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Overview:

During the past year, we have been focusing on the objectives outlined for Specific Aim 1 in our approved statement of work, and have completed an initial experiment related to Specific Aim 2. Testing and packaging of VEE replicon constructs encoding Her-2/neu, IL-12 and IL-18 have been completed. Additionally, a replicon construct encoding murine granulocyte-macrophage colony stimulating factor (mGM-CSF), has been established and tested and a truncated Her-2/neu gene encoding the extracellular and transmembrane domain of the protein is currently being assessed. We are hopeful the reduced size of the Her-2/neu gene will lead to increased levels of RNA and protein expression.

The rat Her-2/neu gene encodes for an 185kDa transmembrane protein. In order that we may have protein to work with experimentally, we have cloned overlapping fragments of the gene into a bacterial expression plasmid. Currently, one protein generated from the bacterial expression system, encompassing parts of the extracellular and transmembrane domains, has been used to demonstrate CD4+ T helper (Th) cell proliferation (Figure 1). For this experiment, mice were immunized intraperitoneally with 100µg of the protein fragment emulsified in incomplete Freund’s adjuvant (IFA). Splenocytes were then recovered and assessed for T cell proliferation by 3H-thymidine uptake. Both rat HER-2/neu and A2-Kβ transgenic mice showed significant proliferative responses. A second fragment, further spanning the extracellular domain, is in the process of being tested.

We have begun to characterize the CD8+ T cell response induced by replicons encoding Her-2/neu in both wild type FVB, and in FVB mice transgenic for the rat HER-2/neu gene. We have repeatedly generated significant CTL responses in wild type FVB mice, but CTL reactivity induced in Her-2/neu transgenic mice has to date been limited (Figure 2). The work of Jaffee et al. has demonstrated that Neu-specific T cell responses can be generated in Her-2/neu transgenic mice immunized with recombinant vaccinia virus expressing Neu. Consequently, we are hopeful that the appropriate combination of replicons encoding Her-2/neu as well as key inflammatory cytokines will overcome this element of immune tolerance seen in the transgenic mice.

One experiment has been completed in which both wild type FVB and Her-2/neu transgenic mice were immunized with replicons encoding Her-2/neu, and subsequently challenged with the F-H2N1 tumor cell line (expressing Her-2/neu). Both immunized and unimmunized Her-2/neu transgenic mice exhibited significant tumor growth whereas both groups of wild type mice rejected the tumor. While there was no significant difference in tumor weight from immunized and unimmunized Her-2/neu transgenic mice, decreased vascularity was noted in tumors recovered from the VEE Her-2/neu immunized mice. While we have no evidence that this decreased vascularity may be attributable to the vaccination, we remain optimistic that the correct combination of antigen and cytokine encoding replicons will be effective in inhibiting tumor growth.

Many of the tools are now in place that will allow us to test various antigen and cytokine combinations for optimal CD4+ Th and CD8+ CTL reactivity. This will further allow us to establish an effective approach of immunotherapy targeting Her-2/neu for the treatment and prevention of breast adenocarcinoma in transgenic mice.
Key Accomplishments:

- Completed *in vitro* testing and packaging of the following VEE replicons:
  
  Full length Her-2/neu  
  IL-12  
  IL-18  
  mGM-CSF  
  IL-2  

- Currently testing truncated Her-2/neu replicon encoding the extracellular and transmembrane domains.

- Cloned fragments of the Her-2/neu gene into a bacterial expression system and generated recombinant protein.

- Demonstrated CD4⁺ Th cell reactivity to one Her-2/neu protein fragment, and are in the process of testing a second fragment.

- Repeatedly generated CD8⁺ CTL reactivity in wild type FVB mice.

- Completed one tumor challenge experiment in VEE Her-2/neu immunized mice.

Reportable Outcomes:

- None.
Figures and Tables:

**Figure 1.** Proliferation in response to Her-2/neu fragment 4. Fragment 4 encompasses nucleotides 1856 – 2585 spanning part of the extracellular domain, the entire transmembrane domain and a small section of the intracellular domain. Her-2/neu transgenic and A2-Kb transgenic mice were twice immunized intraperitoneally with 100 µg fragment 4 emulsified in IFA. Splenocytes were harvested and incubated with the indicated concentration of recombinant protein and the levels of \(^3\)H-thymidine incorporation were measured. Data represents groups of three mice.

**Figure 2.** Standard 5 hr chromium release assay for FVB and FVB/Neu \(^{+/+}\) mice immunized with VEE replicons encoding Her-2/neu (3.0x10^5 IU) and replicons encoding mGM-CSF (3.0x10^4 IU). Mice were immunized in the footpad 3X at 7 day intervals and sacrificed 3 – 5 days following the final injection. Spleen and popliteal lymph node cell suspensions were stimulated in vitro with psorelin inactivated NIH-3T3 cells (H-2\(^b\)) permanently transfected with Her-2/neu and B7-1 encoding constructs. Stimulated lymphocytes were incubated with \(^{51}\)Cr labeled targets in the indicated ratios for 5 hours. 3T3 = non-transfected NIH-3T3 cells. Neu+B7 = NIH-3T3 cells transfected with Her-2/neu and B7.1. Data represents groups of 2 mice.
Summary:

Much of the work described for Specific Aim 1 has now been completed. We have finished the in vitro testing and packaging of the replicon constructs described, and have begun work on additional constructs. Progress has been made characterizing the CD8+ T cell response in both wild type and Her-2/neu transgenic mice with generation of consistent CTL activity in the wild type mice. Additionally, we have cloned overlapping fragments of the Her-2/neu gene into a bacterial expression system, and demonstrated a proliferative response to one of the recombinant proteins. These fragments will be used to evaluate CD4+ T helper responses in immunized mice. We believe an immunotherapeutic protocol consisting of the appropriate combination and antigen and cytokine encoding replicons will be useful for overcoming the tolerance associated with the Her-2/neu transgenic mice.

In our initial tumor challenge experiment with replicons encoding Her-2/neu alone, we failed to mediate rejection of transferred Neu expressing tumors in Her-2/neu transgenic mice. Again, we feel it will be a matter of determining the appropriate cytokine encoding constructs for use in our protocol.

References: