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Emergency Interventions After Severe Traumatic Brain Injury in Rats: Effect on Neuropathology and Functional Outcome

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Traumatic brain injury (TBI) contributes to combat morbidity/mortality. Studies in models of TBI have focused on novel mediators and mechanisms. We used controlled cortical impact (CCI), a contemporary model of TBI in rats to study field-oriented treatments. The following technical objectives were addressed: 1) What is the optimal ventilation strategy? 2) Is hypothermia beneficial? and 3) What is the optimal sedative/analggesic? A fourth objective, combining hypothermia plus other therapies was abandoned due to the limited efficacy of hypothermia. The most important findings/publications include: Objective #1) In a report published in the Journal of Neurosurgery, we demonstrated that early aggressive hyperventilation worsened neuronal death. Objective #2) We published the first report showing that hypothermia was ineffective in the combat-relevant scenario of CCI followed by secondary hypoxemia. That work is in press in Critical Care Medicine. Objective #3) We reported remarkably poor outcome in rats treated with narcotics (fentanyl) versus general anesthesia (isoflurane) after CCI. That work was presented at the 1999 meeting of the National Neurotrauma Society by research trainee Dr. Kimberly Statler, who received the Women in Neurotrauma Award. The paper is in press in the Journal of Neurotrauma. Since narcotics are the current field treatment after TBI, we then completed a seven anesthetic study in our CCI model. This is the first comprehensive assessment of the effect of anesthetic agents on outcome in experimental TBI. Notably, agents that are commonly used in the field (narcotics, benzodiazepines, and propofol) had detrimental effects on selective aspects of outcome. The results of this study will be presented at the National Neurotrauma Society meeting this fall.

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(5) INTRODUCTION

Please note that this publication represents a supplement to our final report, which was submitted in January 2000. There are still a number of manuscripts and abstracts that are either in press, in submission or in preparation. In addition to this revised final report, further supplements will follow for subsequent publications.

Traumatic brain injury (TBI) is an important contributor to combat casualty morbidity and mortality. Although progress has been made in the development of rodent models of TBI (such as the controlled cortical impact (CCI) model), most of the studies with these models have focused on molecular mechanisms of damage. Because there has been a paucity of application of practical emergency interventions in TBI models, we felt that it was important to address this deficiency since this could have important implications for field and emergency management of both soldiers and civilians. Our overall hypothesis is that optimal manipulation of practical interventions applicable in the emergency treatment of severe TBI (respiratory management, temperature control, and sedation) can reduce secondary brain injury in a rat model of brain contusion, and thereby improve functional and/or neuropathological outcome.

Funding year 1

In yr. 1 of funding, we addressed the first Technical Objective, namely, to investigate the effects of mechanical ventilation strategies (as applied by the first responder in the field) on functional and neuropathological outcome in our model. We found that aggressive, prophylactic hyperventilation (HV) applied for 4 h immediately after injury is detrimental (vs normal PaCO₂), and leads to increased neuronal death in selectively vulnerable brain regions. This study was published as a full manuscript in the Journal of Neurosurgery (1). The reviewers indicated that this was an important study that would be cited often. Dr. M. Forbes, a Critical Care Medicine fellow training in research with Dr. Kochanek, authored the study.

To set the stage for the evaluation of therapies after injury (as proposed studies in technical objectives 2-4), it appeared that it would be optimal to have TBI models both with and without a secondary insult-- since such insults are common in the field. This was accomplished by two studies assessing our model (2,3), including, adding a 30-min period of moderate hypoxemia to the CCI. Characterization of that model was described in the 1997 report and was presented in 1998 at the National Neurotrauma Society Meeting (3). During yrs. 2 and 3, we used both the CCI model and the CCI plus secondary hypoxemia model to test therapies.

Funding year 2

In yr. 2 we performed three studies addressing Technical Objective 2 and part of Objective 3. These studies included: 1) assessment of the effect of transient (4 h), moderate hypothermia on outcome after TBI with a secondary insult, 2) assessment of the effect of prolonged (12-h) moderate hypothermia on outcome after TBI, and 3) assessment of the effect of anti-excitotoxic therapy (the NMDA-receptor antagonist MK-
The results of these studies showed that hypothermia (32 °C, for either 4 h or 12 h) reduced DNA damage early after injury, but beneficial effects on long-term outcome could not be demonstrated in the model. It was particularly ineffective after the combined CCI plus hypoxemia. In contrast, we were surprised to find that MK-801 improved functional outcome. However, neither treatment improved brain histopathology after injury. Three research fellows (Drs. C. Robertson, M. Whalen, and R. Ruppel) worked on these projects with the PI (Dr. Kochanek) during yr. 2. Dr. Robertson presented an abstract at the 1999 annual meeting of the Society of Critical Care Medicine (4). That work on hypothermia is in press as a full manuscript in the journal Critical Care Medicine (5). We also reported that 4 h of moderate hypothermia attenuates DNA damage after injury (6,7). That work was presented last year at the Society of Critical Care Medicine Meeting and the manuscript is in preparation. Our work on hypothermia in TBI was summarized in an invited review article published by the International Trauma, Anesthesia and Critical Care Medicine Society (ITACCS)(8). Dr. Randall Ruppel presented the work on MK-801 at the 1999 Meeting of the National Neurotrauma Society (9). It was one of 12 papers selected for oral presentation out of over 200 papers submitted. The manuscript is in preparation for submission to the Journal of Neurotrauma.

Funding year 3

During yr. 3, we carried out a comprehensive study of sedation/analgesia comparing a narcotic (fentanyl) to a conventional general anesthetic (isoflurane). Fentanyl and morphine are the most commonly used narcotics after human head injury while isoflurane is the most commonly used anesthetic in rat models. We reported remarkably poor outcome in rats treated with narcotics (fentanyl) versus general anesthesia (isoflurane) after CCI. That study is extremely relevant since narcotics are the current field treatment for combat casualties. That work was presented by Critical Care Medicine fellow research trainee Dr. Kimberly Statler at the 1999 meetings of the National Neurotrauma Society, the Society for Neuroscience, and the Society of Critical Care Medicine (10-12). Dr. Statler received the 1999 Women in Neurotrauma Award at the National Neurotrauma Meeting and an Educational Scholarship from the Society of Critical Care Medicine. The full paper is in press in the Journal of Neurotrauma (13). The lack of beneficial effect of hypothermia in our model coupled with the remarkably powerful effect of isoflurane suggested the elimination of technical objective 4 in favor of a more comprehensive study of sedatives/analgesics early after CCI (i.e., expansion of proposed technical objective 3).

We recently completed work on the final study in this proposal, namely a comparison of 7 sedative/analgesic treatments applied in a field relevant paradigm. This is the first comprehensive assessment of the effect of anesthetic agents on outcome in experimental TBI. This study showed detrimental effects of narcotics, diazepam, and propofol after TBI, and suggested powerful benefits of isoflurane at or near the time of injury. These findings suggest important implications for acute patient care, especially field application, as well as for experimental models of TBI. Dr. Kimberly Statler will present the results of this study at the National Neurotrauma Society meeting this fall.
(6) BODY

(a) Technical Objective 1: Mechanical ventilation strategies after severe TBI in rats (see summary for 1996-1997 [yr-1] and reference 1, both in appendix).

We showed that aggressive, early HV after TBI augments neuronal death in CA3 hippocampus in a rat model of cerebral contusion. The further reduction of CBF with HV during the low cerebral blood flow state immediately after injury coupled with alkalosis may increase the vulnerability of selected neurons to damage. The results of this study reinforce the meticulous attention necessary to prevent secondary injury after TBI. A risk in the use of HV is demonstrated.

Recommendation: Unless signs of impending herniation (unilateral pupillary dilation, hypertension, bradycardia) are present, this study supports the targeting of normocarbia for mechanical ventilation in the emergency stabilization of the brain trauma victim.

(b) Technical Objectives 2: Testing of field-relevant therapies (notably hypothermia) in experimental models of severe TBI (with and without a secondary hypoxemic insult) in rats.

(b1) Effect of transient (4 h), moderate (32°C) hypothermia on functional and histological outcome after experimental TBI with a superimposed secondary hypoxemic insult in rats. (see summary for 1998-1999 [yr-2] and reference 5, both in appendix).

We tested the effect of 4 h of hypothermia in our model of TBI with a 30-min secondary hypoxemic insult. Hypothermia is effective in a variety of experimental models with transient application (1-4 h) and in a single-center study in humans (32°C applied for 24 h). However, in neither experimental nor clinical TBI has hypothermia been tested when applied after the combination of TBI with a secondary insult. We found no benefit from hypothermia after this field-relevant combined insult. This suggests three possibilities. First, the combination of TBI plus a secondary hypoxemic insult may be so severe that no single treatment will be effective. Second, the insult is so severe that no therapies will be effective. Third, based on our work in technical objective 3, it is possible that a beneficial effect of hypothermia is being masked by using isoflurane anesthesia (viva infra). This work is in press as a full paper in the journal Critical Care Medicine (5).

Recommendation: Even in centers where hypothermia was shown to be effective after TBI, this has not been the case for severely injured patients GCS 3-4. It is likely that severe injuries, such as that modeled by CCI with a 30-min hypoxemic secondary insult, will require combination therapies or may be refractory to all therapy. Also, based on the work by our group on technical objective #3, it will be important in future studies to test hypothermia using a sedative/analgesic approach that is similar to that used in the field or
clinic—i.e., with narcotics. To our knowledge, such a study has never been carried out in a rodent model of TBI.


Based on the aforementioned study, in the CCI model, we sought to test, to our knowledge for the first time in any laboratory, the prolonged application of hypothermia in a rodent model of TBI. This included over 13-h of controlled mechanical ventilation and physiological monitoring. To date, only brief 1-4-h applications have been tested. In this study, we examined TBI without a secondary insult. As described in last year's progress report, we failed to observe important beneficial effects of 12-h of hypothermia on functional or histopathological outcome after CCI. This is a surprising finding which is discussed below.

Recommendation: Studies of 12-h of hypothermia in any experimental animal model are very labor intensive. The negative result of this study suggests one of two possibilities. First, there may be both beneficial and deleterious aspects to the use of hypothermia. Despite promising data from single clinical sites, recently, a randomized, controlled multi-center trial of hypothermia in human head injury failed to yield a positive result. Second, once again based on the work by our group on technical objective #3, it will be important in future studies to test hypothermia using a sedative/analgescic approach that is similar to that used in the field or clinic—i.e., with narcotics. It is our recommendation that this be tried first in a rodent model using either morphine or fentanyl anesthesia followed by either 1 or 4-h of hypothermia vs normothermia.

(b3) Effect of the anti-excitotoxic NMDA-receptor antagonist MK-801 on functional and histological outcome after experimental TBI in rats. (see summary for 1998-1999 [yr. 2], in appendix, for detailed methods).

In the third treatment trial in yr. 2, we sought to test the effect of the traditional NMDA-receptor antagonist MK-801 in our TBI model (without secondary insult). The NMDA antagonist MK-801 was effective in improving both motor function and some aspects of cognitive function after CCI. The motor effects were more dramatic than those seen with hypothermia. In a separate pilot study, MK-801 was not effective when tested in our TBI plus secondary hypoxemia model, again suggesting this insult may be too severe for any single therapy. Although this specific agent is not available for clinical use, it suggests that this category of agents—targeting excitotoxicity—represents a viable strategy.

Recommendation: This finding is particularly relevant since our studies addressing technical objective #3 suggested that an anti-excitotoxic general anesthesia strategy such as isoflurane produced a markedly better outcome than treatment with the narcotic analgesic fentanyl. Although there have been several negative clinical trials of anti-
excitotoxic therapy, there is frequently a delay in administration of treatment for as much
as 6-h in these trials. Several reports have suggested that important components of the
excitotoxic response may occur in the initial 1-2-h after injury. Based on our findings,
anti-excitotoxic strategies should not be abandoned, rather consideration should be
given to the field application of these strategies. In addition, sedative/analgesics
with anti-excitotoxic properties must be extensively studied in experimental TBI, in
both small animal and large animal models.

(c) Technical Objective 3: Testing of the optimal field-relevant sedative/analgesic
therapy in an experimental model of severe TBI in rats (see reference 13 in
appendix).

(c1) Comparison of the effects of TBI on functional and histological outcome
after experimental TBI in rats anesthetized with fentanyl or isoflurane (described
below).

Currently, in clinical practice, fentanyl is the most commonly used emergency
sedative for intubated patients with severe TBI. Fentanyl, has little direct anti-excitotoxic
properties. Thus, to begin investigating this area, we tested how fentanyl treatment
compared to standard isoflurane anesthesia in our model.

Outcome protocol

Rats were initially anesthetized with N₂O:O₂ (2:1) and 4% isoflurane and then
endotracheally intubated and mechanically ventilated. Anesthesia was maintained for the
preparatory surgery with 2 - 2.5% isoflurane and N₂O:O₂ (2:1). Pancuronium bromide
(0.1 mg/kg/h) was given iv for muscle relaxation. Femoral venous and arterial vessels
were cannulated for continuous blood pressure measurement, blood sampling, and
administration of medications. A rectal probe was inserted to monitor core temperature.
The rat was then placed in a stereotaxic frame and a left parietal craniotomy was
performed. The dura and bone flap were left in place until immediately before CCI. A
burr hole was drilled into the left frontal bone for temperature probe placement into the
frontal lobe. Continuously monitored physiologic parameters included arterial blood
pressure and rectal and brain temperatures. Blood glucose, hematocrit, and arterial blood
gas samples were assessed every 15 min for the initial hour and every 30 min thereafter.
PaCO₂ was controlled at 35 - 45 mm Hg. This protocol produced a PaO₂ of greater than
70 mm Hg in all preparations. Both brain and rectal temperatures were maintained at
37.0 ± 0.5 °C.

Rats were allowed to stabilize for 5-min after completion of surgical preparation and
then randomized to receive either fentanyl or isoflurane anesthesia. In the fentanyl group
(n=9), isoflurane was discontinued and 10 μg/kg of fentanyl was administered iv,
followed by a continuous iv infusion at 50 μg/kg/h. In the isoflurane group (n=9),
inspired isoflurane concentration was reduced to 1%, and normal saline, the vehicle for
fentanyl-treated rats, was administered to match the volume received by fentanyl
infusion. Both anesthetic groups continue to receive N₂O:O₂ (2:1). After 30-min
equilibration, TBI was induced by CCI. In pilot studies comparing isoflurane and
fentanyl using our standard CCI model (6-mm tip, 4 m/s velocity, 50 msec duration of deformation and 2.5-mm deformation depth), all of the fentanyl-treated rats developed pulmonary edema and died early after injury. Thus, to compare the effect of isoflurane vs fentanyl on long-term outcome in our model, our standard injury was reduced (2.0-mm deformation depth). After CCI, the bone flap was replaced and sealed with dental cement, and the scalp incision was closed. Anesthesia was continued for 4-h in the isoflurane group and 3.5-h in the fentanyl group to facilitate similar extubation times. At the end of the anesthetic period, rats received 100% oxygen, were allowed to awaken and resume spontaneous breathing, and were then extubated and returned to their cages. Sham rats underwent identical preparation/anesthesia, but no CCI (n=6 per group).

