Enantiomeric Analysis of Ephedrines and Norephedrines

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Final Report

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16. Abstract  
Concerned with variations in abuse potential and control status among various isomers of ephedrines and norephedrines, this study was conducted to develop an effective method for the simultaneous analysis of eight ephedrine-related compounds along with structurally similar cathinones. Among various approaches studied, a 60m HP-5MS (0.25 mm ID, 0.25 µm film thickness) was successfully used to characterize the following compounds that were derivatized with (-)-α-methoxy-α-trifluoromethylphenylacetic acid (MTPA): (+)-cathinone, (-)-cathinone, (+)-norephedrine, (-)-norephedrine, (+)-norpseudoephedrine, (+)-ephedrine, (-)-ephedrine, (-)-pseudoephedrine, (+)-pseudoephedrine. (-)-Cathinone standard was not available but should also be resolvable under this analytical procedure. This method was successfully applied to the analysis of selected cold remedies for characterizing the enantiomeric compositions of the compounds present in these samples.

17. Key Words  
Enantiomer, Ephedrine, Norephedrine, Chiral Derivatization

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**INTRODUCTION**

“Chirality” is currently a topic at the forefront of academic research, as evidenced by the award of the 2001 Noble Prize in Chemistry to “three scientists who devised techniques for catalytic asymmetric synthesis — the use of chiral catalysts to accelerate the production of single-enantiomer compounds for pharmaceutical use and a wide range of other applications.”(1) In the pharmaceutical industry, drug firms are actively involved in developing new drugs as single enantiomers and in carrying out “racemic switches” — redeveloping racemic mixture drugs as single enantiomers — resulting in a significant increase in the percentage of drugs marketed as single enantiomers.(2)

Enantiomeric analysis of abused drugs is also an important issue in forensic laboratories. Data resulting from enantiomeric analysis can (a) provide information for sentencing guidance for certain drug-related offenses; (b) assist in drug-related investigations; and (c) determine whether the drug of concern is derived from a controlled substance. For example, ephedrine and pseudoephedrine (ψ-ephedrine) are common over-the-counter (OTC) pharmaceuticals. They are also frequently used as adulterants in packaging drugs of abuse.(3) (–)-Ephedrine has been a popular precursor for illicit manufacturing of (+)-methamphetamine.(4,5) Investigation of clandestine laboratory activities reported (6,7) the use of ephedra plant (Ma Huang) material for methamphetamine manufacturing; (–)-ephedrine and (+)-ψ-ephedrine in this plant are extracted for conversion to methamphetamine in these illicit manufacturing processes. Thus, the identification of ephedrine and ψ-ephedrine, and their enantiomeric composition in methamphetamine samples, may help identify the drug’s precursor material and provide valuable information to the investigation process. Also of significant analytical concern is the reported false methamphetamine identification in urine specimens due to excessive consumption of ephedrine and ψ-ephedrine.(8)

Enantiomeric analysis of abused drugs in the authors’ laboratories date back to 1981, mainly involving gas chromatographic and nuclear magnetic resonance spectrometric approaches.(9–13) More recent studies utilized liquid chromatography and capillary electrophoresis.(14,15) Concerned with the presence of ephedrine-related compounds in OTC cold remedies and its implications in sport drug testing (16), this study was conducted to develop a method that can be effectively used to determine the enantiomeric compositions of the following structurally related compounds: ephedrines, ψ-ephedrine, norephedrines (phenylpropanolamine, or PPA), norpseudoephedrine (nor-ψ-ephedrine, or cathine), and cathinones. Methods thereby developed were then applied to selected OTC cold remedies to detect the presence and enantiomeric compositions of these compounds.

**EXPERIMENTAL**

**Standards and reagents**

R(+)-Cathinone, S(–)-cathinone, S,R(+)-ephedrine, R,S(–)-ephedrine, S,S(+)–ψ-ephedrine, R,R(–)-ψ-ephedrine, S,S(+)-norephedrine, Internal standard (S,R(+)-ephedrine-d₃, 1 mg/mL in methanol) were purchased from Cerilliant Int. Co. (Austin, TX); S,S(+)-nor-ψ-ephedrine (1 mg/mL) was purchased from Sigma Co. (St. Louis, MO). The structures of these compounds are shown in Figure 1.

