan and Margot P. Cleary
Submitted (abstract included Appendix D)

COMPARISON OF THE EFFECT OF TWO TYPES OF OBESITY ON PROSTATE CANCER DEVELOPMENT IN TRAMP MICE
Melissa J.L. Bonorden, Michael E. Grossmann, Olga P. Rogozina, D.Joshua Liao, Joseph P. Grande and Margot P. Cleary
In preparation

EFFECT OF DIETARY INDUCED OBESITY ON TUMOR FORMATION FROM TRAMP-C2 CELLS
Michael E. Grossmann, Katai J. Nkhata, Nancy K. Mizuno and Margot P. Cleary
In preparation.
CONCLUSIONS:
This has been a frustrating experience over the first two years trying to undertake what we thought was a straightforward approach to inducing obesity in the mice. At this time we hope that our new approach will provide more insightful results into the effect of obesity on prostate cancer.
Reference List


ROLE OF OBESITY AT DIFFERENT AGES IN PROSTATE CANCER DEVELOPMENT IN TRAMP MICE
Margot P. Cleary, Melissa J.L. Bonorden, Olga P. Rogozina, & Nancy K. Mizuno

A number of epidemiological studies have implicated obesity as a risk factor for prostate cancer development. In addition, clinical and biochemical progression of prostate cancer has been reported to be shorter in obese men and mortality from prostate cancer is increased with elevated body weights[11]. Obesity was recently reported to be associated with higher prostate cancer grade at diagnosis, as well as with higher recurrence rates. The potential role of body weight in various aspects of prostate tumorigenesis is of interest given that the incidence of overweight/obesity is increasing throughout the world, and the potential for lifestyle changes to alter body weight status. The goal of the present study is to determine the effect of obesity induced at different ages on the development of prostate cancer using the TRAMP mouse model. Development of prostate cancer in the TRAMP mouse shares a number of similarities with the human disease. The experimental design was to induce obesity at three different ages and follow prostate cancer development. To attain this goal male TRAMP mice (C57BL6 background) were injected with gold-thiogucose (GTG) (0.5-0.8 mg/kg body weight in phosphate buffered saline (PBS)) at 6, 16 and 26 weeks of age. Mice were weighed weekly and palpated to detect prostate tumors. Mice were followed until 46 weeks of age or until disease burden necessitated euthanasia. Serial blood samples over the course of the study are also obtained. Control mice received injections of only the PBS vehicle. For the 26-week cohort 14 mice survived the GTG injection, of which 8 were obese (48.6 ± 2.6 grams). As expected some of the GTG mice did not develop obesity and were designated as non-Obese. Their body weight 39.1 ± 2.5 was in the body weight range of the PBS mice (36.7 ± 1.63). The final body weights for the Obese mice were significantly higher than for the other two groups (ANOVA p <0.05). Fat pad weights followed a similar relationship. Total genital-urinary tract weights were not affected by body weight. Age of prostate tumor detection was not different among the three groups of mice (~33 weeks of age in age). Additionally age at death (~43 weeks of age) was similar among the groups. Serum and tissue analyses are presently being conducted. Additional cohorts of mice injected with GTG at 6 and 16 weeks of age are currently being followed. Overall it appears that GTG has a high toxicity and mortality rate in TRAMP mice (in contrast to our preliminary studies in C57BL6) mice. In the older mice induction of obesity had little effect on the development of prostate cancer. Ongoing studies will address the consequence of obesity on prostate cancer development in younger TRAMP mice.