Motor function, including beam balance and beam walking tasks, was tested by an observer blinded to group assignment on days 1-5 after injury. Morris Water Maze (MWM) testing was performed using an acquisition paradigm on days 14-20 after injury. Lesion volume and hippocampal neuron survival were assessed on day 21.

**ICP Protocol**

![Graph 1: MAP vs time after injury. MAP in fentanyl-treated injured (open circles) and sham (open triangles) rats was ~50 mm Hg higher than in isoflurane-treated rats at all time points (injured shown by closed circles and shams by closed triangles). * p < 0.05, isoflurane vs fentanyl at each time after injury, § p < 0.05, isoflurane vs fentanyl at all time points, including baseline, in shams.](image)

![Graph 2: Beam walking latency vs d after injury. Isoflurane-treated rats recovered by post-injury d 3, fentanyl-treated rats failed to regain normal function by the end of the 5-d period. * p < 0.05, injured vs sham. Beam balance latency showed similar benefit of isoflurane vs fentanyl.](image)

Based both on results of the above protocol showing that fentanyl-treated rats had higher MAP throughout the experiment and on a recent report that increased MAP may exacerbate injury after TBI, ICP and percent brain water were monitored in a separate cohort of rats (n=9 per anesthetic group) subjected to either fentanyl or isoflurane anesthesia and CCI in an identical paradigm to that used to assess functional outcome.

Surgical preparation, randomization, anesthetic administration, and CCI were identical to the outcome protocol, with minor exceptions. Specifically, an intraparenchymal ICP monitor (Codman microtransducer) was inserted through a burr hole in the frontal bone into the contralateral (right) frontal cortex at the time of craniotomy. After CCI, anesthesia was continued and ICP was monitored for 4-h in both anesthetic groups. Cerebral perfusion pressure (CPP) was calculated as the difference between MAP and ICP. Rats were killed by decapitation at the end of the anesthetic period. Brains were immediately removed and a 3-mm coronal slice was made through the center of the contusion. Per cent brain water was determined in the coronal
slice using the wet-dry weight method. Brain water content was determined in both the injured and the homologous region of the uninjured hemispheres.

As an added control, a separate cohort of rats (n=3) was subjected to CCI and allowed to recover without anesthesia. These rats were prepared for CCI under isoflurane anesthesia as above, allowed to recover a tail-pinch response, and then subjected to CCI. Arterial MAP was monitored via a femoral arterial catheter for 4-h during recovery without anesthesia.

Results

Time to extubation did not differ after injury between isoflurane and fentanyl treatment groups (269 ± 7 min vs 275 ± 15 min, p = 0.29). Physiologic values, including PaCO2, PaO2, blood glucose and Hct did not differ between anesthetic groups. In contrast, MAP was higher in injured rats treated with fentanyl compared to their isoflurane counterparts (p < 0.05) during the entire post-trauma period (Fig 1). Similarly, MAP was higher in shams treated with fentanyl vs isoflurane (p < 0.05) during the entire duration of anesthesia (Fig 1). Fentanyl-treated rats had a MAP of ~150 mm Hg compared to ~105 mm Hg in the isoflurane groups.

Rats anesthetized with isoflurane performed better on beam balance and beam walking tasks after TBI compared to their fentanyl counterparts (p < 0.05, Fig 2). Following injury, isoflurane-anesthetized rats also performed better than their fentanyl-treated counterparts during MWM testing with a hidden platform (p < 0.05, Fig 3). Motor and MWM performances did not differ between sham groups.

Lesion volume, expressed as mm³ or as percent of uninjured hemisphere, at 21-d did not differ between treatment groups (see reference 13 in appendix for details). In contrast, neuron counts in the injured CA1 hippocampus were markedly greater.
in isoflurane-treated rats ($p < 0.05$, Fig 4). Neuron counts in the injured CA3 hippocampus, however, did not differ significantly between treatment groups (see reference 13 in appendix for details).

In the ICP protocol, again physiologic values, including PaCO$_2$, PaO$_2$, glucose and Hct, did not differ between anesthetic groups. As in the outcome protocol, MAP was higher in rats treated with fentanyl compared to their isoflurane counterparts ($p < 0.05$). ICP was similar in both anesthetic groups; however, there was a trend toward higher ICP in rats anesthetized with isoflurane by 3-4-h after TBI (Fig 5). This strongly suggests that the higher MAP in fentanyl vs isoflurane treated rats did not exacerbate intracranial hypertension. As expected from the difference in MAP, CPP was higher in the fentanyl treatment group (Fig 6).

Brain water content, assessed at 4-h after injury, was higher in the injured vs uninjured hemisphere ($p < 0.05$) for both anesthetic groups (Fig 7). However, brain water in either the injured or uninjured hemisphere did not differ between isoflurane- and fentanyl-treated rats, indicating that edema was not exacerbated in the fentanyl group.

Average MAP during the 4-h post-injury observation period did not differ significantly between fentanyl-treated rats and those recovering from CCI without anesthesia (157 ± 6.2 mm Hg vs 147 ± 7.1 mm Hg, NS). In contrast, isoflurane-anesthetized rats had lower MAP (105 ± 5.5 mm Hg) vs both fentanyl-treated rats and rats recovering without anesthesia ($p < 0.05$ vs both groups).

**Recommendation:** The theoretical advantages of isoflurane vs fentanyl are compelling; however, explanations for the observed improvement in neurologic outcome after TBI in isoflurane-anesthetized rats remain to be determined. What has become clearer is that anesthetic agents may have considerable impact upon outcome following TBI. The results of this study suggest two important potential ramifications. First, isoflurane may not represent the optimal anesthetic in experimental TBI since it may mask potential benefits of novel therapies. Second, despite common clinical and field use, narcotics such as morphine or fentanyl may not be the optimal sedative/analgesic agent to administer to patients in the acute phase after severe head injury. We feel this has considerable relevance since narcotics
Figure 7: Percent brain water 4 h after TBI. Percent brain water in the injured hemisphere was increased vs respective non-injured hemisphere in both isoflurane- and fentanyl-treated rats; however, brain water did not differ between anesthetic groups. *p < 0.05; isoflurane vs fentanyl.

(either fentanyl or morphine) are first line agents in field or emergency department. Consideration should also be given to the possibility that agents like isoflurane anesthesia could be provided in the field. Our suspicion was that narcotics are not deleterious, rather general anesthetics such as isoflurane are powerfully beneficial, possibly through anti-excitotoxic actions or cerebral blood flow promotion early after injury. Defining the factors responsible for improved outcome with isoflurane may help to direct the clinical application of more optimal sedative/analgesic agents and possibly to identify novel therapies. Finally, since the immediate post-trauma sedative/analgesic regimen had such a powerful effect on both functional and histopathological outcome, we completed a comprehensive comparison of field-relevant sedative/analgesic strategies (vida infra).

(C2) Randomized, blinded study in the rat model of CCI of seven different sedative/analgesic strategies for field use in TBI.

We have recently completed a nine group (seven anesthetic) study in our CCI model. Rats are prepared for TBI exactly as described in our protocol comparing isoflurane and fentanyl above. Anesthesia for surgical preparations is 2% isoflurane in nitrous oxide/oxygen (2:1). After surgical preparation, anesthesia is discontinued until tail-pinch response is obtained and then CCI is delivered. Rats are then randomized to one of the 8 groups below (n = 9 per group, Table 1). There is also a sham group (thus, a total of 9 groups). The sedation or anesthesia is maintained for a period of 60 min and the rats are then weaned and extubated when recovered. Outcome parameters are identical to the isoflurane vs fentanyl outcome study (motor and MWM function; lesion volume, hippocampal CA1 and CA3 cell counts).

<table>
<thead>
<tr>
<th>Table 1. Sedation/analgesia study posttrauma</th>
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<td><strong>Anesthetic/Sedative</strong></td>
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<tr>
<td>Diazepam</td>
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<td>Fentanyl</td>
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<td>Isoflurane</td>
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<td>Ketamine</td>
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<td>Pentobarbital</td>
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<td>Propofol</td>
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<tr>
<td>None¹</td>
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<td>Sham</td>
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¹Isoflurane anesthesia discontinued and TBI immediately on return of tail pinch reflex.

Comment: First, pilot studies were conducted in our CCI model using each anesthetic. The selected doses were based on those commonly reported in the literature with minor adjustments based on our pilot data. As you are aware, our rat facility became infected with a virus midway through the
study, necessitating elimination of all potentially infected animals. The study was restarted in April 2000. Although the physiologic and functional studies are finished, we are still completing the histologic evaluation of brain tissue.

Results

Physiologic parameters (brain temperature, blood gases, and glucose) did not differ between experimental groups, except for MAP (Fig 8). As seen in our prior study, MAP was higher in fentanyl vs isoflurane anesthetized rats (~135 mm Hg vs ~105 mm Hg, p < 0.05). Although there was a trend toward higher MAP in rats treated with diazepam, morphine, or pentobarbital vs isoflurane, these did not reach statistical significance.

Motor function results showed that some agents had detrimental effects on selected outcome parameters. Rats anesthetized with morphine or propofol performed worse than sham on beam balance testing (p < 0.05, Fig 9). Beam balance performances were similar among all other experimental groups. On beam walking tasks, shams performed better than all other experimental groups (p < 0.05, Fig 10). The longest latencies (poorest performances) were again seen with morphine or propofol anesthesia. Beam walking performances were similar for all other treatment groups. Although there were no significant differences between experimental groups in MWM testing with a submerged platform, rats treated with diazepam or fentanyl had a trend toward worse performance than shams (p < 0.10, Fig 11). Similarly, in the visible platform paradigm, rats treated with fentanyl

Fig 8: MAP vs time after injury. MAP is higher in fentanyl (red) vs isoflurane (yellow) anesthetized rats (p < 0.05).

Fig 9: Beam balance latency vs time after injury. Rats treated with morphine (purple) or propofol (burgundy) performed worse than shams (p < 0.05); however, there were no differences between other treatment groups.

Fig 10: Beam walking latency vs time after injury. Sham rats performed better than all other experimental groups (p < 0.05). There were no significant differences among treatment groups.
or morphine had a trend toward worse performance than sham (p < 0.10); however, there were no significant differences between other experimental groups. Assessment of lesion volume and hippocampal cell counts is ongoing.

**Recommendation:** The results of this multi-sedation study show that rats anesthetized with narcotics, diazepam, or propofol had worse functional outcome than sham. **Surprisingly, rats treated with isoflurane only before injury performed as well as those given isoflurane both before injury and for 1 h post-trauma.** Taken in context of our prior study (see C1) showing improved outcome with isoflurane vs fentanyl initiated *prior to injury,* these data suggest that powerful beneficial effects of isoflurane likely occur at or near the time of impact. Considering their common clinical use, the potential detrimental effects of treatment with narcotics, diazepam, or propofol deserve further characterization. Additionally, the use of isoflurane immediately before injury (despite the fact that rats were allowed to recover to tail pinch before injury) appears to mask the effects of the other sedative agents used in the study.

The dramatic influence of early anesthetic management after TBI in our model suggests important implications for the field management of TBI. Although isoflurane may not represent the most practical field agent, novel medications with similar anti-excitotoxic and/or cerebral blood flow promoting effects may well provide a breakthrough in the clinical management of TBI. Our study clearly demonstrates that the an optimal field anesthetic regimen after severe TBI needs to be defined. Clinical studies to evaluate the effects of different anesthetic strategies immediately after impact (i.e. those applied by a field medic) are needed. In addition, further animal studies are warranted to better define the optimal anesthetic properties early after TBI.

As in initial step, we are currently conducting studies to elucidate the potential neuroprotective mechanisms of isoflurane anesthesia by assessing cerebral glucose utilization, cerebral blood flow, and cerebral calcium accumulation during isoflurane vs fentanyl anesthesia in our CCI model of TBI. Finally, we are comparing anesthetic strategies in our CCI model using fentanyl (in lieu of isoflurane) as the preparatory anesthetic agent. We feel that this approach will more accurately simulate the clinical head injury, both in the military and civilian sectors.
(7) KEY RESEARCH ACCOMPLISHMENTS

In order of importance

Sedation/Anesthesia

- Narcotics, the standard field-treatment of victims of severe head injury (after intubation) and a front line treatment in emergency departments in the civilian sector had not been directly compared with general anesthesia in a contemporary rodent model of TBI. After experimental TBI, rats anesthetized at the time of injury with isoflurane vs fentanyl exhibited markedly better functional and histopathological outcomes. Additionally, it appears that powerful beneficial effects of isoflurane occur at or near the time of impact. Despite current clinical use, narcotics may not be the optimal sedative/analgesic early after TBI. Although isoflurane may not represent the most practical field agent, novel medications with similar anti-excitotoxic and/or cerebral blood flow promoting effects may well provide a breakthrough in the clinical management of TBI.

Hyperventilation

- Aggressive hyperventilation for 4-5 h early after TBI is associated with an exacerbation of hippocampal neuronal death in selectively vulnerable brain regions adjacent to the contusion site.

Hypothermia

- Transient, moderate hypothermia, effective after TBI alone in prior studies, was demonstrated to be ineffective after experimental TBI in rats subjected to TBI with a superimposed secondary insult (hypoxemia). This may be clinically important since hypoxic patients have not been randomized in current clinical trials of hypothermia after TBI.

Mechanisms

- Moderate hypothermia reduces markers of injury (such as DNA damage) early after experimental TBI. However, sustained (12 h) of hypothermia was also surprisingly ineffective (on long-term outcome) after experimental TBI in rats. This suggests that although there are beneficial effects of hypothermia, there are potential side effects.

(8) REPORTABLE OUTCOMES

Manuscripts


§Full manuscripts from abstracts (see below) Whalen et al., and Ruppel et al. are also in preparation.

Abstracts and Presentations


**AWARDS**


(2000) Educational Scholarship to Dr. Kimberly Statler from the Society of Critical Care Medicine for her abstract entitled, “Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats.”

**9) CONCLUSIONS**

1. Based on the important finding in this study where the use of fentanyl in our CCI model produced deleterious effects on outcome after TBI, animal models should utilize clinically relevant sedative/analgesic treatments. The beneficial mechanisms of isoflurane (possibly promotion of cerebral blood flow or reduction of excitotoxicity) should be investigated for the development of novel treatments. Isoflurane anesthesia likely provides powerful beneficial effects at or near the time of injury. Sedative agents with anti-excitotoxic and/or cerebral blood flow promoting actions similar to isoflurane may thus represent better alternatives early after TBI. **Narcotics may not be the optimal sedative/analgesic early after TBI. Better field therapy than narcotics must be developed.**

2. Based on studies in rats using the CCI model, we have demonstrated tangible risk to aggressive, indiscriminate hyperventilation early after injury—specifically—augmentation of neuronal death in selectively vulnerable brain regions. This suggests that aggressive hyperventilation should not be indiscriminately used in the field
treatment of TBI, rather it should be applied if there are signs and/or symptoms of herniation. Mild hyperventilation (used in our control group) or normocapnia may be preferable.

3. Based on our studies in rats, hypothermia, although showing some beneficial effects, particularly early after TBI (such as a reduction in DNA damage, etc), may have some deleterious effects which result in only modest overall beneficial effects on long-term outcome. This is particularly true in the setting of severe injury (such as TBI plus secondary hypoxemic insults) where it is possible that there is little to gain except side effects. Also based on our narcotic (fentanyl) vs isoflurane study and our multi-sedation study, hypothermia should be re-examined in future studies with narcotic anesthesia, since beneficial effects of isoflurane may be masking any benefit from hypothermia.

(10) REFERENCES


(11) APPENDIX

1. 1997 Report

2. 1998 Report

3. Curriculum Vitae

4. Manuscripts:


5. Abstracts:


(12) BINDING (N/A)

(13) FINAL REPORTS
   a) Bibliography of all publications and abstracts (see appendix)
   b) List of personnel receiving pay from the research effort
      Patrick M. Kochanek, M.D.
      Peter Safar, M.D.
      Henry Alexander
      Scott Heineman
      Marc Provis
      Linda Amick
Augmented neuronal death in CA3 hippocampus following hyperventilation early after controlled cortical impact

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Minimizing secondary injury after severe traumatic brain injury (TBI) is the primary goal of cerebral resuscitation. For more than two decades, hyperventilation has been one of the most often used strategies in the management of TBI. Laboratory and clinical studies, however, have verified a post-TBI state of reduced cerebral perfusion that may increase the brain's vulnerability to secondary injury. In addition, it has been suggested in a clinical study that hyperventilation may worsen outcome after TBI.

Object. Using the controlled cortical impact model in rats, the authors tested the hypothesis that aggressive hyperventilation applied immediately after TBI would worsen functional outcome, expand the contusion, and promote neuronal death in selectively vulnerable hippocampal neurons.

Methods. Twenty-six intubated, mechanically ventilated, isoflurane-anesthetized male Sprague–Dawley rats were subjected to controlled cortical impact (4 m/s, 2.5-mm depth of deformation) and randomized after 10 minutes to either hyperventilation (Paco2 = 20.3 ± 0.7 mm Hg) or normal ventilation groups (Paco2 = 34.9 ± 0.3 mm Hg) containing 13 rats apiece and were treated for 5 hours. Beam balance and Morris water maze (MWM) performance latencies were measured in eight rats from each group on Days 1 to 5 and 7 to 11, respectively, after controlled cortical impact. The rats were killed at 14 days postinjury, and serial coronal sections of their brains were studied for contusion volume and hippocampal neuron counting (CA1, CA3) by an observer who was blinded to their treatment group.

Mortality rates were similar in both groups (two of 13 in the normal ventilation compared with three of 13 in the hyperventilation group, not significant [NS]). There were no differences between the groups in mean arterial blood pressure, brain temperature, and serum glucose concentration. There were no differences between groups in performance latencies for both beam balance and MWM or contusion volume (27.8 ± 5.1 mm3 compared with 27.8 ± 3.3 mm3) in the normal ventilation compared with the hyperventilation groups, respectively. In brain sections cut from the center of the contusion, hippocampal neuronal survival in the CA1 region was similar in both groups; however, hyperventilation reduced the number of surviving hippocampal CA3 neurons (29.7 cells/hpf, range 24.2–31.7 in the normal ventilation group compared with 19.9 cells/hpf, range 17–23.7 in the hyperventilation group [25th–75th percentiles]; *p < 0.05, Mann–Whitney rank-sum test).

Conclusions. Aggressive hyperventilation early after TBI augments CA3 hippocampal neuronal death; however, it did not impair functional outcome or expand the contusion. These data indicate that CA3 hippocampal neurons are selectively vulnerable to the effects of hyperventilation after TBI. Further studies delineating the mechanisms underlying these effects are needed, because the injudicious application of hyperventilation early after TBI may contribute to secondary neuronal injury.

KEY WORDS • head injury • hyperventilation • alkalosis • hippocampus • rat

TRAUMATIC brain injury (TBI) is often complicated by malignant intracranial hypertension, which is associated with high mortality rates and has been managed using a combination of therapies including osmotherapy, diuretics, sedation, neuromuscular blockade, optimization of cerebral perfusion pressure, and hyperventilation.6,12,32,38,51 Hyperventilation therapy has been an integral part of the clinical armamentarium in the management of severe TBI for more than 20 years;10 this therapy rapidly reduces cerebral blood flow (CBF) and cerebral blood volume in areas of the brain with intact CO2 autoregulation, providing one option in the management of TBI complicated by malignant intracranial hypertension.13,42

In recent studies, however, researchers have defined a state of reduced CBF early after TBI in humans,31 and animals,5,20,25,46,56,57 particularly in the first 8 hours after TBI. Some authors have hypothesized that the brain is more

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vulnerable to secondary injury during this period and that additional reduction of CBF by hyperventilation may attenuate the delivery of important energy substrates.\textsuperscript{5,7,11,30,39,47,48} Yoshida and Marmarou\textsuperscript{49} reported that hyperventilation produced relative ischemia in cat brain after fluid-perfusion injury and demonstrated an increase in brain lactate and inhibition of recovery of the ratio of phospho
creatine to inorganic phosphate. Muizelaar et al.\textsuperscript{45} also demonstrated a loss of brain interstitial bicarbonate buffer after sustained prophyacetic hyperventilation in rabbits. It has been reported that hyperventilation after TBI in animals and humans can reduce CBF to what traditionally have been considered ischemic levels.\textsuperscript{10,24,42} However, defining the ischemic threshold in injured tissue is problematic.\textsuperscript{21,23} Muizelaar et al.\textsuperscript{39} reported that prolonged hyperventilation after TBI in humans may worsen functional outcome, raising questions regarding the appropriate indications and timing for the optimum application of hyperventilation after TBI. Recently published guidelines for the management of severe head injury\textsuperscript{6} state that "in the absence of intracranial hypertension, hyperventilation (PaCO\textsubscript{2} < 35 mm Hg) therapy should be avoided during the first 24 hours after severe TBI . . . ." although "hyperventilation therapy may be necessary for brief periods where there is acute neurologic deterioration. . . ." Consistent with these guidelines, in the setting of acute neurologic deterioration, aggressive hyperventilation is used by both emergency and critical care personnel. In addition, in the initial stabilization of the brain-injured patient, aggressive hyperventilation (appropriate in the setting of impending herniation, or iatrogenic) occasionally occurs in both the prehospital and acute care settings. The specific impact of hyperventilation during this early low-flow period remains to be determined. Despite the availability of well-characterized rodent models of TBI, which reproduce the early posttraumatic reduction in CBF, the effect of aggressive hyperventilation on histopathological and functional outcome has not, to our knowledge, been investigated.

Using a rat model of focal percussive contusion, we hypothesized that aggressive hyperventilation, beginning immediately after TBI and continuing for 5 hours, would worsen functional outcome, expand the contusion, and promote neuronal death in selectively vulnerable hippocampal neurons.

Materials and Methods

Animals and Study Groups

All experimental protocols used in this report were approved by the Animal Care and Use Committee of the University of Pittsburgh. Twenty-six virus-free Sprague–Dawley rats weighing 346 ± 5 g were studied. Food and water were continuously available in their home cages. After TBI the rats were randomly assigned to one of two groups of 13 animals, one receiving normal ventilation (PaCO\textsubscript{2} = 30–40 mm Hg) and one receiving hyperventilation (PaCO\textsubscript{2} = 15–25 mm Hg).

Surgery and Brain Trauma Model

Anesthesia was induced using 4% isoflurane in N\textsubscript{2}O/O\textsubscript{2} (2:1). The rats were endotracheally intubated and mechanically ventilated. The isoflurane concentration was reduced to 2% followed by sterile surgical placement of a femoral arterial catheter for continuous mean arterial blood pressure (MAPB) and arterial blood gas monitoring. Intramuscular injections of penicillin (100,000 U) and gentamicin (10 mg/kg) were given to minimize the risk of infection. Pancuronium bromide was administered at dosages of 0.1 mg/kg/hour via the arterial line to control ventilation. The rats' core temperature was monitored using a rectal probe.

After stereotactically guided head positioning, an incision was made and the scalp was retracted, exposing the left parietal bone. A craniotomy was made using a high-speed dental drill aided by a binocular operating microscope. A burr hole was made 5 mm anterior and 2 mm lateral to the bregma in the left side of the skull and a temperature probe (0.009-in outer diameter) was inserted through the burr hole and 2 mm into the left parietal cortex. The bone flap was left in place and the isoflurane was reduced to 1% followed by a 15-minute equilibration period. The brain temperature was maintained at 37 ± 0.5°C. Normal arterial blood gas levels were achieved in all rats and PaO\textsubscript{2} was maintained at greater than 70 mm Hg.

The TBI was produced using a controlled cortical impact device as recently described\textsuperscript{26} with minor modifications. Fifteen minutes before controlled cortical impact, an arterial catheter and sample were obtained for measurement of arterial blood gas levels, glucose concentration, and hemocrit. The bone flap was then removed and a vertical controlled cortical impact (4 m/second impactor velocity, 2.5-mm deformation depth) was delivered onto the exposed dura overlaying the left parietal cortex. The bone flap was replaced and sealed with dental cement and the scalp was sutured.

Study Design

The study protocol was designed to mimic the aggressive use of hyperventilation (as opposed to normal ventilation) in the immediate posttrauma period in the prehospital as well as early hospital setting. Ten minutes after controlled cortical impact, rats were randomized to either the normal ventilation group (13 animals, PaCO\textsubscript{2} range 30–40 mm Hg) or the hyperventilation group (13 animals, PaCO\textsubscript{2} range 15–25 mm Hg). The ventilator was adjusted to maintain P\textsubscript{acO\textsubscript{2}} or hyperventilation for 5 hours after controlled cortical impact. Arterial blood gas readings were obtained at 30 minutes post–controlled cortical impact, then hourly. The MAPB was recorded every 30 minutes after controlled cortical impact. Brain and rectal temperatures were recorded every 15 minutes.

At 5 hours after controlled cortical impact, anesthesia was discontinued. Temperature probes and the femoral artery catheter were removed and the rat was weaned from mechanical ventilation in the course of 1 hour and underwent extubation. The time to extubation was recorded. After extubation, supplemental O\textsubscript{2} was administered for 30 minutes. When it had fully recovered, the rat was returned to its cage with full access to food and water.

Functional Outcome and Behavior Assessment

Beam Balance. Vestibulomotor function was tested using the beam balance test\textsuperscript{21} in eight rats from each group. One hour before surgery, the rat was placed lengthwise on a 1.5-cm-wide beam suspended above the ground. The time the rat remained on the beam was recorded (up to 60 seconds). The rat was then removed from the beam and the procedure was repeated. Rats were considered trained when they remained on the beam for three consecutive periods of 60 seconds. Beam balance tests were also performed daily on Days 1 to 5 postinjury. Three trials were recorded and averaged each day for each rat.

Morris Water Maze. Cognitive function was tested in the same eight rats from each group using a standard variation of the Morris water maze (MWM) paradigm.\textsuperscript{5,8} A pool 180 cm in diameter and 60 cm deep was painted black and filled with water to a depth of 28 cm. A clear Plexiglas platform 10 cm in diameter and 26 cm high (2 cm below the water surface) was used as the hidden platform. The pool was located in a 2.5 × 2.5-m room with numerous extra-maze cues (for example, posters, pipes, bookcase) that remained constant throughout the experiment. Testing started 7 days after controlled cortical impact to avoid confounding effects of motor deficits. The rats underwent four trials per day for 5 consecutive days to assess spatial memory performance. The rats started each trial once from each of the four possible start locations.
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| TABLE 1 |
| Physiological values in two groups of rats treated with hyperventilation or normal ventilation after TBI* |
|---------------------------------|------------------|---------------------|------------------|---------------------|
|                                  | Baseline | Postrandomization | Baseline | Postrandomization |
| pH                              | 7.39 ± 0.01 | 7.37 ± 0.01        | 7.38 ± 0.01 | 7.53 ± 0.01†     |
| PaCO₂ (mm Hg)                   | 36.7 ± 1.1   | 34.9 ± 0.3         | 37.2 ± 0.9   | 20.3 ± 0.7†      |
| PaO₂ (mm Hg)                    | 165 ± 6      | 167 ± 4            | 168 ± 4      | 180 ± 3†         |
| base deficit (mmol/L)           | 2.7 ± 3.4    | 4.2 ± 0.7          | -0.6 ± 0.9   | 4.8 ± 0.6        |
| serum glucose (mg%)             | 189 ± 9      | 174 ± 6            | 158 ± 10     | 152 ± 9          |
| hct (%)                         | 36 ± 2.3     | 35 ± 0.6           | 32.3 ± 1.5   | 35 ± 0.6         |
| time to extubate (min)          | NA           | 28 ± 6             | NA            | 29 ± 5           |
| brain temperature (°C)          | 36.7 ± 0.1   | 37 ± 0             | 36.5 ± 0.1   | 37 ± 0           |
| rectal temperature (°C)         | 36.5 ± 0.6   | 37 ± 0             | 37.1 ± 0.1   | 37.1 ± 0.1       |
| MABP (mm Hg)                    | 129 ± 4      | 123 ± 4            | 129 ± 8      | 128 ± 3          |

* All values are expressed as mean ± SEM. Abbreviations: hct = hematocrit; NA = not applicable.
† p < 0.05 at 30 minutes postrandomization compared with baseline.

Histopathological Studies

At 14 days after controlled cortical impact (after completion of all of the functional outcome testing), the rats were anesthetized with 5% isoflurane and killed by perfusion fixation using 10% buffered formalin. Their brains were removed and postfixed at 4°C for a minimum of 1 week, and then cryoprotected in sucrose and cut with a cryotome into 10-μm coronal sections at 1-mm increments from the occipital to the frontal lobes and stained with Cresyl violet.