Chiral derivatization reagents and their sources are as follows: (–)-α-methoxy-α-trifluoromethylphenylacetic (MTPA), (S)-(–)-N-(trifluoroacetyl)-prolyl chloride (l-TPC) (Aldrich: St. Louis, MO); 2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyl isothiocyanate, R-(+)–α-phenylethyl isocyanate, 2,3,4-triacetyl-α-D-arabinopyranosyl isothiocyanate (Fluka Chemie Gmbh: Industriestrasse, Buchs, Switzerland). Achiral derivatization reagents and their sources are: 9-fluorenylmethyl chloromate, N-(phenylseleno)-phthalimide, N,O-bis(trimethylsilyl)-acetamide (BSA), pentafluoropropionic anhydride (PFPA), heptafluorobutyric anhydride (HFBA) (Aldrich: St. Louis, MO); 4-carboethoxyhexafluorobutyryl chloride (4-CB) (Lancaster: Windham, NH).

Nineteen readily available OTC cold remedies (13 syrup, 6 capsule) were purchased from local drug stores in the greater Taipei area.

**Sample preparation**

Typical extraction, derivatization, and GC-MS analysis studies utilized 2 mL of standard mixtures or specimens. Standard mixtures were prepared to contain 1000 ng of each analyte following the general procedure described below. Standards obtained from the suppliers (typically 1 mg/mL in methanol) were first diluted to 10 µg/mL (in ethanol). 100 µL of each standard was then taken and mixed into 2 mL of drug-free syrup.
The preparation of OTC samples was as follows. Those in syrup forms were diluted (typically diluting 100 µL to 2 mL), while those in capsule forms were emptied and dissolved into 10 mL of blank syrup with further dilution (typically diluting 20 µL to 2 mL).

To determine extraction efficiency, the following procedure was used to derive the amount of the analytes resulting from the process without the extraction step. A mixture containing all analytes of interest was prepared by mixing 100 µL each of the diluted standards (10 µg/mL) in a clean tube. The mixture was dried under nitrogen and then processed in parallel with other standard mixtures that had completed the extraction step.

**Derivatization procedure**

Standard mixtures and OTC specimens in aqueous solutions were extracted and derivatized following either a one-step or two-step procedure as described below. Using l-TPC as an example, the one-step procedure involved mixing 2 mL sample, 100 µL internal standard ((+)-ephedrine-d₃, 10 µg/mL), 0.5-mL saturated K₂CO₃ solution, 50 µL l-TPC, and 6 mL ethyl acetate for 10 min. The mixture was then centrifuged (5 min), followed by removing the upper layer to a clean tube that was dried under a nitrogen stream. The residue was typically reconstituted with 200-µL ethyl acetate of which 1 µL was used for each GC-MS analysis.

Using MTPA derivatization as an example, the two-step process was carried out as follows. Typically, the internal standard, 2 mL standard mixture (or specimen), and 0.5 mL saturated K₂CO₃ solution were mixed for 30 sec. The mixture was then extracted with 6-mL ethyl acetate by shaking (10 min), followed by centrifugation (5 min). The upper layer was transferred into a clean tube and dried under nitrogen. For the derivatization step, the residue was added 50 µL N,N-dicyclohexycarbodiimide and 100 µL MTPA. The reaction mixture was thoroughly mixed, then incubated at 70°C for 20 min. This same two-step procedure was used when HFBA was used for derivatization, except that 1 mL of 2 N NaOH, instead of 0.5 mL saturated K₂CO₃ solution, was used prior to the addition of ethyl acetate for extraction.

**GC-MS analysis**

GC-MS analysis was performed on a HP 5890 Series II GC interfaced to an HP 5971 MS (Agilent: Palo Alto, CA). Two columns used in this study were: 25 m HP 5MS (0.20 mm ID, 0.33 µm film thickness) and 60 m HP 5MS (0.25 mm ID, 0.25 µm film thickness) from Agilent (Wilmington, DE). Helium carrier gas flow rate was 1.0 mL/min. The injector and GC-MS interface temperatures were maintained at 250 and 280°C, respectively. Temperature of the GC oven was programmed using different parameters for the analysis of products derived from different derivatization reagents (Table 1). For the 60 m column, a typical GC-MS run took 30 min or less.

The MSD was initially operated under full-scan mode to derive the retention time and full-scan mass spectrum information for each analyte. This information was then used to identify each analyte in standard mixtures and OTC specimens. Full-scan mass spectra were further used for the selection of ions suitable for use in selected ion monitoring (SIM) mode.

**RESULTS AND DISCUSSION**

**Resolution**

As shown in Section 2.1, a total of 11 derivatization reagents (5 chiral and 6 achiral) were included in this study. MTPA was found to be the most effective enantiomeric derivatization reagent, allowing complete base-line resolution of the 10 structurally closely related compounds of interest shown in Figure 2. (–)-Cathine was not available for this study; however, it should have been resolved, were it included in the mixture.

Shown in Figure 3 are the mass spectra of (+)-Ψ-norephedrine (A), (+)-norephedrine (B), (+)-Ψ-ephedrine (C), (+)-Ψ-ephedrine (D), and (+)-ephedrine-d₂ (E) (all as MTPA derivatives). Mass spectra of the corresponding (–)-omers are practically indistinguishable and, therefore, are not shown.

Derivatization products resulting from the commonly used l-TPC also result in good resolution of the analytes, with the exception of (–)-ephedrine and (–)-Ψ-ephedrine. Furthermore, ions that may be used for designating the analytes and their deuterated analogs are less characteristic.

When the determination of analytes’ enantiomeric compositions is not needed, HFBA-derivatization was found very effective (Fig. 4). Analysis time can be further reduced by increasing the column temperature following the elution of cathine and norephedrine (peaks 1 and 2 in Fig. 4). Mass spectra of representative compounds with HFBA derivatization are shown in Figure 5.

**Evaluation of analytical parameters**

Common analytical parameters, such as limits of detection and quantitation (LOD and LOQ) and extraction efficiency, have been studied. Evaluations were performed on MTPA and HFBA derivatizations using ephedrine as the exemplar compound. Results listed in Table 2 were established using the criteria and procedure described below.

Commonly adapted criteria were used to confirm the presence of a specific analyte in a test sample, i.e., ions...
monitored for a specific analyte have to present at an acceptable retention time (±2%) with acceptable intensity ratios (±20%) of that established by a standard. The LOD was defined as the lowest concentration of a standard solution meeting the above criteria, while LOQ was defined as the lowest concentration of a standard solution that met these criteria and with an observed analyte concentration that is within ±20% of the targeted value.

A series of standard solutions with the following concentrations of ephedrine were used for LOD and LOQ evaluations: 2000, 1000, 500, 250, 100, 80, 60, 40 ng/mL. Applying the criteria described above, the method’s LOD and LOQ were determined to be 60 and 80 ng/mL for both HFBA and MTPA derivatives.

**Application to the analysis of common OTC cold remedies**

As reported in an earlier study (16), various ephedrine-related compounds were found in readily available OTC cold remedies. Attempts to correlate the occurrences and concentrations of these compounds in OTC remedies with the analytical findings derived from testing athletes during sport-competition events have not been conclusive. With this in mind, the authors thought an additional dimension of information (enantiomeric composition) may help studies of this nature. Thus, various chiral and achiral derivatization approaches were explored, of which the most effective ones were applied to the analysis of a limited number of OTC cold remedies (from 19 manufacturers). Preliminary data shown in Table 3 are promising, and further studies will be pursued and applied to a comprehensive list of OTC remedies, selected prescription medicines, and relevant urine specimen sets.

**CONCLUSION**

An effective methodology has been established for the analysis of the following structurally related compounds and their enantiomers: cathinone, ephedrine, Ψ-ephrine, norephedrine, and nor-Ψ-ephrine. Using MTPA as the derivatization reagent, the resulting products can be baseline resolved by a 60 m HP 5MS capillary column. HFBA is effective when enantiomeric compositions are not needed. Preliminary application studies have also shown great potentials in providing an additional dimension of information (enantiomeric compositions) for source-tracing studies.

**REFERENCES**


(8) Department of Health and Human Services, Notice to All DHHS/NIDA Certified Laboratories, Feb. 4, 1991.


Figure 1. Structures of ephedrine and structurally related compounds.
Figure 2. Ion chromatogram of MTPA-derivatives: (+)-Cathinone (1), (–)-cathinone (2), (+)-norephedrine (3), (–)-norephedrine (4), (+)-nor-Ψ-ephrine (5), (+)-ephrine-d₃ (6), (+)-ephrine (7), (–)-ephrine (8), (–)-Ψ-ephrine (9), and (+)-Ψ-ephrine (10) (all as MTPA-derivatives).
Figure 3. Mass spectra of (+)-Ψ-norephedrine (A), (+)-norephedrine (B), (+)-Ψ-ephedrine (C), (+)-ephedrine (D), and (+)-ephedrine-d₃ (E) (all as MTPA-derivatives).
**Figure 4.** Ion chromatogram of HFBA-derivatives: Cathine (1), (±)-norephedrine (2), (±)-ephedrine (3), (±)-γ-ephedrine (4), (+)-ephedrine-d₃ (5) (all as HFBA-derivatives).
Figure 5. Mass spectra of (+)-Ψ-norephedrine (A), (+)-norephedrine (B), (+)-Ψ-ephedrine (C), (+)-ephedrine (D), and (+)-ephedrine-d$_3$ (E) (all as HFBA-derivatives).
### Table 1. Gas chromatograph oven temperature programming parameters for the analysis of analytes resulting from three derivatization reagents.

