Finite Element Analysis of the Hierarchical Structure of Human Bone

The objective of this study was to develop an analytical model of the basic hierarchical structure of the human bone. The model computed the stiffness of composite collagen fibers comprised of collagen fibrils and hydroxyapatite mineral crystals. Next, the stiffness of the concentric lamella was computed utilizing the stiffness of the collagen fibers and layer information. Finally, the effective stiffness of the bone was estimated. In order to determine the stiffness of the collagen fiber, a three-dimensional finite element model was developed and a simple analytical model was derived. The simple analytical model was validated using the finite element results. The lamination theory of unidirectional fibrous composites was used to calculate the stiffness of the lamella and eventually the bone stiffness. A series of parametric studies were conducted to understand what parameter(s) affected the stiffness of the bone most significantly. This information will be useful when an artificial bone structure is designed.
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FINITE ELEMENT ANALYSIS OF THE HIERARCHICAL STRUCTURE OF HUMAN BONE

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ABSTRACT

The objective of this study was to develop an analytical model of the basic hierarchical structure of the human bone. The model computed the stiffness of composite collagen fibers comprised of collagen fibrils and hydroxyapatite mineral crystals. Next, the stiffness of the concentric lamella was computed utilizing the stiffness of the collagen fibers and layer information. Finally, the effective stiffness of the bone was estimated. In order to determine the stiffness of the collagen fiber, a three-dimensional finite element model was developed and a simple analytical model was derived. The simple analytical model was validated using the finite element results. The lamination theory of unidirectional fibrous composites was used to calculate the stiffness of the lamella and eventually the bone stiffness. A series of parametric studies were conducted to understand what parameter(s) affected the stiffness of the bone most significantly. This information will be useful when an artificial bone structure is designed.
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I. INTRODUCTION

This thesis research developed both an analytical model and a finite element model of the hierarchical structure of human bone. The analytical model was validated using the results from the finite element model. Before we delve into the aspects of each model, it is important to understand the structure of the human bone. Figure 1 shows molecular to ultra to micro to macro components of the bone, which will be discussed in the subsequent section.

Figure 1. Hierarchical Structure of Human Bone (From: Rockwood, 2001)
A. HUMAN BONE STRUCTURE

Starting with the molecular structure of bone we have hydroxyapatite mineral crystals, commonly known as apatite. These mineral crystals provide for the hardness and brittle nature of bone (Apatite, 2001).

The hydroxyapatite lies in sections around a collagen fibril. The collagen has great tensile strength and allows for bone’s elastic behavior which complements the properties of the apatite (Collagen, 2001). Together these materials make up the composite fiber and account for the molecular structure of bone.

The ultra component of the bone structure consists of many concentric lamellas. In nature these lamella are oriented $45^\circ$ from the longitudinal axis. Each layer of the lamellae are at opposing angles and there are 3-20 lamella comprising the lamellae (Caceci, 2001).

The osteon or the Haversion system is the micro component of bone. This is made up of many lamellae encircling an inner region to provide protection. This inner region consists of:

- The Haversion canal which contains one or two small blood vessels and a nerve (Osteon, 2002).
- Lacunae; which are holes for one bone cell (osteocyte) to live (Osteon, 2002).
- Canaliculi which enable osteocytes to communicate with one another (Osteon, 2002).
- Volkman’s canal which enables adjacent osteons to communicate with each other (Osteon, 2002).
The macro components of bone make up bone as most of us know it today. As seen in Figure 1, there are many different parts which make up the outer structure of the bone. This research did not account for every one, but instead grouped them together to account for the 30% of bone which was not analyzed with the finite element model or with the analytic predictions. In other words our final values for the properties of bone were based upon the lamella structure and multiplied by 70%.

B. OBJECTIVES

The objective of this study was to develop an analytical model of the basic hierarchical structure of the human bone, consisting of collagen and hydroxyapatite. The model computed the stiffness of composite collagen fibers comprised of collagen fibrils and hydroxyapatite mineral crystals. Once the model was developed, the stiffness of the concentric lamella was computed utilizing the stiffness of the collagen fibers and layer information. Finally, the effective stiffness of the bone was estimated.

In order to determine the stiffness of the collagen fiber, a three-dimensional finite element model was developed and a simple analytical model was derived. The simple analytical model was validated using the finite element results. The lamination theory of unidirectional fibrous composites was used to calculate the stiffness of the lamella and eventually the bone stiffness. A series of parametric studies were conducted to understand what parameter(s) affected the stiffness of the bone most significantly.
II. FINITE ELEMENT MODEL

A. MODEL OVERVIEW

A three-dimensional, fixed displacement, finite element model of the most basic human bone structure was developed that is capable of predicting the total stress of the composite fiber and therefore, through analytical calculations, the elastic modulus of the composite fiber. The model is on a nanometer scale and represents one section of the composite fiber, including collagen and hydroxyapatite mineral crystals. The commercial finite element package MSC/PATRAN was used for pre and post processing and MSC/NASTRAN was used for analysis.