Contusion Volume. We used a computerized image analysis system to outline the margin of the contusion and the sectional area of the contusion at each 1-mm increment was calculated by an observer (M.L.F.) who was blinded to the treatment group. Contusion volume in each rat was calculated as the sum of these sections.

Hippocampal Cell Counting. Neuronal loss in hippocampal regions CA1 and CA3 pyramidal layers was quantified. A coronal section cut from the dorsal hippocampus underlying the area of contusion, approximately 2.6 mm posterior to the bregma, was used for analysis in each rat. The regions were visualized at × 100 magnification, then localized and counted at × 400 by an observer (R.S.B.C.) blinded to treatment group. Only complete cells with a clearly defined body and nucleus were counted. Surviving pyramidal CA1 and CA3 hippocampal neurons were counted in six separate × 400 fields for each region in both hemispheres. Sections were excluded if the boundary of the contusion extended into the pyramidal layers of the hippocampus or if fixation artifacts precluded accurate counting. Data are reported as the average number of surviving neurons per high-power field for the CA1 and CA3 hippocampal regions in both the ipsilateral and contralateral hemispheres.

Statistical Analysis

Survival was compared between groups using Fisher’s exact test. Between group comparisons of physiological parameters, beam balance, and MWM latencies were made using one- or two-way analysis of variance (ANOVA) for repeated measures where appropriate and post-hoc tests with appropriate correction for multiple comparisons. Contusion volume was normally distributed and was compared between groups using Student’s t-test. Hippocampal neuronal survival in CA1 and CA3 was not normally distributed and was compared between groups using the Mann-Whitney rank-sum test. Significance was defined at a probability level of less than 0.05.

Sources of Supplies and Equipment

Pancuronium bromide and gentamicin were purchased from Elkins-Sinn, Cherry Hill, NJ, and penicillin was acquired from Upjohn, Kalamazoo, MI. The stereotactic head positioning system was obtained from David Kopf, Tujunga, CA. The temperature probe was purchased from Physitemp Corp., Clifton, NJ. The video tracking system (Poly-Trak) was acquired from San Diego Instrument, Inc., San Diego, CA, and the image analysis system (MCID) was from Imaging Research, St. Catharines, Ontario, Canada.

Results

Physiological Parameters

Baseline and 30-minute postrandomization physiological data are presented for all measured parameters in Table 1. After randomization, there was a marked increase in pH and decrease in PaCO₂ in the hyperventilation group (compared with baseline, p < 0.05). Hyperventilation was also associated with a small increase (12 mm Hg) in PaO₂ compared with baseline (p < 0.05). This difference was attributable to the increased minute ventilation and mean airway pressure in the hyperventilation group. At no time were any of the rats hypoxemic (PaO₂ < 100 mm Hg). The entire time course of PaCO₂, arterial pH, MABP, and brain temperature after TBI is given for both groups in Fig. 1. The PaCO₂ and pH levels differed between groups at all time points after randomization (p < 0.05). The MABP and brain temperature were similar in both groups.

Five of 26 rats died during the 14-day study, with all deaths occurring on the day of injury. Two rats remained unresponsive postinjury and were unable to demonstrate any spontaneous respiratory effort for 1 hour after discontinuation of anesthesia and were therefore killed. Three rats developed pulmonary edema and/or respiratory distress and died soon after extubation. There were no differences in mortality between groups (two of 13 in the normal ventilation group compared with three of 13 in the hyperventilation group). There were no differences between groups in time to extubation (Table 1).
Fig. 1. Graphs showing time course of (A) PaCO₂ (mm Hg), (B) arterial pH, (C) MABP (mm Hg), and (D) brain temperature (°C) in all rats treated with either normal ventilation (triangles w/ solid line, 13 animals) or hyperventilation (squares w/ broken line, 13 animals) after controlled cortical impact. *p < 0.05 for normal ventilation compared with hyperventilation. Data are expressed as the mean ± standard error of the mean (SEM).

**Functional Outcome Assessment**

**Beam Balance.** There was no difference between groups in motor performance latencies over time (F₁₈ = 0.17, p < 0.69, Fig. 2). Maximum impairment of performance occurred on Days 1 or 2 in both groups, and eventually returned to baseline. Beam balance performance did not differ significantly between normal ventilation and hyperventilation groups.

**Morris Water Maze.** There was no difference between normal ventilation and hyperventilation groups in the time needed to find the hidden platform in the MWM test (F₁₈ = 0.50, p < 0.50, Fig. 3). In addition, there was a statistically nonsignificant tendency (t₁₈ = 1.77, p < 0.065) for the rats in the hyperventilation group to swim slower than the rats in the normal ventilation group (30.8 ± 1.0 compared with 35.4 ± 2.1 cm/second).

**Histopathological Studies**

**Contusion Volume.** At the injury level selected for this study, the contusion was generally restricted to the parietal cortex beneath the impact site. Contusion volume in both groups is shown in Fig. 4. There was no difference between groups (27.8 ± 5.1 mm³ in the normal ventilation group compared with 27.8 ± 3.3 mm³ in the hyperventilation group) in this outcome parameter.

**Hippocampal Cell Counting.** Figure 5 shows the number of surviving neurons/hpf in the CA1 and CA3 regions of the dorsal hippocampus ipsilateral to the contusion. There were no differences in the number of surviving CA1 hippocampal neurons between groups after controlled cortical impact. There was, however, a further reduction in the number of surviving CA3 neurons in the hyperventilation group after controlled cortical impact compared with the normal ventilation group (normal ventilation 29.7, range 24.2–31.7 neurons/hpf, compared with hyperventilation 19.9, range 17–23.7 neurons/hpf; median [25th–75th percentiles], p < 0.05). Neuronal cell counts in the CA1 and CA3 regions of the hemisphere contralateral to the contusion did not differ in either the normal ventilation or hyperventilation groups (CA1 counts = 55.3, range 52.1–59.1).
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Fig. 3. Graph showing MWM performance latency to find a hidden platform (mean ± SEM, in seconds) by rats on Days 7 to 11 after controlled cortical impact. There was no difference between groups (triangles = normal ventilation [eight animals]; squares = hyperventilation [eight animals]) when performances were compared using ANOVA with repeated measures.

Fig. 4. Bar graph depicting mean lesion area (left y-axis, mm²) compared with distance from occiput (mm) measured 14 days after controlled cortical impact (open bars, normal ventilation [11 rats]; closed bars, hyperventilation [10 rats]). Contusion volume (mm³) was calculated as the sum of these areas in each group and is depicted as the cumulative volume (right y-axis) in the normal ventilation (triangles) and hyperventilation (squares) groups. There was no difference between groups in contusion volume (normal ventilation, 27.8 ± 5.1 mm³ compared with hyperventilation, 27.8 ± 3.1 mm³, mean ± SEM).

[Graph showing MWM performance latency to find a hidden platform (mean ± SEM, in seconds) by rats on Days 7 to 11 after controlled cortical impact. There was no difference between groups (triangles = normal ventilation [eight animals]; squares = hyperventilation [eight animals]) when performances were compared using ANOVA with repeated measures.]

[Bar graph depicting mean lesion area (left y-axis, mm²) compared with distance from occiput (mm) measured 14 days after controlled cortical impact (open bars, normal ventilation [11 rats]; closed bars, hyperventilation [10 rats]). Contusion volume (mm³) was calculated as the sum of these areas in each group and is depicted as the cumulative volume (right y-axis) in the normal ventilation (triangles) and hyperventilation (squares) groups. There was no difference between groups in contusion volume (normal ventilation, 27.8 ± 5.1 mm³ compared with hyperventilation, 27.8 ± 3.1 mm³, mean ± SEM).]

Discussion

In a model of controlled cortical impact–induced focal contusion in rats, aggressive hyperventilation for 5 hours after TBI augments neuronal death in the CA3 region of the hippocampus ipsilateral to the contusion. However, hyperventilation did not worsen motor function or cognitive outcome, as assessed using standard beam balance and MWM paradigms, respectively, and did not increase contusion volume.

Hippocampal CA3 neurons are selectively vulnerable to delayed neuronal death after TBI.2,8,19,49,52,55 Theories about the mechanisms underlying this process remain speculative. Potential mechanisms include ischemia, TBI-induced excitotoxicity, apoptosis, and inflammation.8,19,49

Yamakami and McIntosh16,57 reported reduced CBF as early as 15 and 30 minutes after TBI. Using a piglet model of TBI, Pfenninger, et al.,49 reported CBF reduction as early as 5 minutes post-TBI. Some flow levels were in the range consistent with ischemia. We have previously demonstrated that the hippocampus and cortex ipsilateral to the impact show marked flow reduction (at least 60%) at 2 hours after TBI in the controlled cortical impact model.25 Cerebral blood flow approaches ischemic levels in the core of the contusion at 2 hours postinjury. Although we have not evaluated the reactivity status of the cerebral circulation to changes in PaCO₂ at 2 hours after TBI in this model, we have reported that CO₂ reactivity is impaired, although still present (62–71% of baseline) in and around the contusion at 24 hours after controlled cortical impact in rats.16

Hyperventilation rapidly reduces cerebral blood volume and intracranial pressure (ICP).11 In some studies, this intervention has been associated with CBF values consistent with ischemia or brain tissue hypoxia.10,11,14,43,46 After global cerebral ischemia in dogs, hyperventilation did not increase neuronal death;55 however, the brains were assessed at 8 hours after reperfusion, and neuronal death may be delayed. Although ischemia may be considered a contributing mechanism in the observed augmented neuronal death, ischemia alone is an inadequate explanation for our findings in light of the preservation of CA1 neurons. Although CA1 neurons are known to be selectively vulnerable to ischemic injury,23 they were not affected by hyperventilation in this paradigm. Furthermore, in our model, CA1 neurons are more proximal to the point of impact in the cortex compared with CA3 neurons. The lack of CA1 neuronal death in light of ischemic and (presumed) anatomical vulnerability weighs against ischemia and primary injury as putative mechanisms of neuronal death in the hippocampus in this model. One limitation in this study is that neuronal counting using traditional histological methods may underestimate cell loss because of a loss of hippocampal volume.52 We did not use stereological methods in this study. However, CA1 neuronal counts did not differ between groups and were equivalent to those observed in sham-injured animals studied in our laboratory in prior published46 and unpublished work. In addition, comparisons were only made between injured groups within this study.

Hyperventilation produces cerebral vasoconstriction.

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and alkalosis.\textsuperscript{40} Alkalosis exacerbates N-methyl-D-aspartate receptor–mediated neurotoxicity.\textsuperscript{17,18,21,40} As a result of aggressive hyperventilation, the rats in our study were quite alkaloic as indicated by arterial pH measurements. Although we did not measure brain pH, a decrease in PaCO$_2$ immediately reduces brain interstitial pH.\textsuperscript{40} Although alkalosis appears to have deleterious effects on neurons, acidosis has been shown to have both beneficial and detrimental effects. Giffard, et al.,\textsuperscript{17} and Takadera, et al.,\textsuperscript{44} reported a neuroprotective effect of acidosis via an attenuation of the N-methyl-D-aspartate receptor activation in vitro. Rosner and Becker\textsuperscript{46} reported a deleterious effect of tissue acidosis after experimental TBI in cats. The spatial distribution of brain pH around the contusion and in the hippocampus has not been determined for either normal ventilation or hyperventilation conditions in our model.

Finally, the potential effects of hyperventilation on other mechanisms such as posttraumatic seizures or axonal injury may contribute to the enhanced vulnerability of CA3 neurons. The lateralization of the deleterious effects also raises the possibility that spreading wave depression may be a component of the neurotoxic milieu after TBI in this model of focal contusion.\textsuperscript{29} It could also be the case that the combined effect of alkalosis and further flow reduction by hyperventilation is deleterious in regions vulnerable to excitotoxicity such as CA3. Early, aggressive, or prophylactic hyperventilation, therefore, in the context of reduced CBF, may potentiate excitotoxic mechanisms and augment neuronal death.

Aggressive hyperventilation in the early low-flow period did not worsen functional outcome or expand the contusion, failing to support a significant portion of our initial hypothesis. Ultimate contusion size, in controlled cortical impact or other models of focal contusion, is relatively refractory to manipulation by a variety of interventions;\textsuperscript{38} however, application of hypothermia, particularly prior to injury, reduces contusion volume resulting from controlled cortical impact and lateral fluid-percussion injury.\textsuperscript{13,14} Although we chose rather aggressive hyperventilation in an attempt to produce a maximum effect, we did not test the effect of hyperventilation on a milder contusion, which may be more manipulable to secondary insults. The contusion penumbra has not been clearly defined in either of the standard rodent TBI models (controlled cortical impact or fluid-percussion) for any level of injury. It is possible that selectively vulnerable CA3 hippocampal neurons are the only potential target for a deleterious effect of hyperventilation in our model. However, the effect of hyperventilation on the survival of neurons in the dentate gyrus or hilus (all vulnerable to TBI)\textsuperscript{9,20} was not assessed.

Hippocampal damage and memory deficits are common after TBI in humans.\textsuperscript{26,28} This study did not reveal any added effect of hyperventilation on functional outcome deficits as measured by beam balance and MWM latencies. A number of factors may have contributed to this. Our sample size may have limited statistical power; however, this sample size was adequate to detect the exacerbation of functional deficits by the addition of 30 minutes of moderate hypoxemia (PaO$_2$ 40 mm Hg) in our model.\textsuperscript{8} Second, the cognitive deficits in this model are modest compared with those detailed in previous reports.\textsuperscript{15} Bilateral hippocampal damage may be necessary to create more marked functional deficits.\textsuperscript{30,37} In addition, CA3 damage may not mediate post-TBI memory deficits, as manifested in MWM test results. Finally, the specific functional outcome paradigm may not have the necessary sensitivity to detect subtle functional deficits. For example, more demanding MWM paradigms have been used by other investigators.\textsuperscript{27,35} However, in support of the testing strategy used, our hypothesis was that hyperventilation would worsen functional deficits.

This study does not completely address the uncommon situation in which, soon after severe head injury, marked intracranial hypertension is observed. Hyperventilation may in fact be life saving in its ability to impede herniation. Similarly, we did not measure ICP or titrate ventilation to control cerebral perfusion pressure, and we evaluated only one level of hyperventilation and injury severity. We did not attempt to model the clinical scenario of optimum titration of ventilation when ICP is increased. In the clinical setting, some investigators have demonstrated a wide variety of beneficial effects of hyperventilation under those conditions, such as homogenization of CBF, normalization of cerebral glucose uptake, and improvement in autoregulation.\textsuperscript{12,41} Rather, we chose the worst-case scenario, aggressive hyperventilation during the early posttrauma period when flow is already low and excitotoxicity is peaking.\textsuperscript{43} However, our study does show that hyperventilation is associated with a tangible risk to vulnerable neurons in the controlled cortical impact model. To our knowledge, this is the first in vivo study demonstrating that hyperventilation can augment neuronal injury after TBI, suggesting that there is indeed a tradeoff associated with this intervention.