Support: DOD PC050284 and the Hormel Foundation.
Appendix B

Characterization of Adiponectin Receptor Expression and Function in TRAMP Prostate Tumors and the TRAMP-C2 Cell Line

Michael E. Grossmann, Nancy K. Mizuno, Melissa J. L. Bonorden, Amitabha Ray, and Margot P. Cleary
Hormel Institute, University of Minnesota, Austin, MN
Presented at the AACR Frontiers in Cancer Prevention Meeting- Philadelphia PA December 2007

Introduction: Obesity is associated with increased risk for more aggressive prostate cancer (Pca) as defined by an increase in the risk of Pca death and an increased chance of progression after surgery. Obesity may mediate its effects on Pca in part due to factors secreted from adipose tissue. One factor potentially involved in the interaction between Pca and obesity is adiponectin, also known as adipocyte complement-related protein of 30 kDa (Acrp30). Lower serum Acrp30 levels have been reported for Pca patients compared to patients with benign prostatic hyperplasia or healthy controls. In addition, lower expression levels of Acrp30 receptors are found in prostate tumors as compared to healthy prostate tissue. Here we assessed how Acrp30 impacted cell growth in vitro in the TRAMP-C2 cell line which is derived from a TRAMP prostate tumor and determined Acrp30 receptor expression in the TRAMP model.

Procedures: TRAMP-C2 cells (ATCC) were used in growth assays (CC8 kit Dojindo Laboratories). Whole cell extracts were obtained using Phosphosafe extraction reagent from Novagen for determination of adiponectin receptors (AdipoR1 and R2) and signaling proteins by western blot. Antibodies were from Santa Cruz Biotechnology except antibodies to AdipoR1 (Abcam Inc), AdipoR2 (Phoenix Pharmaceuticals, Inc.) and anti-rabbit secondary (Cell Signaling Inc.).TRAMP mice were euthanized at 50 weeks and urogenital tracts plus abnormal growths/tumors removed. Sections were stained with the rabbit ABC staining system for AdipoR1 and R2.

Results: There was a dose-related reduction in proliferation of the TRAMP-C2 cells after 48 hours in response to the addition of Acrp30. The difference in proliferation was statistically significant at physiological Acrp30 concentrations of 10 and 20 ug/ml (Student’s t-test p<0.03 and 0.02 respectively) compared to untreated cells. Western blots indicated that AdipoR1 and AdipoR2 are both expressed by TRAMP-C2 cells. We also identified increases or decreases in phosphorylation of several growth associated signaling proteins with western blots. Acrp30 increased levels for both ERK1 and ERK2. The phosphorylation of Stat3 was decreased by the addition of fetal calf serum but this decrease was blocked by Acrp30.

We also found that tumors from TRAMP mice expressed the two receptors for Acrp30, AdipoR1 and AdipoR2. Using immunohistochemical analysis we found expression of AdipoR1 in prostate tumor tissue from TRAMP mice was mostly in epithelial cells on the apical membrane. AdipoR2 was present in the same areas as AdipoR1 but the staining was lower. Western blot analysis of frozen tissue from the same mice also indicated expression of AdipoR1 and AdipoR2 in prostate tumor tissue.

Conclusions: Here, we are the first to report the presence of AdipoR1 and AdipoR2 in prostate tumor tissues from TRAMP mice and in the TRAMP-C2 cell line which is derived from the prostate tumor of a TRAMP mouse. The receptors appear to be functional since proliferation of TRAMP-C2 cells was inhibited by addition of Acrp30. This decrease in cell growth may be attributable to increased signaling through ERK 1/2 since Acrp30 increased the phosphorylation of ERK 1/2. We are currently investigating the levels of Acrp30 in vivo with ongoing mouse studies in relationship to body weight and Pca development. Support from DOD PC 050284 and The Hormel Foundation.
Appendix C.

Impact of Two Types of Obesity on Prostate Cancer in the TRAMP Mouse
Melissa J.L. Bonorden, Michael E. Grossmann, O.P. Rogozina, D. Joshua Liao, Joseph P. Grande and Margot P. Cleary
To be presented at AACR meeting Denver, CO April 2009