B. GEOMETRY

One section of the composite fiber was modeled, as shown in Figure 2. Beginning with a solid cylinder to represent a portion of the collagen fibril, 64 nm in length with a radius of 60 nm, hydroxyapatite mineral crystals, 48 nm in length and 2nm in width, were placed on the outside of the cylinder. These two materials made up the composite collagen fiber. The elastic modulus of each component was varied in order to analyze the individual effect on the elastic modulus of the composite. The stiffness of collagen was adjusted from 10 to 30 GPa at 5 GPa intervals. Separately, the stiffness of hydroxyapatite was adjusted from 700 GPa to 450 GPa at 50 GPa intervals. These values were chosen at random to determine the trend of the composite stiffness as each value was changed.
C. STATIC RESPONSE

The finite element model was run multiple times with different values for the elastic modulus of collagen and hydroxyapatite to determine which material affected the elastic modulus of the composite most dramatically. Applying boundary conditions enabled us to fix the translation in the x-direction, fix the translation in the y-direction, and fix rotation in all directions. Applying a fixed displacement to the model in the z-direction, we were able to determine the stress at each point of the composite fiber. Through analytic calculations, we then determined the average stress over the cross sectional
area, to ultimately obtain the elastic modulus of the composite fiber \( E = \frac{\sigma}{\varepsilon} \), where \( E \) = elastic modulus, \( \sigma \) = stress, and \( \varepsilon \) = strain. Once these results were tabulated we used them to validate our analytic model.

Figure 3 is an example of the output of the model when \( E_{\text{apatite}} = 450 \text{ GPa} \) and \( E_{\text{collagen}} = 20 \text{ GPa} \). The various colors represent the stresses on the given portion of the composite material. The analytic calculations to determine the average stress were accomplished on the cross sectional area (face of the fiber). The model is on a nanometer scale, so to obtain a meaningful value, we need to divide each stress value by the average stress over the cross sectional area to obtain the stress concentration. For example, the white value for stress is \( 2.79 \times 10^{-11} \text{ N/nm}^2 \) which when converted to common dimensions, becomes \( 2.79 \times 10^7 \text{ N/m}^2 \) or 27.9 MPa. We divide this value of stress by the average stress obtained through earlier calculations and obtain, 6.50 stress per unit stress, the stress concentration. This value is explained to give the reader a general idea of what they are seeing in Figure 3, and will make more sense when we discuss our results.
From the finite element model, we gained insight into the composite fiber properties and used the results to validate a simple analytic model.
III. ANALYTIC SOLUTION

An analytic model was derived in a sequence of three separate computer programs. The first computed the stiffness of composite collagen fibers comprised of collagen fibrils and hydroxyapatite mineral crystals. Once this analytic model was validated against the results of the finite element model, the stiffness of the concentric lamella was computed utilizing the stiffness of the collagen fibers and layer information. Finally, the effective stiffness of the bone was estimated.

A. FORCE-DEFORMATION BEHAVIOR OF A TYPICAL UNIFORM AXIAL-DEFORMATION ELEMENT

An analytic model was derived and the results from the finite element model were used as validation for this model. As a starting point, a MATLAB computer program using uniform axial deformation (spring constant) theory was developed. It enabled the user to change the values for the elastic modulus of hydroxyapatite and collagen to determine the longitudinal stiffness of the composite fiber. Figure 4 shows a schematic of the composite fiber.

![Spring Constant Schematic of Composite Fiber](image)

Figure 4. Spring Constant Schematic of Composite Fiber
We can compare Figure 4 to Figure 5 to see how the spring model correlates to the composite model.

![Diagram showing sections L1, L2, L3](image)

Figure 5. 2-D Finite Element Model Schematic Showing Sections

The basic theory behind this solution is

$$\frac{1}{k} = \frac{1}{k_1} + \frac{1}{k_2} + \frac{1}{k_3}$$

termed the force-deformation behavior of a typical uniform axial-deformation element (Craig, 2000); where $k =$ the total stiffness coefficient (force required to produce a unit elongation of the member), $k_1 =$ the stiffness coefficient for the first section of composite, $k_2 =$ the stiffness coefficient for the second section of the composite, and $k_3 =$ the stiffness coefficient for the third section of the composite. Note that $k_2 = k_{2a} + k_{2c}$ and that $k_{2a} =$ hydroxyapatite stiffness coefficient for the second section and $k_{2c} =$ the collagen stiffness coefficient for the second section. Solving this equation for $k$ we find
and from theory we know that \( k = \frac{AE}{L} \), where \( A = \text{Area} \). We substitute this definition for \( k \), into the composite formula and find

\[
k_1 = \left( \frac{A_1 E_c}{L_1} \right) = \frac{\pi r^2 E_c}{L_1}
\]
\[
k_3 = \left( \frac{A_3 E_c}{L_3} \right) = \frac{\pi r^2 E_c}{L_3}
\]

but the solution for \( k_2 \) is somewhat more involved as it has to be treated as a composite material. In determining \( k_2 \) we analyze a cross section of the mineral crystal represented in figure 6, where \( \sigma = \text{normal stress} \), \( \tau = \text{shearing stress} \), \( d\sigma = \text{change in normal stress} \), \( dz = \text{length of the cross section} \), and \( s = \text{width and height of cross section} \).