**Conclusions**

We have demonstrated that aggressive, early hyperven-
Augmented neuronal death following hyperventilation post-TBI

tilation after TBI augments neuronal death in CA3 hippocampus. The further reduction of CBF with hyperventilation during the low CBF state immediately after severe TBI, coupled with alkalosis, may increase the vulnerability of selected neurons to traumatic injury. Further studies are needed to delineate the relative contributions of these mechanisms to the observed effects. The results of this study reinforce that meticulous attention is necessary to prevent secondary injury after TBI, and a risk in the use of hyperventilation is demonstrated.

Acknowledgments

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Therapeutic Hypothermia After Traumatic Brain Injury or Hemorrhagic Shock:

From Mild Cooling to Suspended Animation

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Objectives:

1. To familiarize the reader with contemporary studies on the application of resuscitative hypothermia in the treatment of traumatic brain injury and hemorrhagic shock.

2. To describe the potential mechanisms for the beneficial effects of hypothermia in these settings.

3. To present some recent findings from both laboratory and clinical studies of resuscitative hypothermia conducted at the University of Pittsburgh.

4. To discuss possible side effects and limitations of the application of therapeutic hypothermia.

5. To discuss future directions for novel applications of hypothermia in combination with pharmacologic interventions.

Historical Perspective

One of the earliest reports of the potential beneficial effects of hypothermia in the treatment of traumatic brain injury was described by Charles Phelps in 1897 in his classic textbook *Traumatic injuries of the brain and its membranes* (1). It is fitting that this monograph was assembled on the 100th anniversary of this remarkable description.

"The shaving of the head, which had been advised as a means of facilitating diagnosis, is at the same time a measure of treatment... The essential advantage... to be derived from this procedure is that it permits the effective application of the ice-cap, which next to trephination, ... is most nearly a directly curative
resource... It is contraindicated in hemorrhages and cerebral lacerations when uncomplicated by serious contusion; but, as those lesions are constantly thus complicated, it may be held a proper resort when such symptoms are manifest, without regard to exact diagnosis.”

In the early 1940s, Fay (2,3) examined the deliberate application of hypothermia in traumatic brain injury, and this was followed by several additional series of case reports and uncontrolled trials between 1943 and 1979 by other pioneers in this field including Woringen et al (4), Sedzimir (5), Lazorhes and Campan (6), and Rosomoff (7) in traumatic brain injury, Albin et al (8) in spinal cord injury, Bigelow et al (9) and Swan et al (10) in cardiothoracic surgery, Rosomoff et al (11) in focal cerebral ischemia, Siebke et al (12) and Conn et al (13) in near drowning, Wolfe (14), Benson et al (15), Ravitch and Safar (16) in cardiopulmonary arrest, and Rush et al (17) in the application of deep hypothermia for total circulatory arrest. Although remarkable effects were suggested in many of these reports, they failed to demonstrate convincingly that hypothermia was beneficial and did not result in the widespread application of resuscitative hypothermia. These reports were complicated by a number of difficulties including variation in depth and duration of hypothermia, and failure to include concurrent normothermic controls. In addition, reports of potential infectious complications in patients treated with the sustained application of moderate hypothermia (18) tempered enthusiasm for further studying resuscitative hypothermia in a controlled fashion.
Laboratory studies supporting the application of therapeutic hypothermia in traumatic brain injury and hemorrhagic shock

In the mid 1980s there was renewed interest in the laboratory investigation of the deliberate application of therapeutic hypothermia for protection (induced before the insult) or resuscitation (induced after the insult). This work was focused predominantly in models of global cerebral ischemia in rats and monkeys (19-23), cardiopulmonary arrest (24-28) and near drowning in dogs (29). Central to this resurgence in interest in hypothermia was the development of three novel concepts: 1) that remarkably mild hypothermia (a temperature reduction of between 3° and 5°C) was effective in reducing secondary brain damage (19-30), 2) that the duration of mild hypothermia necessary for a beneficial effect might be transient --as short as 1 or 2 hours (19,28) and 3) that brain temperature, not body temperature, was the critical therapeutic target (19). The chance discovery of the efficacy of mild, transient hypothermia in these studies revived the importance of hypothermia research because mild and transient hypothermia are safer and easier to induce than the previously tried moderate, sustained hypothermia. It is important to define the approximate temperature ranges commonly used to describe specific depths of therapeutic hypothermia. Generally accepted definitions of these ranges are mild (34° to 36°C), moderate (28° to 32°C), deep (15° to 25°C), and profound (<15°C) hypothermia (31).

Specific investigation of the application of therapeutic hypothermia in the treatment of traumatic brain injury was renewed by the report of Clifton et al (32) who observed an
inverse correlation between functional outcome and brain temperature (between 30° and 40°C). This was followed by a series of reports from several laboratories further defining the beneficial effect of hypothermia in a wide variety of models (both rodent and canine) of traumatic brain injury (33-37).

Recent controlled laboratory studies of the utility of resuscitative hypothermia in models of hemorrhagic shock developed from the initial work of Crippen et al in our center (38) and of Meyer and Horton (39). This resuscitative effect was demonstrated in models of both controlled (38,40) and uncontrolled (41) hemorrhagic shock (Figure 1), and with both mild and moderate hypothermia (42,43).

In controlled laboratory studies addressing an additional hemorrhagic shock-related application of deliberate hypothermia, Tisherman et al (44,45) investigated the application of deep and profound hypothermic circulatory arrest to enable resuscitative surgery that would otherwise be impossible. Our series of studies into “suspended animation” has culminated so far in the study by Capone et al (46) who reported complete recovery of the brain in dogs after normothermic hemorrhagic shock of 1 hour followed by profound hypothermic circulatory arrest of 1 hour. This application of resuscitative hypothermia is being further developed as a possible novel therapeutic approach to the management of pulseless battlefield casualties, specifically, “suspended animation” for transport and repair of otherwise lethal extracranial wounds. “Suspended animation” could be induced and reversed by portable cardiopulmonary bypass (47) and followed by subsequent delayed resuscitation (48).
Why hypothermia: Proposed mechanisms for the beneficial effects of deliberate hypothermia in traumatic brain injury and hemorrhagic shock

Laboratory and clinical trials in cerebral resuscitation from ischemic or traumatic brain injury have repeatedly highlighted the tremendous challenge involved in demonstrating reproducible efficacy, in a wide variety of injury models or injury types, when a single therapeutic agent is used (49-51). The complex, multifactorial nature of the cascades of secondary damage purported to occur in both ischemic and traumatic brain injury strongly suggests the need for multimodal therapies (23,48-52). A similar multifactorial pathogenesis is proposed in the evolution of visceral damage after hemorrhagic shock (31). A great deal of evidence suggests that hypothermia favorably and simultaneously influences a large number of secondary injury mechanisms including; energy failure (53), oxidant injury (54,55), delayed neuronal death (19,36), excitotoxicity (56), intracranial hypertension (37) edema formation (35,57), cytoskeletal protein degradation (58), blood-brain barrier permeability (59), IL-1β production (60) (Figure 2), and neutrophil accumulation (61). It is very likely that some critical combination of beneficial effects on these mechanisms is responsible for the success of therapeutic hypothermia in experimental and clinical trials.

Clinical investigation of therapeutic hypothermia in traumatic brain injury

Although there is a much larger body of laboratory data supporting the use of mild, transient, resuscitative hypothermia in ischemic rather than traumatic brain injury,
clinical application of deliberate hypothermia has been spearheaded in controlled trials after traumatic brain injury. Uncontrolled trials of moderate hypothermia in patients after traumatic brain injury looked promising (57,62) but were abandoned because of management problems. Marion et al (63) reported a beneficial effect of moderate (32°C), transient (24 hours) hypothermia on intracranial hypertension in adults with severe closed head injury. A reduction in the need for other therapies for control of intracranial hypertension was observed. Clifton et al (64) reported a reduction in the incidence of posttraumatic seizures in adults treated with moderate hypothermia for 48 hours after severe head injury. A trend toward improved outcome was also observed. Similarly, Shiozaki et al (65) reported efficacy of mild hypothermia in controlling refractory intracranial hypertension in patients with severe traumatic brain injury. Most recently, Marion et al (66) demonstrated that moderate (32°C), transient (24 hours) hypothermia improved functional outcome as measured with the Glasgow outcome scale at 6 months after severe traumatic brain injury in 82 patients randomized to either hypothermia or normothermia. This beneficial effect extended to 12 months in the subgroup of patients with admission Glasgow coma score of 5 to 7 (Table 1). In addition, reductions in IL-1β and glutamate concentrations were demonstrated in cerebrospinal fluid samples from hypothermic vs normothermic patients, suggesting the possibility of beneficial effects of hypothermia on posttraumatic inflammation and excitotoxicity, respectively. Remarkably, a significant reduction in cerebral metabolic rate for oxygen was not observed (63,66), suggesting that this beneficial effect was not
due to a simple reduction in cerebral oxidative metabolic demands. A multicenter randomized controlled clinical trial of 48 hours of hypothermia vs normothermia in the treatment of human head injury is currently underway.

Potential limitations and complications of the application of deliberate hypothermia

Hypothermia is associated with potentially limiting side effects. Suppression of acute inflammation (67) and an increased infection risk (18) are concerns. These complications appear to be importantly related to the duration of hypothermia and the underlying condition that is being treated. In traumatic brain injury, Marion et al (66) and Clifton et al (64) did not observe increases in the incidence of infection with 24 hour and 48 hour applications of hypothermia, respectively. However, longer applications of hypothermia may have considerable risk (18). In addition, application of mild hypothermia in settings not associated with ischemia but associated with considerable infection risk (such as elective abdominal surgery in patients with malignancies) increases infection rates (68).

Coagulopathy is suggested as another potential complication of hypothermia. However, in the studies of severely head injured patients by Marion et al (56), platelet counts and prothrombin times did not differ significantly between groups, and no difference in posttrauma intracranial hematomas or other hemorrhagic complications were noted despite the fact that some of the patients had multiple trauma. Cardiac arrhythmias were also not observed. The threshold for these complications appears to be
temperatures below 30°C (69,70). On the other hand, a recent report (71) suggested that morbid cardiac events after non-cardiac surgery were more common in mildly hypothermic patients compared to those who remained normothermic.

Finally, although the systemic complications appear relatively minimal for the transient (24 hour) application of mild or moderate hypothermia, one area of investigation that deserves further study is that of the effect of hypothermia on regenerative and endogenous defense mechanisms in brain. Goss et al (60) reported that 4 hours of moderate hypothermia resulted in a sustained inhibition of nerve growth factor production in brain after experimental contusion in rats. Nerve growth factor is an important homeostatic molecule in the central nervous system that upregulates antioxidant defenses and prevents apoptosis. The ramifications of this effect of hypothermia on brain parenchyma is currently under investigation.

Finally, another potential limitation of resuscitative hypothermia may be that it produces a temporary rather than sustained effect -- i.e., delays rather than ameliorates damage. This possibility was first suggested in classic studies of the effect of hypothermia on acute inflammation (67,72), and was re-introduced in work by Dietrich et al (73) in models of global cerebral ischemia, where brief episodes (1-3 hours) of hypothermia only delayed death of neurons in selectively vulnerable brain regions. Recent work by Colbourne et al (74), however, suggests that longer durations of hypothermia may produce permanent benefit.
Future directions

Some of the most intriguing recent work in the therapeutic application of hypothermia in laboratory studies involves the combination of hypothermia with other therapies. Dietrich et al (75) reported that combination of 3 hours of moderate hypothermia with sustained administration of the glutamate antagonist MK-801 produced a synergistic beneficial effect on neuronal survival in a model of global cerebral ischemia (Figure 3). Similar reports have been suggested for the combination of hypothermia and other therapies (76). Additional promising strategies that will require further study include the combination of hypothermia with either growth factors (60), anti-inflammatory agents or flow promoting treatments (60,77).

References


Table 1. Glasgow Outcome Scores at 3, 6, and 12 months after severe head injury in patients treated with 24 hours of hypothermia (32 °C) or normothermia, from the study by Marion et al (66). A beneficial effect of hypothermia versus normothermia on outcome was demonstrated in all patients at 6 months after injury. In patients with initial Glasgow Coma Scores of 5-7, a favorable effect of hypothermia vs normothermia was observed at 3, 6 and 12 months after injury. Reprinted from the *New England Journal of Medicine* with permission.
<table>
<thead>
<tr>
<th>GLASGOW OUTCOME SCORE</th>
<th>AT 3 MONTHS</th>
<th>AT 6 MONTHS</th>
<th>AT 12 MONTHS</th>
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<tr>
<td></td>
<td>HYPOThERMIA</td>
<td>NORMOTHERMIA</td>
<td>HYPOThERMIA</td>
</tr>
<tr>
<td>All patients</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1 (Death)</td>
<td>8 (20)</td>
<td>9 (21)</td>
<td>8 (20)</td>
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<tr>
<td>2 (Vegetative state)</td>
<td>6 (15)</td>
<td>11 (26)</td>
<td>3 (8)</td>
</tr>
<tr>
<td>3 (Severe disability)</td>
<td>11 (28)</td>
<td>15 (36)</td>
<td>7 (18)</td>
</tr>
<tr>
<td>4 (Moderate disability)</td>
<td>8 (20)</td>
<td>4 (10)</td>
<td>7 (18)</td>
</tr>
<tr>
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<td>40</td>
</tr>
<tr>
<td>P value‡</td>
<td>0.12</td>
<td>0.05</td>
<td></td>
</tr>
</tbody>
</table>

Patients with coma score of 5 to 7

|                       |             |             |             |             |             |             |
| 1 (Death)             | 2 (9)       | 5 (19)      | 2 (9)       | 6 (23)      | 2 (9)       | 6 (23)      |
| 2 (Vegetative state)  | 2 (9)       | 7 (27)      | 1 (5)       | 3 (12)      | 1 (5)       | 4 (15)      |
| 3 (Severe disability) | 6 (27)      | 9 (35)      | 3 (14)      | 8 (31)      | 3 (14)      | 6 (23)      |
| 4 (Moderate disability) | 6 (27) | 3 (12)      | 4 (18)      | 6 (23)      | 5 (23)      | 2 (8)       |
| 5 (Mild or no disability) | 6 (27) | 2 (8)       | 12 (55)     | 3 (12)      | 11 (50)     | 8 (31)      |
| Total                 | 22          | 26          | 22          | 26          | 22          | 26          |
| P value‡              | 0.01        |             | 0.01        |             | 0.04        |             |

*Percentages may not add to 100 because of rounding.