<table>
<thead>
<tr>
<th>Derivatization reagent</th>
<th>Starting (°C)</th>
<th>Hold (Min)</th>
<th>Rate (°C/Min)</th>
<th>End (°C)</th>
<th>Hold (Min)</th>
<th>Rate (°C/Min)</th>
<th>End (°C)</th>
<th>Hold (Min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HFBA</td>
<td>60</td>
<td>0</td>
<td>5</td>
<td>200</td>
<td>0</td>
<td>25</td>
<td>250</td>
<td>—</td>
</tr>
<tr>
<td>l-TPC</td>
<td>160</td>
<td>5</td>
<td>5</td>
<td>250</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>MTPA</td>
<td>160</td>
<td>0</td>
<td>5</td>
<td>220</td>
<td>1</td>
<td>25</td>
<td>250</td>
<td>—</td>
</tr>
</tbody>
</table>

### Table 2. Evaluation of common analytical parameters resulting from FHBA and MTPA derivatizations.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>HFBA</th>
<th>MTPA</th>
</tr>
</thead>
<tbody>
<tr>
<td>Recovery (%) (a)</td>
<td>72 ± 4 (b)</td>
<td>90 ± 7 (b)</td>
</tr>
<tr>
<td>LOD (ng/mL)</td>
<td>60</td>
<td>60</td>
</tr>
<tr>
<td>LOQ (ng/mL)</td>
<td>80</td>
<td>80</td>
</tr>
</tbody>
</table>

\(a\) Evaluated using triplicates of 2-mL standard solutions containing 500 ng/mL ephedrine.

\(b\) Mean ± standard deviation.

### Table 3. Enantiomeric composition (µg/mL) of the targeted 10 analytes found in various cold remedies.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Derivative</th>
<th>Cathinone (+)</th>
<th>Norephedrine (+)</th>
<th>Nor-(\psi)-ephedrine (+) (c)</th>
<th>Ephedrine (+)</th>
<th>(\psi)-Ephedrine (+)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2</td>
<td>MTPA</td>
<td>—</td>
<td>—</td>
<td>0.039 (b)</td>
<td>1.84</td>
<td>0.565</td>
</tr>
<tr>
<td></td>
<td>HFBA</td>
<td>—</td>
<td>—</td>
<td>0.123</td>
<td>1.50</td>
<td>0.850</td>
</tr>
<tr>
<td>9</td>
<td>MTPA</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>2.39</td>
<td>1.02</td>
</tr>
<tr>
<td></td>
<td>HFBA</td>
<td>—</td>
<td>—</td>
<td>2.33</td>
<td>0.951</td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>MTPA</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>1.36</td>
</tr>
<tr>
<td></td>
<td>HFBA</td>
<td>—</td>
<td>—</td>
<td>1.27</td>
<td></td>
<td>0.600</td>
</tr>
<tr>
<td>15</td>
<td>MTPA</td>
<td>0.186</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td></td>
<td>HFBA</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>16</td>
<td>MTPA</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td></td>
<td>HFBA</td>
<td>—</td>
<td>—</td>
<td>0.549</td>
<td>48.1</td>
<td></td>
</tr>
<tr>
<td>17</td>
<td>MTPA</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td></td>
<td>HFBA</td>
<td>—</td>
<td>—</td>
<td>0.111</td>
<td>0.089</td>
<td></td>
</tr>
<tr>
<td>19</td>
<td>MTPA</td>
<td>1.35</td>
<td>26.8</td>
<td>2130</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td></td>
<td>HFBA</td>
<td>1.19</td>
<td>2340</td>
<td>—</td>
<td>0.940</td>
<td>—</td>
</tr>
</tbody>
</table>

\(a\) Below detection limit (60 ng/mL as established for ephedrine).

\(b\) Below LOD and LOQ as established for ephedrine. However, distinct chromatographic peaks and mass spectra were observed and the listed concentrations were estimated.

\(c\) No standard was available; thus, these analytical findings are tentative.