![Cross section of Hydroxyapatite](image)

Figure 6. Cross section of Hydroxyapatite

The first step is to sum the forces over the cross section, and from statics we know that \( \sum F = 0 \) and \( \sigma = \frac{F}{A} \). Utilizing
this, we find that \( \sum F = (\sigma + d\sigma) s^2 - \sigma s^2 - \tau sdz = 0 \). Simplifying this equation and integrating we find that \( \sigma(z) = \sigma_o + \frac{\tau z}{s} \) where \( \sigma_o = 0 \) as there is no stress at the end of the cross section. Assuming that \( \tau \) is constant, \( \sigma = \frac{\tau z}{s} \) and the maximum stress occurs at the center of the section, as seen in figure 7,

\[
\sigma_{\text{max}} = \frac{\tau l}{2s} \quad \text{and} \quad \sigma_{\text{avg}} = \frac{\tau l}{2s} = \frac{\tau l}{4s}.
\]

Figure 8 represents the shear stress over one section.
Figure 8. Shear Stress Over One Section of Hydroxyapatite

From figures 7 and 8 we find that $F_{avg} = \sigma_{avg} A = \frac{\tau s}{4} = \frac{F_{shear}}{4}$. From this equation we determine that our value for $k_{2a}$ must have a factor of 0.25 in the equation, therefore,

$$k_2 = \left( \frac{A_a E_a}{L_2} \right) + \left( \frac{A_c E_c}{L_2} \right) = \left( \frac{n A_a E_a}{4L_2} \right) + \left( \frac{\pi r^2 E_c}{L_2} \right)$$

where $r =$ radius, the $c$ subscript represents collagen, the $a$ subscript represents apatite, and $n =$ the number of sections of apatite around the fibril. We can now
determine \( k \), which in turn enables us to determine the elastic modulus of the composite fiber, using

\[
E_{\text{composite}} = \frac{kl}{\pi r^2}
\]

The value obtained here for \( E_{\text{composite}} \) is the longitudinal elastic modulus of the composite fiber. The results from this analytic process were used to validate our model and as a basis to move on to the next step which was to find the elastic modulus of the composite fiber when oriented at various angles.

**B. LAMINATION THEORY OF UNIDIRECTIONAL FIBROUS COMPOSITES**

The lamination theory of unidirectional fibrous composites was utilized to derive the next portion of the analytic model. The value for the longitudinal elastic modulus of the composite fiber was the starting point. This value was used in the transformed stiffness matrix to produce the elastic modulus of a single fiber oriented at various angles. The angles of orientation included 15°, 30°, 45°, 60°, -15°, -30°, -45°, and -60°.

The following stress-strain relations for isotropic materials were used (Jones, 1975):

\[
\begin{bmatrix}
\sigma_1 \\
\sigma_2 \\
\tau_{12}
\end{bmatrix} =
\begin{bmatrix}
Q_{11} & Q_{12} & 0 \\
Q_{12} & Q_{22} & 0 \\
0 & 0 & Q_{66}
\end{bmatrix}
\begin{bmatrix}
\varepsilon_1 \\
\varepsilon_2 \\
\gamma_{12}
\end{bmatrix}
\]

in which \( Q_{ij} \) are the reduced stiffnesses; specifically, \( Q_{11} = \frac{E}{1-\nu^2} \) where \( \nu \) is Poisson’s ratio and assumed to be 0.3, and \( Q_{12}, Q_{22}, \) and \( Q_{66} \) are all zero. Using these results, we
then calculated the transformed reduced stiffnesses using the following stress-strain relations in the xy coordinate directions (Jones, 1975):

\[
\begin{bmatrix}
\sigma_x \\
\sigma_y \\
\tau_{xy}
\end{bmatrix} =
\begin{bmatrix}
\bar{Q}_{11} & \bar{Q}_{12} & \bar{Q}_{16} \\
\bar{Q}_{12} & \bar{Q}_{22} & \bar{Q}_{26} \\
\bar{Q}_{16} & \bar{Q}_{26} & \bar{Q}_{66}
\end{bmatrix}
\begin{bmatrix}
\varepsilon_x \\
\varepsilon_y \\
\gamma_{xy}
\end{bmatrix}
\]

in which \( \bar{Q}_{ij} \) are the transformed reduced stiffnesses; specifically (Jones, 1975),

\[
\bar{Q}_{11} = Q_{11} \cos^4 \theta + 2(Q_{12} + 2Q_{66}) \sin^2 \theta \cos^2 \theta + Q_{22} \sin^4 \theta
\]

\[
\bar{Q}_{12} = (Q_{11} + Q_{22} - 4Q_{66}) \sin^2 \theta \cos^2 \theta + Q_{12} \left( \sin^4 \theta + \cos^4 \theta \right)
\]