Epidemiological studies suggest that body weight plays a role in prostate cancer development and obesity is associated with higher cancer grade and greater recurrence and mortality rates. To clarify these issues two studies were undertaken using the TRAMP mouse model of prostate cancer. First obesity was induced at 6, 16 and 26 wk of age by injection of gold-thioglucose (GTG) (0.5-0.8 mg/kg body weight in phosphate buffered saline (PBS). Mice were weighed and palpated weekly to detect prostate tumors until 46 wk of age or until disease burden necessitated euthanasia. Overall GTG had high toxicity and mortality rates in TRAMP mice (in contrast to preliminary studies in wildtype C57BL6 mice) resulting in limited data from the 6- and 26-wk cohorts. For the 26-wk cohort 14 mice survived the GTG injection, of which 8 were obese (48.6 ± 2.6 grams) (ANOVA p <0.05) compared to the non-Obese GTG mice with a body weight of 39.1 ± 2.5 grams, which was similar to PBS injected mice (36.7 ± 1.63). Fat pad weights had a similar relationship. Genital-urinary tract (GUT) weights were not affected by body weight. Age of prostate tumor detection (~33 wk) or death (~43 wk) was not different among the groups. Similar results were obtained for the 6-wk cohort despite their much longer exposure to obesity. In general GTG-Obese mice had lower metastases rates although GUT pathology was similar to lean mice. In the second study TRAMP mice were fed a moderately high fat (33% fat calories) diet from 6 wk of age. Based on body weight gain from 6-18 wk of age mice were divided into Obesity-Prone, Overweight and Obesity-Resistant groups (n=24). A Low-Fat group (n=24) was included for comparison to the Obesity-Resistant mice. Due to low body weights mice that died prior to 30 wk of age were removed from calculations resulting in final numbers ranging from 16-21/group. Final body weights of Obesity-Prone mice were significantly heavier than Obesity-Resistant mice. Fat pad weights of Obesity-Prone mice were significantly heavier than all other groups. When normalized to body weight, fat pad weights of Obesity-Prone mice were significantly greater than those of Low-Fat mice. There were no significant effects of body weight or diet on GUT weight or GUT relative to body weight among the groups. There were no differences in age to tumor detection (29-32 wk) or death (37-41 wk) among the groups. Metastases rates (63-73%) were similar for all groups except for the Obesity-Resistant mice which had a rate of 43%. There was a trend for Obesity-Prone and Overweight mice to have lower incidence of PIN and higher incidence of moderately to poorly differentiated prostate tissues compared to the Obesity-Resistant and Low-Fat mice. These findings are consistent with epidemiological evidence indicating obesity’s role in prostate cancer is associated with more aggressive disease. Diet-induced obesity provided a better obesity related model than did GTG in TRAMP mice. (Support AICR, DOD and The Hormel Foundation).
Appendix D

Role of the Adiponectin Leptin Ratio in Prostate Cancer

Michael E. Grossmann, Nancy K. Mizuno, Melissa J.L. Bonorden, Amitabha Ray, Irina Sokolchik, Meena L. Narasimhan and Margot P. Cleary

Abstract

The effects of obesity on prostate cancer are complex and controversial. In order to better understand these effects we have investigated two different proteins that are secreted by adipose tissue. Adiponectin and leptin have been studied individually for how they affect proliferation, signaling and apoptosis of prostate cancer. However, while both are expressed by virtually all people the effect of the ratio of the two on prostate cancer initiation and progression remains to be elucidated. Transgenic adenocarcinoma mouse prostate (TRAMP) mice develop prostate cancer in a manner similar to that of humans. The TRAMP-C2 cell line was derived from a TRAMP tumor and can be utilized to investigate effects of various treatments in vitro or in vivo. Here we have investigated the effects of adiponectin, leptin or the combination of both in this model of prostate cancer. We found that tumors from TRAMP mice express both adiponectin and leptin receptors. Also, addition of adiponectin to TRAMP-C2 prostate cancer cells reduced proliferation. Leptin treatment alone did not significantly alter proliferation. However, when leptin was combined with adiponectin at ratios seen in obese individuals, leptin was able to block the ability of adiponectin to reduce cell proliferation. Adiponectin and leptin can alter signaling of the ERK pathway as shown by changes in phosphorylation. The plant protein osmotin has previously been shown to activate adiponectin receptors and we found that osmotin reduces proliferation of TRAMP-C2 cells in a manner similar to adiponectin. This work suggests that adiponectin, leptin and their associated receptors may play an important role in prostate cancer.