Evaluation of Levofloxacin Pharmacodynamics in a mouse Model of Inhalational *Bacillus anthracis*

H.S. Heine, W.R. Byrne, J. Bassett, L. Miller, USAMRIID
G. L. Drusano, Ordway Research Institute

Disclaimer
Opinions, interpretations, conclusions, and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.
**Evaluation of Levofloxacin Pharmacodynamics in a mouse Model of Inhalational Bacillus anthracis**

US Army Medical Research Institute of Infectious Diseases


14. **ABSTRACT**

15. **SUBJECT TERMS**

<table>
<thead>
<tr>
<th>a. REPORT</th>
<th>b. ABSTRACT</th>
<th>c. THIS PAGE</th>
<th>17. LIMITATION OF ABSTRACT</th>
<th>18. NUMBER OF PAGES</th>
<th>19a. NAME OF RESPONSIBLE PERSON</th>
</tr>
</thead>
<tbody>
<tr>
<td>unclassified</td>
<td>unclassified</td>
<td>unclassified</td>
<td>UU</td>
<td>23</td>
<td></td>
</tr>
</tbody>
</table>

Standard Form 298 (Rev. 8-98)
Prescribed by ANSI Std Z39-18
Antibiotic Assessment Plan

• Antibiotics licensed or in clinical human trials in the U.S.

• Screen antibiotics by in-vitro assay (MICs)

• Best candidates by MIC and pharmacokinetics (PK) tested in mouse aerosol challenge model

• Best candidates in mouse model tested in non-human Primate model.
Balb/c

- Inbred
- Intermediate sensitivity $\text{LD}_{50} 8 \times 10^4$
- Consistent with previous data by subcutaneous challenge (Welkos SL; I&I 1986)
Treated Infection Burden

Days

0 5 10 15 20 25 30 35 40 45 50
Conclusions

• **Balb/c mouse is a suitable model**
  – For inhalational anthrax
  – For screening of antibiotic efficacy in a rodent model
  – As a first step before evaluation of antibiotic efficacy in non-human primates

• **Ciprofloxacin 30mg/Kg, Q12 is an appropriate standard of effective treatment**
  – 21 days sufficient
  – 14 days not sufficient

• **Anthrax spore burden in the lung/mediastinum does not appear to be significantly affected by antibiotic treatment**
Peripheral (organ) Compartment ($C_p$) + Bacteria ($X_{T/R}$)

IP injection

Central Blood Compartment ($C_c$)

- $k_{cp} \rightarrow C_p$
- $k_{pc} \downarrow C_p$

\[ \frac{dC_p}{dt} = k_{cp}C_c - k_{pc}C_p \]

\[ \frac{dC_a}{dt} = -k_aC_a \]

\[ \frac{dC_c}{dt} = k_aC_a + k_{pc}C_p - k_{cp}C_c - k_eC_c \]

\[ \frac{dX_S}{dt} = K_{GS}X_S \times L - f_{KS}(C_c^{H\xi}) \times X_S \]

\[ \frac{dX_R}{dt} = K_{GR}X_R \times L - f_{KR}(C_c^{H\xi}) \times X_R \]

\[ L = (1 - (X_R + X_S)/POPMAX) \]

\[ f_{\psi\xi}(C_c^{H\xi}) = \frac{K_{max}\xi \cdot C_c^{H\xi}}{C^{H\xi}_{50\xi} + C_c^{H\xi}} \]

\[ Y_1 = X_T = X_S + X_R \]

\[ Y_2 = X_R \]
Terminology

MIC - Minimum Inhibitory Concentration

Cmax - Maximum concentration (PK)

AUC - Area under the Curve (PK)

ΔTMIC - Time above MIC (PK)

AUC/MIC ratio

Cmax/MIC
Hollow fiber System allows simulation of human PK in vitro

- Useful for dose ranging and schedule dependency determinations
- Allows examination of different classes (beta lactams, fluoroquinolones, etc.)