\[
\bar{Q}_{22} = Q_{11} \sin^4 \theta + 2(Q_{12} + 2Q_{66}) \sin^2 \theta \cos^2 \theta + Q_{22} \cos^4 \theta
\]

\[
\bar{Q}_{16} = (Q_{11} - Q_{12} - 2Q_{66}) \sin \theta \cos^3 \theta + (Q_{12} - Q_{22} + 2Q_{66}) \sin^3 \theta \cos \theta
\]

\[
\bar{Q}_{26} = (Q_{11} - Q_{12} - 2Q_{66}) \sin^3 \theta \cos \theta + (Q_{12} - Q_{22} + 2Q_{66}) \sin \theta \cos^3 \theta
\]

\[
\bar{Q}_{66} = (Q_{11} + Q_{22} - 2Q_{12} - 2Q_{66}) \sin^2 \theta \cos^2 \theta + Q_{66} \left( \sin^4 \theta + \cos^4 \theta \right)
\]

The transformed elastic modulus of the composite fiber was equivalent to \( \bar{Q}_{11} \). This value was then used as a starting point in the last step of the analytic model derivation to determine the actual elastic modulus of human bone.

C. LAMELLAE AND BONE ELASTIC MODULUS

Considering actual human bone structure, the composite fibers were wrapped in a concentric lamella, each oriented at opposing angles. From the previous section, we determined the elastic modulus of a given composite fiber oriented at a given angle. The last step in the sequence
to derive the analytic model started with these results and determined first, the elastic modulus of the lamellae structure, and second, the elastic modulus of bone.

The lamellae was assumed to have 12 layers, as an average between the 3–20 lamella stated in the references (Caceci, 2001). The transformed elastic modulus for each angle ($15^\circ$, $30^\circ$, $45^\circ$, $60^\circ$, $-15^\circ$, $-30^\circ$, $-45^\circ$, and $-60^\circ$), from the previous derivation using the transformed matrix, was the starting point. Radius and area ($A=\pi r^2$) calculations were made for each layer of the concentric lamella as well as for the total area of the lamellae. To calculate the elastic modulus of the entire lamellae, the following formula was used:

$$\frac{\sum_{i=1}^{12} A_i E_{\text{angle}}}{\text{TotalArea}}$$

Once the stiffness of the lamellae was obtained, the only thing which remained was to make an estimate to determine the actual stiffness of the entire bone. To do this, we assumed that the lamellae represented 70% of the human bone structure. We multiplied the lamellae stiffness by 0.7 to account for this 70% and obtained the elastic modulus of human bone.
IV. RESULTS AND DISCUSSION

A. EFFECT OF ELASTIC MODULUS OF COLLAGEN AND HYDROXYAPATITE ON STRESS CONCENTRATION OF COMPOSITE FIBER

From the figures below we determine that the elastic modulus of collagen has a similar impact on the stress concentration of the composite fiber as the elastic modulus of hydroxyapatite, although collagen may be somewhat more influential. The stress results of the finite element model enabled us to determine the stress concentration over the composite fiber utilizing the following formula:

\[
\text{Stress Concentration} = \frac{\text{Localized High Stress}}{\text{Uniform Stress over Remainder}} = \frac{\sigma_{\text{max}}}{\sigma_{\text{avg}}}
\]

1. Effect of Elastic Modulus of Collagen on Stress Concentration of Composite Fiber

Figure 9 shows how the stress concentration over the composite fiber decreases as the stiffness of collagen increases. This decrease is as expected because as the stiffness of collagen increases its value gets closer to the stiffness of apatite. From theory, the closer the stiffness of the two materials, the lower the stress concentration should be (Ugural, 1995). The results shown in Figure 9 agree with theory.
2. Effect of Elastic Modulus of Hydroxyapatite on Stress Concentration of Composite Fiber

Figure 10 shows that as the stiffness of apatite increases, the stress concentration increases or maintains its approximate value. As with Figure 9, these results agree with theory because as you increase the stiffness of apatite you are moving farther away from the stiffness of collagen, thereby increasing the stress concentration.
When analyzing Figure 9 and 10 together, we can look at the ratio of change of the stress concentration. For example, beginning with Figure 9 and a collagen stiffness of 10 GPa (with $E_{\text{apatite}} = 700$ GPa) the stress concentration is 10.5, a 50% change in collagen stiffness would give us 15 GPa and a stress concentration of 9.5. So there is one unit of change in stress concentration with a 50% change in collagen stiffness. Now, looking at Figure 10, and an apatite stiffness of 450 (Ecollagen = 15 GPa) the stress concentration is 8, a 50% change in apatite stiffness would give us 675 GPa and a stress concentration of 9. Here there is also one unit of change in stress concentration with a 50% change in apatite stiffness.
B. EFFECT OF ELASTIC MODULUS OF COLLAGEN AND HYDROXYAPATITE ON COMPOSITE ELASTIC MODULUS

Figures 11 and 12 show the effect of varying the elastic modulus of hydroxyapatite and collagen on the resulting elastic modulus of the composite. As discussed below, the changes in collagen have a much greater impact on the composite than do the changes in hydroxyapatite.