⁺One patient was lost to follow-up.

‡P values are for comparisons of all five outcomes in the hypothermia and normothermia groups.
Figure Legends

Figure 1. Survival after uncontrolled hemorrhagic shock (UHS) in rats from the study of Kim et al (78). The insult in all groups is comprised of a volume controlled initial hemorrhage followed by tail amputation. Treatments include normothermia (Nth, Group 1), hypothermia (Hth, Group 2, 30°C applied between 15 min and 120 min), normothermia plus lactated Ringers (LR) fluid resuscitation (Nth + LR, Group 3), or hypothermia plus fluid resuscitation (Hth + LR, Group 4). Survival to 72 hours was maximal in rats treated with hypothermia plus LR. Reprinted from the Journal of Trauma with permission.

Figure 2. Desitometric analysis of RNA gel blot hybridizations for IL-1β message before, and at serial times after experimental cerebral contusion in rats, from the work of Goss et al (60). Filled circles represent data from rats maintained at brain temperature 37 °C, while solid squares represent data from rats maintained at a brain temperature of 32 °C for 4 hours after injury. A marked increased in IL-1β message was observed at 5.5 hours after injury which was partially attenuated by hypothermia. Reprinted form the Journal of Neurotrauma with permission.

Figure 3. Bar graph from the work of Dietrich et al (75) showing the number of normal appearing neurons in striatum at 2 months after sham operation or cerebral ischemia in rats treated with normothermia (37 °C), the glutamate receptor antagonist MK-801,
hypothermia (30 °C), or the combination of hypothermia plus MK-801. Neuronal survival was maximal after treatment with the combination of moderate hypothermia and MK-801. Reprinted from the *Journal of Cerebral Blood Flow and Metabolism* with permission.

**CME Questions**

1. Randomized, controlled clinical trials of moderate hypothermia in cerebral resuscitation have progressed most rapidly for which of the following insults?
   a. Stroke
   b. Cardiopulmonary arrest
   c. Traumatic brain injury
   d. Near drowning
   e. Hemorrhagic shock

2. This factor re-vitalized interest in the potential application of deliberate hypothermia to cerebral resuscitation.
   a. Remarkably mild hypothermia (a temperature reduction of between 3° and 5°C) was effective in reducing secondary brain damage in laboratory models.
   b. Brain temperature, not body temperature, was a critical therapeutic target.
   c. The duration of hypothermia necessary for a beneficial effect in some
models was transient --as short as 1 hour,

d. b and c

e. All of the above

3. Controlled clinical trials of moderate hypothermia, applied for 24 hours, suggest that the incidence of side effects such as coaguloapthy, arrhythmias, or infectious complications are not significantly different from that seen during normothermia.
   a. True
   b. False

4. Based on the results of clinical trials, beneficial effects of mild or moderate hypothermia in injured brain may result from a reduction of which of the following?
   a. Excitotoxicity
   b. Regeneration
   c. Inflammation
   d. a and c
   e. All of the above

5. Recent studies in laboratory models of hemorrhagic shock have demonstrated that application of mild or moderate hypothermia during the “golden hour” reduces mortality rate.
   a. True
   b. False
Answers: 1 (c), 2 (e), 3 (a), 4 (d), 5 (a)
Figure 3

![Graph showing number of normal appearing neurons under different conditions](image-url)
No long-term benefit from hypothermia after severe traumatic brain injury with secondary insult in rats

Courtney L. Robertson, MD; Robert S. B. Clark, MD; C. Edward Dixon, PhD; Henry L. Alexander, BS; Steven H. Graham, MD, PhD; Stephen R. Wisniewski, PhD; Donald W. Marion, MD; Peter J. Safar, MD; Patrick M. Kochanek, MD

Objectives: To evaluate the effect of application of transient, moderate hypothermia on outcome after experimental traumatic brain injury (TBI) with a secondary hypoxemic insult.

Design: Prospective, randomized study.

Setting: University-based animal research facility.

Subjects: Male Sprague-Dawley rats.

Interventions: All rats were subjected to severe TBI followed by 30 mins of moderate hypoxemia, associated with mild hypotension. Rats were randomized to three groups: a) normothermia (37°C ± 0.5°C); b) immediate hypothermia (32°C ± 0.5°C initiated after trauma, before hypoxemia); and c) delayed hypothermia (32°C ± 0.5°C after hypoxemia). The brain temperature was controlled for 4 hrs after TBI and hypoxemia.

Measurements and Main Results: Animals were evaluated after TBI for motor and cognitive performance using beam balance (days 1–5 after TBI), beam walking (days 1–5 after TBI), and Morris Water Maze (days 14–18 after TBI) assessments. On day 21 after TBI, rats were perfused with paraformaldehyde and brains were histologically evaluated for lesion volume and hippocampal neuron counts. All three groups showed marked deficits in beam balance, beam walk, g, and Morris Water Maze performance. However, these deficits did not differ between groups. There was no difference in lesion volume between groups. All animals had significant hippocampal neuronal loss on the side ipsilateral to injury, but this loss was similar between groups.

Conclusions: In this rat model of severe TBI with secondary insult, moderate hypothermia for 4 hrs posttrauma failed to improve motor function, cognitive function, lesion volume or hippocampal neuronal survival. Combination therapies may be necessary in this difficult setting. (Crit Care Med 2000; 28:000–000)

Keywords: traumatic brain injury; hypothermia; hypoxemia

Secondary insults after experimental traumatic brain injury (TBI) have been shown to exacerbate disturbances in key physiologic variables, including hypoperfusion, energy failure, cerebral edema, and electroencephalographic suppression (1–3). In addition, animals subjected to a secondary hypoxemic insult after TBI have worse motor and histologic outcomes than those subjected to TBI alone (1, 3, 4). After severe TBI, patients often experience a variety of secondary systemic insults related to extracerebral traumatic injury. As many as 20% to 50% of patients presenting with severe TBI have experienced a period of hypoxemia (5–7). Postmortem findings of head-injured patients (8) demonstrate evidence of ischemic neuronal death throughout the brain. Similarly, clinical studies have demonstrated higher morbidity and mortality among head-injured patients who had experienced a secondary insult, specifically hypoxemia or hypotension (7). Often, the most severely devastated patients are those who experience the combination of TBI with hypoxemia and hypotension.

Hypothermia has been used as a successful treatment modality after brain injury in many experimental models and clinical settings. Neuroprotective effects of hypothermia in animal models include attenuation of release of excitatory amino acids (9–11), reduction in hydroxyl radicals (9) and inflammatory mediators (12, 13), and reduction in disruption of the blood-brain barrier (14). In the setting of experimental TBI, hypothermia improves outcome (15, 16). In models of fluid percussion injury (15) and controlled cortical impact (CCI) (16), reductions in both functional and motor deficits are observed in animals treated with moderate hypothermia after TBI when compared with normothermic animals. Transient, moderate hypothermia applied after global or focal ischemic insult in animal models has improved histologic outcomes (17–19). Clinical studies have similarly demonstrated improvements in functional outcomes (20) and intracranial pressure (21, 22) in patients treated with moderate hypothermia.

Despite the importance of secondary insults to clinical outcome after TBI, the variety of experimental models of TBI and secondary insult that have been developed, and the success of hypothermia in both clinical and experimental TBI, the effect of the application of hypothermia in the setting of TBI with secondary insult has not been studied. We hypothesized that moderate hypothermia would improve outcome after CCI with secondary insult in rats.
MATERIALS AND METHODS

This study was approved by the University of Pittsburgh Animal Care and Use Committee. The care and handling of animals were in accord with National Institute of Health guidelines.

Experimental Protocol. Virus-free male Sprague-Dawley rats (329–460 g) were studied. The animals were allowed free access to food and water before and after surgery. All surgical procedures were performed using aseptic technique.

Anesthesia was induced in a plastic jar with 4% isoflurane (Anaquest, Memphis, TN) in oxygen. The trachea was intubated with a 14-gauge angiocatheter and the lungs were mechanically ventilated with 2% isoflurane/66% N₂O/oxygen. A femoral arterial catheter (PE-50) was inserted for continuous monitoring of blood pressure and arterial blood sampling. Pancuronium bromide (0.1 mg/kg/hr) (Elkins-Sinn, Cherry Hill, NJ) was given for immobilization. A rectal probe was inserted to monitor core temperature.

Traumatic Brain Injury Model. The head was fixed in a stereotactic device (David Kopf, Tujunga, CA) and a midline scalp incision was made to expose the parietal bone. A craniotomy was made over the left parietal cortex with a dental drill, using the coronal and interparietal sutures as margins. The intact dura and bone flap were left in place until immediately before trauma. A temperature probe (outside diameter, 0.009 inch) (Physiostemp, Clifton, NJ) was inserted through a burr hole into the left parietal cortex 5 mm anterior to the bregma and 2 mm lateral to the sagittal suture. Rats were equilibrated under anesthesia (1.1% isoflurane/66% N₂O/oxygen) at a brain temperature of 37°C ± 0.5°C for 30 min before TBI. An arterial blood sample (0.5 mL) was obtained 15 min before trauma to verify normal arterial blood gas tensions, serum glucose, and hematocrit.

TBI was performed using the CCI device (23, 24), with minor modifications to the procedure previously described (25). Briefly, after removal of the bone flap, injury was produced using a device with a 6-mm metal impactor tip that is pneumatically driven in the vertical plane at a predetermined depth, velocity, and duration of brain deformation. For all studies, a depth of penetration of 2.5 mm, a velocity of 4.0 ± 0.2 m/sec, and a duration of deformation of 50 msec was used. After trauma, the bone flap was replaced and sealed with dental cement (Koldmount, Vernon Bantshoff, Albany, NY) and the scalp incision was closed.

Secondary Insult. Beginning 1 min after CCI, all rats underwent a 30-min period of hypoxemia as previously described (4). Air and oxygen were blended to achieve an FIO₂ of 11% (1.1% isoflurane/74% N₂O/19% air/6% oxygen). This produced a PaO₂ range of 46–51 and MAP 50–54. Arterial blood gas samples were obtained in all rats at 10 min and 25 min during the hypoxic period.

Hypothermia. Rats were randomized into three groups: normothermia, immediate hypothermia, and delayed hypothermia. Hypothermia (32°C) was achieved by the external application of ice packs to the head to lower brain temperature over a 15-min period. This temperature was maintained for 4 hrs and the brain was rewarmed over 1 hr. Arterial blood gas samples were obtained every hour during the hypothermia period. Physiologic variables (MAP, brain temperature, rectal temperature) were recorded every 30 min during hypothermia and during rewarming.

The immediate hypothermia group (n = 10) had brain cooling initiated immediately after trauma, coincident with the onset of hypoxemia. The delayed hypothermia group (n = 10) had cooling initiated on the completion of hypoxemia, 30 mins after TBI. The normothermia group (n = 19) had brain temperature maintained at 37°C ± 0.5°C throughout the experimental period. At the end of the experiment, after completion of rewarming, anesthesia was discontinued. Rats were extubated, placed in 100% oxygen for an additional 30 mins, then returned to their cages, where they were allowed free access to food and water.

Motor Function Assessments. Gross vestibulomotor function was assessed using a beam-balance task (24, 26). The rats were trained by three trials before TBI to obtain a baseline measurement. Beam-balance latency (up to 60 sec) was measured on days 1–5 after TBI.

Fine vestibulomotor function and coordination were assessed using a beam-walking task (27). Performance was assessed by measuring the rat's latency to traverse the beam and enter a goal box. Beam-walking latency was measured on days 1–5 after TBI.

Cognitive Assessment (Morris Water Maze). Water-maze testing started on day 14 postinjury. The hidden platform task assesses the rat's ability to learn spatial relations between distal cues and the escape platform. Performance is impaired by cortical and hippocampal lesions. We used a variant of the Morris water maze (28). Rats were given 120 secs to find the hidden platform. If the rat failed to find the platform within 120 secs, it was placed on the platform by the experimenter. Rats were given four swimming trials per day for 5 consecutive days. Water maze tests were given on days 14–18 after TBI. The last 2 days of testing consisted of a visible platform task in which the platform was raised 2 cm above the water surface. This visible task controls for potential nonspecific visual or motor deficits.

Lesion Volume Analysis. At 21 days after TBI, rats were anesthetized and perfused with 500 mL of 4% buffered formaldehyde. Brains were removed and postfixed for a minimum of 1 wk at 4°C and cryoprotected in sucrose. Coronal sections (10-μm) were prepared through the entire brain at 1-mm intervals from the occiput. Sections were stained with cresyl violet. In the serial sections taken at 1-mm intervals, the margins of both the contusion and the total left hemisphere were outlined by a blinded observer using image analysis (Image Research). Contusion and hemispheric areas were measured. Contusion volume was calculated and expressed as mm³.

Hippocampal Cell Counts. Neuronal loss in hippocampal CA1 and CA3 pyramidal layers was quantified using a method previously described by Clark et al. (4). A coronal section through the dorsal hippocampus underlying the area of contusion was used for analysis. This location was −2.6 mm posterior to bregma. The regions were visualized at magnification ×400 by a blinded observer. CA1 and CA3 hippocampal neurons were counted in six separate fields for each region in both injured and uninjured hemispheres. Only complete cells with a defined cell body and intact nucleus were counted. Hippocampal neuron survival was reported as the average number of surviving neurons per high power field ipsilateral to injury.

Statistical Analysis. All data are presented as mean ± SEM. Because of the large number of physiologic variables recorded, comparisons of physiologic data were made using a multiple regression analysis, evaluating the effect of treatment and time for each variable. Beam balance, beam walking, and Morris water maze data were analyzed using repeated-measures analysis of variance using GB-STAT statistical software. Lesion volumes and hippocampal neuron counts were compared using the Kruskal-Wallis test and Dunn's test. A significance level of p < .05 was used for all tests.

RESULTS

Physiologic Variables. Physiologic data (including brain and rectal temperature, MAP, arterial pH and blood gases, blood glucose, and hematocrit) from all animals that survived the experimental protocol are presented in Table 1. There were no differences between groups in pH, PaCO₂, and hematocrit. As expected, the groups differed in brain and rectal temperatures (p < .05), and this difference was seen between groups when controlling for time. Brain temperature remained at 37°C ± 0.2°C in the normothermia group. Brain temperature decreased from 37.2°C ± 0.1°C before trauma to 32.0°C ± 0.1°C by 25 mins of cooling in the immediate hypothermia group. Brain temperature decreased from 37.1°C ± 0.1°C before trauma to 32.1°C ± 0.1°C by 25 mins of cooling in the delayed hypothermia group.