The original hollow fiber system was used by Blaser, Dudley & Zinner
Levo vs. B. anthracis

![Graph showing the comparison of Levo and B. anthracis over time.](image)

- **Control**
- **Human 300**
- **Monkey 150**
- **Monkey 200**
- **Monkey 300**
- **Monkey 500**
- **Monkey 1000**
- **Cipro**

*Note: Log CFU values are plotted against time (days) for each category.*
Levo vs. B. anthracis

Control
Levo Human AUC/MIC = 300
Cipro AUC/MIC = 256 QD
Cipro AUC/MIC = 128 Q12h
Levo AUC/MIC = 183 Q12h
Levo AUC/MIC = 183 H0, 62.5 H8, 37.5 H12
Levo AUC/MIC = 183 H0, 50 H12
Preliminary Conclusions

- Levofloxacin achieved stasis (at best) on a once-daily schedule of administration with rhesus pharmacokinetics.
- Levofloxacin eradicated *Bacillus anthracis* when human PK was employed.
- Emergence of resistance was seen for the lower AUC/MIC ratio regimens in a time-dependent manner for levofloxacin.
In order to determine the validity of these findings, we designed and carried out an aerosol challenge of the Ames strain of *Bacillus anthracis* in a Balb/c mouse model.

- The challenge amount was 50-75 LD$_{50}$ of Ames spores (LD$_{50} = 8 \times 10^4$ spores)
- Schedules of Q6h, Q12h and Q24h were evaluated,
- Total daily doses of 37.5, 75, 150, 225, 300 mg/Kg
• Population modeling of mouse levofloxacin PK was performed \(T1/2 = 1.1 \text{[mean PK values]} - 1.8 \text{[median PK values]} \text{ h})

• AUC/MIC ratios of 0 (control) to 176 were examined on each administration schedule

• Treatment was for 21 days

• Surviving animals were followed off therapy

• Time to death was one endpoint examined by stratified Kaplan-Meier analysis and also by Cox Proportional Hazards Modeling
Schedule of administration was highly significant ($p < 0.000001$)

Schedule is best handled as a stratification variable, not as a covariate

AUC/MIC was not significant alone, but was a a covariate when added to Schedule as a stratification variable ($p = 0.0012$)
When Q24h dosing is removed, there is no difference by Schedule.

Now AUC/MIC is highly significant alone ($p = 0.00012$).
Total Counts

Spore Counts (heat shock)
Preliminary Conclusions

- The mouse system showed that the adverse impact of Q24h dosing schedule predicted by the hollow fiber system did, indeed, occur.
- Human PK and “Humanized” rhesus PK killed the organisms with NO emergence of resistance for levofloxacin.
- When ciprofloxacin was administered as twice the exposure once daily (same 24 hour exposure), there was immediate (24 hour) emergence of resistance and regimen failure.
- The ciprofloxacin failure with resistance is likely due to pump overexpression.
Preliminary Conclusions

- The mouse model demonstrated good effect with relatively low AUC/MIC ratios on Q12h and Q6h dosing (likely due to an intact immune system)

- The levofloxacin failures with resistance were probably due to target site mutation (not proven)

- We are currently investigating the impact of sporulation
Preliminary Conclusions

- The rhesus challenge with Levo/Cipro Rx is taking place currently
- Good protection is being seen with “Humanized” levofloxacin dosing
- The hollow fiber model predicts both success (rhesus) and failure (mouse Q24h dosing)
**Lab Animal Usage**

Research was conducted in compliance with the Animal Welfare Act and other Federal statutes and regulations relating to animals and experiments involving animals and adheres to principles stated in the Guide for the Care and Use of Laboratory Animals, National Research Council, 1996. The facility where this research was conducted is fully accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International.

**Funding Acknowledgement**

The research described herein was sponsored by the (ie. Medical Biological Defense Research Program, U.S. Army Medical Research and Materiel Command)] [Project/Grant/Agreement/Contract No. X].
Acknowledgements

THE “A” TEAM
Jennifer Bassett               Col. Russ Byrne
Lynda Miller                  Dr. Dave Waag
Anthony Bassett               Dr. Bruce Ivins
Marilyn England               Dr. Kei Amemiya
Steve Tobery                  Dr. George Drusano
Vet Med Div.
Aerobiology Dept.