1. Effect of Elastic Modulus of Collagen on Elastic Modulus of Composite

Figure 11 shows as the elastic modulus of hydroxyapatite is kept constant, the elastic modulus of the composite increases linearly as the elastic modulus of collagen increases.

![Graph showing the effect of collagen elastic modulus on composite elastic modulus](image_url)

**Figure 11. Effect of Collagen Elastic Modulus on Composite Elastic Modulus**
Six different values for the hydroxyapatite elastic modulus are shown with each line on the graph.

2. **Effect of Elastic Modulus of Hydroxyapatite on Elastic Modulus of Composite**

Figure 12 shows the slight increase of the elastic modulus of the composite as the elastic modulus of hydroxyapatite is increased.

![Graph showing the effect of hydroxyapatite elastic modulus on composite elastic modulus.](image.png)

**Figure 12.** Effect of Hydroxyapatite Elastic Modulus on the Composite Elastic Modulus

The plot shows three different values for the elastic modulus of collagen with each different line. We see that varying the elastic modulus of hydroxyapatite does not have a dramatic effect on the resulting properties of the composite material.
C. FINITE ELEMENT MODEL COMPARED TO AXIAL DEFORMATION CALCULATIONS

After utilizing the finite element model to obtain values for the elastic modulus of the composite, we wanted to validate our simple analytic model using this data. Figures 13-18 plot the FEM data with our analytic data from axial deformation calculations. Both results compare very well.

1. Constant Collagen Elastic Modulus

Figures 13-15 show the proximity of our finite element model calculations to the analytic data obtained from axial deformation calculations when varying the elastic modulus of hydroxyapatite. Errors are likely due to approximations and hand calculations used to obtain average stress results from the finite element model.

Figure 13, specifically shows the difference between the FEM and analytic data for $E_{\text{collagen}} = 10$ GPa. Note that the lines have the same slope and are within reasonable tolerances of each other.
Figure 13. FEM vs Analytic Data for $E_{\text{collagen}} = 10$ GPa

Figure 14, specifically shows the difference between the FEM and analytic data for $E_{\text{collagen}} = 20$ GPa. The lines are almost right on top of each other, showing excellent correlation between the FEM data and the analytic calculations.
Figure 14. FEM vs Analytic Data for $E_{\text{collagen}} = 20$ GPa

Figure 15, specifically shows the difference between the FEM and analytic data for $E_{\text{collagen}} = 30$ GPa. This graph shows a bit more deviance from the FEM data to the analytic data, most likely due to the error in approximating the average stress used to find the composite elastic modulus, and due to round off error.
Overall, we determined from Figures 13-15, that when the elastic modulus of collagen is held constant, the FEM and analytic data correlate relatively well.

2. Constant Hydroxyapatite Elastic Modulus

Figures 16-18 show the proximity of our finite element model calculations to the analytic data obtained from axial deformation calculations when varying the elastic modulus of collagen. Errors are likely due to approximations and hand calculations used to obtain average stress results from the finite element model.

Figure 16, specifically shows the difference between the FEM and analytic data for $E_{\text{apatite}} = 450$ GPa. Again, we see that the values are relatively close.
Figure 16. FEM vs Analytic Data for $E_{\text{apatite}} = 450$ GPa

Figure 17, specifically shows the difference between the FEM and analytic data for $E_{\text{apatite}} = 550$ GPa. As with Figure 14, the lines are almost right on top of each other, showing excellent correlation between the FEM data and the analytic calculations.
Figure 17. FEM vs Analytic Data for $E_{\text{apatite}} = 550$ GPa

Figure 18, specifically shows the difference between the FEM and analytic data for $E_{\text{apatite}} = 650$ GPa. As with Figure 15, this graph shows a bit more deviance from the FEM data to the analytic data, most likely due to the error in approximating the average stress used to find the composite elastic modulus, and due to round off error. The values are still relatively close.
Overall, we determined from Figures 16-18, that when the elastic modulus of hydroxyapatite is held constant, the FEM and analytic data correlate relatively well.

D. EFFECT OF VARIOUS LAMELLA ANGLES ON ELASTIC MODULUS OF BONE

Figures 19-24 show the elastic modulus of bone when each layer of the lamellae is oriented at various angles. For example, there are 12 concentric lamella, and the first lamella is oriented at 15°, the second at -15°, the third at 15°, the fourth at -15°, and so on, until you reach the twelfth lamella. Four possible orientations are shown in the graphs in this section.
1. **Constant Collagen Elastic Modulus**

Figures 19-21 show the elastic modulus of bone when the lamellae are oriented at various angles and the elastic modulus of collagen is kept constant. With all three figures it is obvious that the overwhelming contributor to the elastic modulus of bone, is the elastic modulus of collagen.