Groups differed in MAP when controlling for time (p < .05). Initial MAP in the normothermia group was 91–92 mm Hg and decreased to 62–63 mm Hg during...
Table 1. Physiologic data

<table>
<thead>
<tr>
<th>Group</th>
<th>Brain Temperature (°C)</th>
<th>Rectal Temperature (°C)</th>
<th>MAP (mm Hg)</th>
<th>PH</th>
<th>PaCO₂ (mm Hg)</th>
<th>PaO₂ (mm Hg)</th>
<th>Glucose (mg/dL)</th>
<th>HCT (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Baseline</td>
<td>37.1 ± 0.1</td>
<td>37.3 ± 0.1</td>
<td>91 ± 3</td>
<td>7.44 ± 0.01</td>
<td>39 ± 1</td>
<td>169 ± 4</td>
<td>182 ± 10</td>
<td>40 ± 0</td>
</tr>
<tr>
<td>Insult</td>
<td>37.0 ± 0.1</td>
<td>37.3 ± 0.0</td>
<td>92 ± 3</td>
<td>7.43 ± 0.01</td>
<td>40 ± 1</td>
<td>46 ± 1</td>
<td>173 ± 13</td>
<td>39 ± 0</td>
</tr>
<tr>
<td>10 mins</td>
<td>37.1 ± 0.1</td>
<td>37.1 ± 0.1</td>
<td>63 ± 4</td>
<td>7.42 ± 0.01</td>
<td>40 ± 1</td>
<td>47 ± 1</td>
<td>154 ± 5</td>
<td>39 ± 0</td>
</tr>
<tr>
<td>25 mins</td>
<td>37.0 ± 0.0</td>
<td>37.1 ± 0.1</td>
<td>62 ± 3</td>
<td>7.43 ± 0.01</td>
<td>39 ± 1</td>
<td>160 ± 3</td>
<td>145 ± 4</td>
<td>39 ± 0</td>
</tr>
<tr>
<td>1 hr</td>
<td>37.1 ± 0.0</td>
<td>37.2 ± 0.0</td>
<td>84 ± 2</td>
<td>7.41 ± 0.01</td>
<td>37 ± 1</td>
<td>172 ± 3</td>
<td>148 ± 4</td>
<td>38 ± 0</td>
</tr>
<tr>
<td>3 hrs</td>
<td>37.1 ± 0.1</td>
<td>37.2 ± 0.0</td>
<td>85 ± 3</td>
<td>7.42 ± 0.01</td>
<td>37 ± 1</td>
<td>172 ± 3</td>
<td>148 ± 4</td>
<td>38 ± 0</td>
</tr>
<tr>
<td>32°C—Immediate (n = 10) Baseline</td>
<td>37.2 ± 0.1</td>
<td>37.3 ± 0.1</td>
<td>103 ± 2</td>
<td>7.45 ± 0.01</td>
<td>40 ± 1</td>
<td>157 ± 3</td>
<td>171 ± 8</td>
<td>41 ± 1</td>
</tr>
<tr>
<td>Insult</td>
<td>37.2 ± 0.0</td>
<td>37.3 ± 0.1</td>
<td>104 ± 3</td>
<td>7.44 ± 0.01</td>
<td>37 ± 1</td>
<td>51 ± 1</td>
<td>166 ± 11</td>
<td>39 ± 0</td>
</tr>
<tr>
<td>10 mins</td>
<td>32.5 ± 0.2</td>
<td>32.5 ± 0.2</td>
<td>74 ± 5</td>
<td>7.44 ± 0.01</td>
<td>37 ± 1</td>
<td>50 ± 1</td>
<td>208 ± 22</td>
<td>39 ± 0</td>
</tr>
<tr>
<td>25 mins</td>
<td>32.0 ± 0.1</td>
<td>32.1 ± 0.1</td>
<td>68 ± 3</td>
<td>7.45 ± 0.01</td>
<td>37 ± 1</td>
<td>201 ± 5</td>
<td>166 ± 11</td>
<td>39 ± 0</td>
</tr>
<tr>
<td>1 hr</td>
<td>31.9 ± 0.1</td>
<td>32.0 ± 0.1</td>
<td>91 ± 4</td>
<td>7.45 ± 0.01</td>
<td>37 ± 1</td>
<td>193 ± 6</td>
<td>182 ± 15</td>
<td>39 ± 0</td>
</tr>
<tr>
<td>3 hrs</td>
<td>31.2 ± 0.1</td>
<td>32.0 ± 0.1</td>
<td>94 ± 3</td>
<td>7.42 ± 0.01</td>
<td>38 ± 0</td>
<td>193 ± 6</td>
<td>182 ± 15</td>
<td>39 ± 0</td>
</tr>
<tr>
<td>32°C—Delayed (n = 10) Baseline</td>
<td>37.1 ± 0.1</td>
<td>37.3 ± 0.1</td>
<td>85 ± 2</td>
<td>7.43 ± 0.01</td>
<td>39 ± 1</td>
<td>176 ± 7</td>
<td>196 ± 14</td>
<td>41 ± 1</td>
</tr>
<tr>
<td>Insult</td>
<td>37.1 ± 0.1</td>
<td>37.3 ± 0.1</td>
<td>84 ± 2</td>
<td>7.42 ± 0.01</td>
<td>39 ± 1</td>
<td>176 ± 7</td>
<td>196 ± 14</td>
<td>41 ± 1</td>
</tr>
<tr>
<td>10 mins</td>
<td>36.9 ± 0.1</td>
<td>37.1 ± 0.1</td>
<td>54 ± 3</td>
<td>7.43 ± 0.01</td>
<td>39 ± 1</td>
<td>173 ± 10</td>
<td>39 ± 0</td>
<td></td>
</tr>
<tr>
<td>25 mins</td>
<td>36.9 ± 0.1</td>
<td>37.1 ± 0.1</td>
<td>50 ± 2</td>
<td>7.41 ± 0.01</td>
<td>39 ± 1</td>
<td>173 ± 10</td>
<td>39 ± 0</td>
<td></td>
</tr>
<tr>
<td>1 hr</td>
<td>31.9 ± 0.1</td>
<td>31.9 ± 0.1</td>
<td>79 ± 2</td>
<td>7.43 ± 0.01</td>
<td>38 ± 1</td>
<td>205 ± 3</td>
<td>188 ± 11</td>
<td>39 ± 0</td>
</tr>
<tr>
<td>3 hrs</td>
<td>31.9 ± 0.1</td>
<td>32.0 ± 0.1</td>
<td>71 ± 5</td>
<td>7.41 ± 0.01</td>
<td>36 ± 0</td>
<td>215 ± 3</td>
<td>182 ± 9</td>
<td>37 ± 1</td>
</tr>
</tbody>
</table>

MAP, mean arterial pressure; HCT, hematocrit.

Values are mean ± SEM.

hypoxemia, returning to a postinsult level of 84–85 mm Hg. The immediate hypothermia group started with a baseline MAP of 103–104 mm Hg, decreased to 68–74 mm Hg during hypoxemia, and returned to 91–94 mm Hg postinsult. The delayed hypothermia group had the lowest overall MAP, starting with a baseline of 84–85 mm Hg, decreasing to 50–54 mm Hg during hypoxemia and returning to 71–76 mm Hg postinsult.

Groups also differed in PaO₂ and glucose when controlling for time (both p < .05). The differences between groups appeared modest and unlikely to be clinically significant. Glucose levels were also different between groups over time. The normothermia group had initial glucose levels of 182 mg/dL and decreased to levels of 148 mg/dL at 3 hrs postinsult. The immediate and delayed hypothermia groups had more stable glucose levels.

Survival. Survival rate to 21 days for completion of motor and cognitive testing was 79% for the normothermia group, 80% for the immediate hypothermia group, and 62% for the delayed hypothermia group. Motor performance, cognitive performance, lesion volume analysis, and hippocampal neuron counts are reported on all animals that survived to cognitive testing at 20 days and were able to swim for water maze testing.

Motor Performance. All three groups showed a decrease in beam balance performance and an increase in beam walking latency after trauma. However, there was no difference in beam balance duration (Fig. 1) or beam walking latency (Fig. 2) between normothermic and hypothermic groups.

Cognitive Performance (Morris Water Maze). All three groups showed a marked latency in finding the submerged platform on days 14–18 after trauma. However, there was no difference between groups in performance in the water maze (Fig. 3). The groups were similar in discovery of the visible platform on days 19–20 after trauma (Fig. 3).

Lesion Volume Analysis. Lesion volumes (mm³) measured at 21 days after TBI are shown in Table 2. There appeared to be a reduction in lesion volume in the hypothermia vs. normothermic groups. However, this reduction in lesion volume was not statistically different from normothermia. Lesion area at various distances from the occiput is shown in Figure 4.

Hippocampal Neuron Counts. Surviving hippocampal neuron counts are shown in Table 3. Both normothermic and hypothermic animals had significant hippocampal neuronal loss on the side ipsilateral to injury. For comparison, average neuron counts in CA1 and CA3 hippocampus on the side contralateral to injury were 48–56 cells/high-power field. There were no significant differences in the number of surviving CA1 or CA3 hippocampal neurons in the hypothermic vs. normothermic groups.

DISCUSSION

To our knowledge, this is the first study to evaluate the effect of hypothermia on severe experimental TBI with secondary insult. Surprisingly, no difference in motor performance, cognitive performance, lesion volume, or hippocampal neuronal survival was observed with the application of moderate hypothermia after severe TBI with secondary insult in a rat model. Also unexpectedly, both the
Figure 2. Beam walking latency in normothermia (solid circles), immediate hypothermia (open circles), and delayed hypothermia (triangles) on days 1–5 after controlled cortical impact (CCI). Values are mean ± SEM.  

Figure 3. Morris Water Maze performance in normothermia (solid circles), immediate hypothermia (open circles), and delayed hypothermia (triangles) on days 14–18 after controlled cortical impact (CCI); and visible platform latency in all groups on days 19–20 after CCI. Values are mean ± SEM.  

Table 2. Lesion Volume

<table>
<thead>
<tr>
<th>Group</th>
<th>Lesion Volume (mm³)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normothermia</td>
<td>65.3 ± 6.9</td>
</tr>
<tr>
<td>Immediate HT</td>
<td>50.2 ± 8.2</td>
</tr>
<tr>
<td>Delayed HT</td>
<td>53.7 ± 7.9</td>
</tr>
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</table>

HT, hypothermia.  
*p = .32 by analysis of variance. Values are expressed as mean ± SEM.  

immediate hypothermia group, with hypothermia initiated after trauma and before secondary hypoxemia, and the delayed hypothermia group, with hypothermia applied after both brain trauma and hypoxemia, demonstrated similar functional and histologic deficits when compared with each other and to the normothermia group.  

Beneficial effects of hypothermia on histopathologic outcome after TBI have been demonstrated by several investigators. The timing of this histologic evaluation appears to be important. Dietrich et al. (29) showed a reduction in cortical contusion volume and frequency of necrotic cortical neurons in rats that received 3 hrs of immediate posttrauma hypothermia (30°C) after parasagittal fluid percussion injury. These animals were evaluated at 3 days posttrauma. However, in the same model, investigators found no difference in hippocampal CA1, CA3, CA4, or dentate neuronal survival in rats receiving posttrauma hypothermia compared with normothermic animals when brains were analyzed at 8 wks after TBI (30). In models of ischemic brain injury, transient application of therapeutic hypothermia has also shown a temporary beneficial effect on hippocampal neuronal survival. Early evaluation revealed decreased hippocampal CA1 cell loss, but this protection by posttrauma hypothermia (30°C) was not seen in the animals evaluated at 2 months after ischemia insult (31). In our model, severe TBI was followed by 30 mins of hypoxemia. All animals showed a reduction in systemic blood pressure during the hypoxicemic period, likely highlighting an ischemic component to the secondary insult.  

Table 3. Hippocampal neuronal survival

<table>
<thead>
<tr>
<th>Group</th>
<th>CA1 Neurons (cells/hpf)*</th>
<th>CA3 Neurons (cells/hpf)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normothermia</td>
<td>19.4 ± 4.2</td>
<td>19.8 ± 4.6</td>
</tr>
<tr>
<td>Immediate HT</td>
<td>13.2 ± 8.7</td>
<td>15.6 ± 7.3</td>
</tr>
<tr>
<td>Delayed HT</td>
<td>13.7 ± 5.8</td>
<td>18.5 ± 7.3</td>
</tr>
</tbody>
</table>

hpf, high powered field; HT, hypothermia.  
*Intraplantar to injury. Values are expressed as mean ± SEM.  

The amount of tissue loss after experimental TBI varies greatly dependent on the model. This model of severe CCI followed by 30 mins of hypoxemia produced lesion volumes of 50–65 mm³. This is much larger than contusion volumes seen in other traumatic injury models, such as lateral fluid percussion (2.14 mm³) (27), or in similar CCI models without hypoxemia (~30 mm³) (5). The severe insult produced in this model might explain the failure of posttrauma hypothermia to show a significant reduction in lesion volume. However, other experimental TBI models applying hypothermia after injury have also failed to reduce necrotic volumes (30, 32). Cherian et al. (33) showed increasing sizes of contusion volume as the degree of secondary insult (bilateral carotid occlusion) increased after CCI in rats. In addition, it is likely that the lesion volume observed at 21 days postsinjury is the result of damage by many different mechanisms operating in the early and late posttrauma phases. Clark et al. (4) has demonstrated cells with either necrotic or apoptotic phenotypes in various brain regions after TBI in a similar model with hypoxemia. It is unclear if earlier assessment would have revealed more hippocampal protection with posttraumatic hypothermia in this model. However, our goal is to favorably influence long-term outcome. Hypothermia as a single treatment modality, and applied for only 4 hrs after TBI, might be unable to reduce overall lesion volumes in such a model.  

Previous experimental studies applying moderate hypothermia after TBI have demonstrated protection against motor
and spatial memory deficits after both CCI (16) and fluid percussion injury (30, 34). However these models did not include a period of hypoxemia or any other secondary insults after trauma. This secondary hypoxic insult worsens histologic outcome and could therefore worsen behavioral outcome after TBI. In a similar model of CCI with secondary hypoxemia, rats demonstrated progressively worse motor function (beam-balance latency) with increasing amounts of postrumta hypoxemia (4). This trend was seen even in rats who received mild hypoxemia (PaO2, 55–63 mm Hg) after CCI. In a fluid percussion injury model, Ishige et al. (1) showed significantly worse neurologic status scores in rats that underwent impact injury followed by a 30-min period of hypoxemia (PaO2, 35–40 mm Hg) vs. those injured without secondary hypoxemia. These neurologic deficits were also not observed in rats that received hypoxia alone.