![Graph showing E bone effected by lamella angles (E collagen = 10 GPa)](image)

**Figure 19.** Bone Elastic Modulus with $E_{\text{collagen}} = 10$ GPa

Figure 19, specifically addresses the case when $E_{\text{collagen}} = 10$ GPa. We see that the various values for the elastic modulus of bone are similar for each value of hydroxyapatite, but as the degree of orientation is increased, the bone elastic modulus decreases, as expected.
The increasing angle of orientation moves each lamella further away from its strictly longitudinal configuration, and therefore causes the bone elastic modulus to continue to decrease. With collagen being so soft, the stiffness of bone remains relatively low for each value of $E_{\text{apatite}}$.

![Graph showing Bone Elastic Modulus with $E_{\text{collagen}}$ = 20 GPa](image)

**Figure 20.** Bone Elastic Modulus with $E_{\text{collagen}} = 20$ GPa

Figure 20, specifically addresses the case when $E_{\text{collagen}} = 20$ GPa. The same trend in $E_{\text{bone}}$ is recognized. The further away from zero degrees the lamella orientation gets, the lower the value for bone elastic modulus. We can see what a difference the harder collagen ($E = 20$ GPa vice $E = 10$ GPa) makes on the bone elastic modulus when comparing Figure 20 to Figure 19.
Figure 21. Bone Elastic Modulus with $E_{\text{collagen}} = 30 \text{ GPa}$

Figure 21, shows results with $E_{\text{collagen}} = 30 \text{ GPa}$. When raising collagen’s elastic modulus this high, we see a marked increase in the slope of each line, and therefore a greater increase in the property of bone. Although, when getting further away from the zero degree orientation (i.e., the $60^\circ$ orientation) the values for the stiffness of bone are very similar to previous values with the softer collagen. This is due to the high degree of orientation for each concentric lamella.

2. Constant Hydroxyapatite Elastic Modulus

Figures 22-24 show the elastic modulus of bone when the lamellae are oriented at various angles and the elastic modulus of hydroxyapatite is kept constant. With all three figures we see the same trend of bone stiffness, and
realize that the major contributor to its properties are that of collagen. Note the striking similarities between figures 22, 23, and 24.

![Diagram showing bone elastic modulus with different lamella angles and apatite modulus of 450 GPa.](image)

**Figure 22.** Bone Elastic Modulus with $E_{\text{apatite}} = 450$ GPa

Figure 22 shows the specific case of $E_{\text{apatite}} = 450$ GPa. As in the previous figures we see a decreasing trend of the elastic modulus of bone as the degree of orientation gets greater and greater. This is again due to the greater distance from the longitudinal orientation.
Figure 23. Bone Elastic Modulus with $E_{\text{apatite}} = 550$ GPa

Figure 23 shows the specific case of $E_{\text{apatite}} = 550$ GPa. Remarkably similar to Figure 22, providing an example of how the properties of bone are not reliant on the properties of hydroxyapatite, but instead on the properties of collagen.

Figure 24 depicts the same trend in results, but with $E_{\text{apatite}} = 650$ GPa.
Figure 24. Bone Elastic Modulus with $E_{\text{apatite}} = 650$ GPa
V. CONCLUSIONS AND RECOMMENDATIONS

A. CONCLUSIONS

The objective of this study was to derive an analytic model of the basic hierarchical structure of the human bone. The model computed the stiffness of composite collagen fibers comprised of collagen fibrils and hydroxyapatite mineral crystals. Next, the stiffness of the concentric lamella was computed utilizing the stiffness of the collagen fibers and layer information. Finally, the effective stiffness of the bone was estimated. In order to determine the stiffness of the collagen fiber and validate the analytic model, a three-dimensional finite element model was developed. Once the model was validated, a series of parametric studies were conducted to understand what parameter(s) affected the stiffness of the bone most significantly.

The results of the parametric study demonstrate that the elastic modulus of collagen is much more vital in determining the stiffness of bone than is the elastic modulus of hydroxyapatite. Both materials had a relatively similar effect on the stress concentration of the composite fiber. Therefore, if one were to construct an artificial human bone they would need to pay close attention to the material used to replicate collagen, and should not be so concerned about the material used for hydroxyapatite. Another important result, was that the lower the degree of orientation from the centerline, the higher the elastic modulus of bone. But what was not analyzed in this study was the effect of torsion on the bone stiffness. To combat
the effects of torsion, would require a higher degree of orientation of the lamella.

B. RECOMMENDATIONS

The following recommendations for future research include:

The current finite element model was analyzed for the axial load. Future work could consist of analyzing the model with different loading conditions including a torsional and a bending load.

This research utilized the elastic modulus of hydroxyapatite and collagen in determining the elastic modulus of bone. In the future, other bone properties such as the transverse modulus and shear modulus could be analyzed to determine if the same trends occur. The interface strength between the collagen and apatite could be another important parameter to analyze.
%This program is used to calculate the total stiffness coefficient for one section of collagen fiber (with apatite) as well as the Elastic Modulus of the composite fiber.

Theory

\[ \frac{1}{k} = \frac{1}{k_1} + \frac{1}{k_2} + \frac{1}{k_3} \]

\[ \frac{1}{k} = \frac{k_2 k_3 + k_1 k_3 + k_1 k_2}{k_1 k_2 k_3} \]

clear
clc

r = 60e-9; %radius in meters
Ec = 10e9; %Modulus of collagen in Pascals (N/m^2)
L1 = 8e-9; %length of initial and final section of collagen in meters
L2 = 48e-9; %length of apatite in meters
n = 12; %nr of pieces of apatite around circumference
Aa = 4.97955e-9*4*2e-9; %Area of apatite crystal (edge length*4*width)
Ea = 700e9; %Modulus of apatite in Pascals (N/m^2)
L = 2*L1 + L2; %Total length of composite

k1 = (pi*r^2*Ec)/L1;

alpha = 0.25;

k2 = k21 + k22 (collagen + apatite)
\%k2 = (AE/L)c + (AE/L)a \\
k2a = \alpha*(n*Aa*Ea)/L2; \\
k2c = (\pi*r^2*Ec)/L2; \\
k2 = k2a + k2c; \\
k3 = k1; \\

k = (k1*k2*k3)/((k1*k2) + (k2*k3) + (k1*k3)); \\

\%k = (AE/L)total \\
Ecomposite = ((k*L)/(\pi*r^2))
APPENDIX B – ELASTIC MODULUS OF COMPOSITE FIBER ORIENTED AT DIFFERENT ANGLES MATLAB PROGRAM

%To determine modulus of elasticity for collagen at 15, 30, 45, 60, -15, -30, -45, -60 degree angles.

clc
clear

nu = 0.3;
Efiber = 33e9;
data = [];
i=0;

%Try angles of 15,30,45,60,-15,-30,-45,-60 degrees

for theta = [(pi/12) (pi/6) (pi/4) (pi/3) (-pi/12) (-pi/6) (-pi/4) (-pi/3)];
    Q11 = Efiber/(1-nu^2);
    Q11bar = Q11*(cos(theta))^4;
    Q12bar = Q11*(sin(theta))^2*(cos(theta))^2;
    Q22bar = Q11*(sin(theta))^4;
    Q16bar = Q11*(sin(theta))*(cos(theta))^3;
    Q26bar = Q11*(sin(theta))^3*(cos(theta));
    Q66bar = Q11*(sin(theta))^2*(cos(theta))^2;

    %Qbar = [Q11bar Q12bar Q16bar; Q12bar Q22bar Q26bar; Q16bar Q26bar Q66bar];
    Qbar = zeros(3,3);  %Set matrix of zeros as most values are assumed zero
    Qbar(1,1) = Q11bar;  %Force value for (1,1)
    Qbar(2,2) = Q22bar;  %Force value for (2,2)
theta
Qbar;
S11 = 1/Qbar(1,1);
S22 = 1/Qbar(2,2);

E1bar = 1/S11                %Value for E1
E2bar = 1/S22;               %Value for E2
i = i+1;

data = [data; Q11bar Q22bar];
% disp([' Q11bar   Q22bar']);
data;

end
% Assuming that there are 12 layers of collagen which make up the lamella and that each layer is 120 nm thick. Take the values for E from the "transformedmatrix" m-file for various angles.

clc
clear

% theta = pi/12 (15 degrees)
E1bar15 = 31.568e+09;
E1barn15 = E1bar15;

% theta = pi/6 (30 degrees)
E1bar30 = 20.398e+09;
E1barn30 = E1bar30;

% theta = pi/4 (45 degrees)
E1bar45 = 9.0659e+009;
E1barn45 = E1bar45;

% theta = pi/3 (60 degrees)
E1bar60 = 2.2665e+009;
E1barn60 = E1bar60;

% radius calculations for each of 12 layers (assuming radius of each layer is equivalent to diameter of a single collagen fiber)
r1 = 60e-9;
r2 = r1 + 120e-9;
r3 = r2 + 120e-9;
\begin{verbatim}
r4 = r3 + 120e-9;
r5 = r4 + 120e-9;
r6 = r5 + 120e-9;
r7 = r6 + 120e-9;
r8 = r7 + 120e-9;
r9 = r8 + 120e-9;
r10 = r9 + 120e-9;
r11 = r10 + 120e-9;
r12 = r11 + 120e-9;

%Area calculations for each layer
A1 = pi*r1^2;
A2 = pi*(r2^2-r1^2);
A3 = pi*(r3^2-r2^2);
A4 = pi*(r4^2-r3^2);
A5 = pi*(r5^2-r4^2);
A6 = pi*(r6^2-r5^2);
A7 = pi*(r7^2-r6^2);
A8 = pi*(r8^2-r7^2);
A9 = pi*(r9^2-r8^2);
A10 = pi*(r10^2-r9^2);
A11 = pi*(r11^2-r10^2);
A12 = pi*(r12^2-r11^2);

%Total area of lamella
Atotal = pi*r12^2;
Atotalsum = A1+A2+A3+A4+A5+A6+A7+A8+A9+A10+A11+A12;

%Total Modulus of Elasticity (E1)

%Try opposite layers (15,-15,15,-15...)
\end{verbatim}
E_{1\text{total15}} = \frac{(A1*E_{1\text{bar15}}) + (A2*E_{1\text{barn15}}) + (A3*E_{1\text{bar15}}) + (A4*E_{1\text{barn15}}) + (A5*E_{1\text{bar15}}) + (A6*E_{1\text{barn15}}) + (A7*E_{1\text{bar15}}) + (A8*E_{1\text{barn15}}) + (A9*E_{1\text{bar15}}) + (A10*E_{1\text{barn15}}) + (A11*E_{1\text{bar15}}) + (A12*E_{1\text{barn15}}))}{A_{\text{total}}};

% Multiply by 0.7 to account for 70% of bone
E_{1\text{bone15}} = E_{1\text{total15}} \times 0.7

% Try opposite layers (30,-30,30,-30...)
E_{1\text{total30}} = \frac{(A1*E_{1\text{bar30}}) + (A2*E_{1\text{barn30}}) + (A3*E_{1\text{bar30}}) + (A4*E_{1\text{barn30}}) + (A5*E_{1\text{bar30}}) + (A6*E_{1\text{barn30}}) + (A7*E_{1\text{bar30}}) + (A8*E_{1\text{barn30}}) + (A9*E_{1\text{bar30}}) + (A10*E_{1\text{barn30}}) + (A11*E_{1\text{bar30}}) + (A12*E_{1\text{barn30}}))}{A_{\text{total}}};

% Multiply by 0.7 to account for 70% of bone
E_{1\text{bone30}} = E_{1\text{total30}} \times 0.7

% Try opposite layers (45,-45,45,-45...)
E_{1\text{total45}} = \frac{(A1*E_{1\text{bar45}}) + (A2*E_{1\text{barn45}}) + (A3*E_{1\text{bar45}}) + (A4*E_{1\text{barn45}}) + (A5*E_{1\text{bar45}}) + (A6*E_{1\text{barn45}}) + (A7*E_{1\text{bar45}}) + (A8*E_{1\text{barn45}}) + (A9*E_{1\text{bar45}}) + (A10*E_{1\text{barn45}}) + (A11*E_{1\text{bar45}}) + (A12*E_{1\text{barn45}}))}{A_{\text{total}}};

% Multiply by 0.7 to account for 70% of bone
E_{1\text{bone45}} = E_{1\text{total45}} \times 0.7

% Try opposite layers (60,-60,60,-60...)
E_{1\text{total60}} = \frac{(A1*E_{1\text{bar60}}) + (A2*E_{1\text{barn60}}) + (A3*E_{1\text{bar60}}) + (A4*E_{1\text{barn60}}) + (A5*E_{1\text{bar60}}) + (A6*E_{1\text{barn60}}) + (A7*E_{1\text{bar60}}) + (A8*E_{1\text{barn60}}) + (A9*E_{1\text{bar60}}) + (A10*E_{1\text{barn60}}) + (A11*E_{1\text{bar60}}) + (A12*E_{1\text{barn60}}))}{A_{\text{total}}};

% Multiply by 0.7 to account for 70% of bone
E_{1\text{bone60}} = E_{1\text{total60}} \times 0.7
%Try random layers (15,-15,30,-30,45,-45,15,-15,30,-30,45,-
%45)

\[ E_{1\text{total}r1} = \frac{(A1*E1\text{bar}15)+(A2*E1\text{bar}n15)+(A3*E1\text{bar}30)+(A4*E1\text{bar}n30)+(A5*E1\text{bar}45)+(A6*E1\text{bar}n45) + (A7*E1\text{bar}15)+(A8*E1\text{bar}n15) + (A9*E1\text{bar}30)+(A10*E1\text{bar}n30)+(A11*E1\text{bar}45)+(A12*E1\text{bar}n45))}{A_{\text{total}}}; \]

%Multiply by 0.7 to account for 70% of bone

\[ E_{1\text{boner}1} = E_{1\text{total}r1}*0.7 \]

%Try random layers (45,-45,60,-60,45,-45,60,-60,45,-45,60,-
%60)

\[ E_{1\text{total}r2} = \frac{(A1*E1\text{bar}45)+(A2*E1\text{bar}n45)+(A3*E1\text{bar}60)+(A4*E1\text{bar}n60)+(A5*E1\text{bar}45)+(A6*E1\text{bar}n45) + (A7*E1\text{bar}45)+(A8*E1\text{bar}n60) + (A9*E1\text{bar}45)+(A10*E1\text{bar}n45)+(A11*E1\text{bar}60)+(A12*E1\text{bar}n60))}{A_{\text{total}}}; \]

%Multiply by 0.7 to account for 70% of bone

\[ E_{1\text{boner}2} = E_{1\text{total}r2}*0.7 \]
LIST OF REFERENCES


INITIAL DISTRIBUTION LIST

1. Defense Technical Information Center
   Fort Belvoir, VA

2. Dudley Knox Library
   Naval Postgraduate School
   Monterey, CA

3. Professor Young W. Kwon ME/KW
   Department of Mechanical Engineering
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