Clinical studies of patients with head injury have also reported marked worsening of outcome variables in the setting of TBI with secondary insult such as hypoxemia or hypotension. In an analysis of 717 patients from the Traumatic Coma Data Bank, Chesnut et al. (7) found that hypoxia and hypotension were independently associated with increases in morbidity and mortality from severe head injury. This study showed a marked shift toward vegetative/dead outcomes in patients who endured hypoxia (PaO2 ≤60 mm Hg) or hypotension (systolic blood pressure ≤90) during the prehospital or resuscitation period. This is especially relevant to our model of TBI because rats underwent a planned 30-min period of hypoxemia, which was also associated with a decrease in systemic blood pressure. The difference in MAP between groups may have caused additional experimental differences. The delayed hypothermia group had an overall lower MAP trend and may have experienced more significant secondary ischemic flows.

Clinical studies applying hypothermia after TBI have yielded a variety of positive results. In a phase II study of moderate hypothermia, Clifton et al. (35) found a reduction in incidence of posttraumatic seizures. Shiozaki et al. (22) documented improved control of intracranial pressure with the application of mild (34°C) hypothermia after conventional therapies had failed to control intracranial pressure (22). Most recently, Marion et al. (20) demonstrated faster recovery of functional outcome with the application of moderate hypothermia (32°C) after severe TBI in 82 patients randomized to either normothermia or hypothermia for 24 hrs after injury. Relevant to our findings, these clinical trials showed important distinctions in the subset of very severely injured patients. Marion’s study included all patients with initial Glasgow Coma Scale (GCS) scores of ≤8, but the beneficial effect of hypothermia only extended to 12 months in the subset of patients with an initial GCS score of 5–7 (20). In the study by Shiozaki et al. (22), the subset of patients admitted with GCS scores of 3–4 had a much lower incidence of favorable outcome at 6 months after injury (only 1 of 22 patients, 4.5%) vs. the group with GCS scores of 5–7 (11 of 40 patients, 27.5%), despite the application of mild hypothermia. Importantly, Marion et al. (20) excluded patients with hypoxia or hypotension.

Our model of CCI with hypoxemia results in a very severe injury relative to other models. In addition, during the 30-min period of hypoxemia, the rats experienced a significant drop in their mean arterial pressure. In a recent experimental TBI model (36), posttraumatic application of moderate hypothermia (30°C for 3hrs) resulted, after rewarming, in a lower cerebral perfusion without a corresponding decrease in cerebral glucose utilization, creating a state of metabolism to blood flow mismatch. This may be especially important in the clinical setting in which severely head-injured patients undergo a secondary insult of hypoxia and/or hypotension before initiation of treatment for their head injury. Analysis of trauma patients has consistently revealed worse outcomes in patients who experienced sustained hypothermia in the prehospital setting vs. those who remained normotensive. Specifically, Chesnut et al. (7) showed hypothermia was associated with a 150% increase in mortality rate. Wald et al. (37) also found that prehospital hypothermia doubled the incidence of adverse outcome (37). In a review of pediatric trauma patients, Pigula et al. (38) found that hypothermia significantly increased the mortality rate. As a result, patients with significant secondary insult have been excluded from evaluation in clinical trials (20). This subset clearly represents a population in which currently available interventions, even those proven to be effective in the sets of TBI alone, may have limited efficacy.

There are many mechanisms of cell injury and death after TBI. Investigators have shown additional pathophysiologic mechanisms operating when secondary insults were added to the already vulnerable traumatically injured brain (1, 3, 33, 39). Recent experimental investigations have focused on the period after trauma, during which secondary insults may potentiate neuronal damage, utilizing a wide variety of therapies. However, many of these therapies have been tested in models of TBI where oxygenation and ventilation of animals after TBI are controlled. Given that many head-injured patients present with preceding hypoxemia, it may be important to reassess these therapies in models simulating this clinical setting. It is possible that proven treatments may be less effective in a model with applied hypoxemia and accompanying hypotension. Novel therapies targeting this complex clinical scenario have yet to be developed.

In conclusion, in this rat model of severe CCI with hypoxemia, moderate hypothermia for 4 hrs posttrauma failed to improve hippocampal neuron survival, lesion volume, motor function, or cognitive function. Combination therapies or development of novel therapies may be necessary to see significant improvement in outcome in this difficult setting.

ACKNOWLEDGMENT

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amine, haloperidol and experience interact to affect rate of recovery after motor cortex injury. Science 1982; 217:855–857


**AWARDS**


(2000) Educational Scholarship to Dr. Kimberly Statler from the Society of Critical Care Medicine for her abstract entitled, “Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats.”

**9. CONCLUSIONS**

1. Based on the important finding in this study where the use of fentanyl in our CCI model produced deleterious effects on outcome after TBI, animal models should utilize clinically relevant sedative/analgesic treatments. The beneficial mechanisms of isoflurane (possibly promotion of cerebral blood flow or reduction of excitotoxicity) should be investigated for the development of novel treatments. Isoflurane anesthesia likely provides powerful beneficial effects at or near the time of injury. Sedative agents with anti-excitotoxic and/or cerebral blood flow promoting actions similar to isoflurane may thus represent better alternatives early after TBI. **Narcotics may not be the optimal sedative/analgesic early after TBI. Better field therapy than narcotics must be developed.**

2. Based on studies in rats using the CCI model, we have demonstrated tangible risk to aggressive, indiscriminate hyperventilation early after injury—specifically—augmentation of neuronal death in selectively vulnerable brain regions. This suggests that aggressive hyperventilation should not be indiscriminately used in the field
treatment of TBI, rather it should be applied if there are signs and/or symptoms of herniation. Mild hyperventilation (used in our control group) or normocapnia may be preferable.

3. Based on our studies in rats, hypothermia, although showing some beneficial effects, particularly early after TBI (such as a reduction in DNA damage, etc), may have some deleterious effects which result in only modest overall beneficial effects on long-term outcome. This is particularly true in the setting of severe injury (such as TBI plus secondary hypoxemic insults) where it is possible that there is little to gain except side effects. Also based on our narcotic (fentanyl) vs isoflurane study and our multi-sedation study, hypothermia should be re-examined in future studies with narcotic anesthesia, since beneficial effects of isoflurane may be masking any benefit from hypothermia.

(10) REFERENCES


(11) APPENDIX

1. 1997 Report

2. 1998 Report

3. Curriculum Vitae

4. Manuscripts:


5. Abstracts:


(12) BINDING (N/A)

(13) FINAL REPORTS
   a) Bibliography of all publications and abstracts (see appendix)
   b) List of personnel receiving pay from the research effort
      Patrick M. Kochanek, M.D.
      Peter Safar, M.D.
      Henry Alexander
      Scott Heineman
      Marci Provins
      Linda Amick
ISOFLURANE IMPROVES LONG-TERM NEUROLOGIC OUTCOME VS
FENTANYL AFTER TRAUMATIC BRAIN INJURY IN RATS

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M.D.,1,2,6 Stephen R. Wisniewski, Ph.D.,4 Steven H. Graham, M.D., Ph.D.,7
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Running Title: Isoflurane vs fentanyl after TBI in rats

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Abstract

Despite routine use of fentanyl in patients after traumatic brain injury (TBI), it is unclear if it is the optimal sedative/analgesic agent. Isoflurane is commonly used in experimental TBI. We hypothesized that isoflurane would be neuroprotective vs fentanyl after TBI. Rats underwent controlled cortical impact (CCI) and received 4 h of N₂O:O₂ (2:1) and either fentanyl (10 µg/kg iv bolus, 50 µg/kg/h infusion) or isoflurane (1% by inhalation) with controlled ventilation. Shams underwent identical preparation, without CCI. Functional outcome (beam balance, beam walking, Morris water maze [MWM] tasks) was assessed over 20 days. Lesion volume and hippocampal neuron survival were quantified on d 21. Additional rats underwent identical CCI and anesthesia with intracranial pressure (ICP) monitoring, and brain water content was assessed.

Motor and MWM performances were better in injured rats treated with isoflurane vs fentanyl ($p < 0.05$). CA1 hippocampal damage was attenuated in isoflurane-treated rats ($p < 0.05$). Fentanyl-treated rats had higher mean arterial blood pressure after injury ($p < 0.05$); however, ICP and brain water were similar between groups.

Isoflurane improved functional outcome and attenuated damage to CA1 vs fentanyl in rats subjected to CCI. Isoflurane may be neuroprotective by augmenting cerebral blood flow and/or reducing excitotoxicity, not by reducing
ICP or brain water content. Alternatively, fentanyl may be detrimental. Isoflurane may mask beneficial effects of novel agents tested in TBI models. Additionally, fentanyl may not be optimal early after TBI in humans.

**Key words:** sedation, analgesia, anesthesia, head injury, narcotics, opioids
Introduction

In current clinical practice, opioids are routinely administered after traumatic brain injury (TBI). Fentanyl is one of the first-line agents because of its short half-life and low incidence of hypotension. Despite standard clinical use, it remains unclear if fentanyl represents the optimal sedative/analgesic agent in the acute period following TBI. Unlike the clinical arena, opioids are rarely used in experimental TBI. In fact, most models of TBI use isoflurane or pentobarbital anesthesia.

Much of the study of opioids in TBI has focused on the actions of endogenous opiates, such as dynorphin, and specific opiate receptor effects (Hayes et al., 1990; Lyeth et al., 1993, 1995; McIntosh et al., 1987; Vink et al., 1990). Although mu receptor agonists, such as morphine and fentanyl, have been shown to have some beneficial effects after central nervous system injury (Lyeth et al., 1993, 1995), recent studies in cerebral ischemia and focal cryogenic lesion suggest that isoflurane may be neuroprotective compared to fentanyl (Miura et al., 1998; Murr et al., 1993, 1995; Soonthorn-Brant et al., 1999). In rats subjected to global cerebral ischemia, isoflurane reduced neuronal damage and improved motor function compared to fentanyl (Miura et al., 1998). Similarly, after focal ischemia, rats anesthetized with isoflurane had smaller infarct volumes than those receiving fentanyl. Lesion volumes in rats treated with fentanyl were similar to
those in unanesthetized control rats (Soonthon-Brant et al., 1999). Isoflurane has been reported to enhance post-insult cerebral blood flow (CBF), produce widespread increases in brain surface PO₂ and reduce edema in a rabbit model of focal cryogenic lesion. Conversely, both CBF and regional PO₂ were decreased, and edema was increased, in rabbits anesthetized with fentanyl (Murr et al., 1993, 1995). Using a fluid-percussion model in cats, DeWitt et al, addressed the question of whether fentanyl was detrimental in TBI and found that fentanyl produced no adverse effects compared to vehicle. However, in that study, fentanyl was administered to cats already anesthetized with isoflurane (Bedell et al., 1998).

To our knowledge, isoflurane has not been directly compared to fentanyl in a contemporary model of TBI with long-term functional outcome and histologic assessment. We hypothesized that isoflurane would be neuroprotective compared to fentanyl when administered before and early after TBI. To test our hypothesis, we directly compared fentanyl and isoflurane anesthesia in a controlled cortical impact (CCI) model of TBI.

Materials and Methods

Virus-free, mature male (280 - 400g, total n=51) Sprague-Dawley rats were used in this study. The rats had free access to food and water before and
after surgery. All studies were approved by the University of Pittsburgh Animal Care and Use Committee. All surgical procedures were performed using aseptic technique.

**Outcome Protocol**

Rats were initially anesthetized with N₂O:O₂ (2:1) and 4% isoflurane (IsoFlo, Abbott Laboratories, North Chicago, IL) via a nose cone and then endotracheally intubated with a 14-guage angiocatheter and mechanically ventilated. Anesthesia was maintained for the duration of surgical preparation with 2 - 2.5% isoflurane and N₂O:O₂ (2:1). Femoral venous and arterial vessels were cannulated for continuous blood pressure measurement, blood sampling, and administration of medications. Pancuronium bromide (0.1 mg/kg/h, Elkins-Sinn, Cherry Hill, NJ) was given intravenously for muscle relaxation. A rectal probe was inserted to monitor core temperature. The rat was then placed in a stereotaxic frame (David Kopf, Tujunga, CA) and a left parietal craniotomy (7mm x 8mm) was performed using a high-speed dental drill. The dura and bone flap were left in place until immediately before CCI. A burr hole was drilled into the left frontal bone for temperature probe (2.28-mm outside diameter, Physiotemp Corp., Clifton, NJ) placement into the frontal lobe. Continuously monitored physiologic parameters included mean arterial blood pressure (MAP) and rectal and brain
temperatures. Parameters monitored intermittently included blood glucose, hematocrit, and arterial blood gas samples, which were assessed every 15 minutes for the initial hour and every 30 minutes thereafter. Throughout the experiment, PaCO₂ was controlled at 35 - 45 mm Hg. This protocol produced a PaO₂ of greater than 70 mm Hg in all preparations. Both brain and rectal temperatures were maintained at 37.0 ± 0.5°C.

Rats were allowed to stabilize for 5 min after completion of surgical preparation and then randomized to receive either fentanyl or isoflurane anesthesia. In the fentanyl group (n = 9), isoflurane was discontinued and 10 µg/kg of fentanyl (50 µg/ml, Elkins-Sinn, Cherry Hill, NJ) was administered intravenously, followed by a continuous intravenous infusion at 50 µg/kg/h. In the isoflurane group (n = 9), inspired isoflurane concentration was reduced to 1%, and normal saline, the vehicle for fentanyl-treated rats, was administered to match the volume received by fentanyl infusion. Both anesthetic groups continued to receive N₂O:O₂ (2:1). After 30 min of equilibration, TBI was induced by CCI using a pneumatic-driven piston device that has been shown (with isoflurane anesthesia) to deliver a reliable and reproducible degree of injury with a mortality rate of less than 5% (Dixon et al., 1991; Kochanek et al., 1995). In pilot studies comparing isoflurane and fentanyl using our standard CCI model (6-mm tip, 4 m/s velocity, 50 msec duration of deformation and 2.5-mm deformation depth),
all of the fentanyl-treated rats developed pulmonary edema and died early after injury. Thus, to compare the effect of isoflurane vs fentanyl on long-term outcome in our model, our standard injury was reduced (6 mm tip, 4 m/s velocity, 50 msec duration of deformation and 2.0-mm deformation depth). After CCI, the bone flap was replaced and sealed with dental cement, and the scalp incision was closed. Anesthesia was continued for 4 h in the isoflurane group and 3.5 h in the fentanyl group to facilitate similar extubation times (Miura et al., 1998). At the end of the anesthetic period, rats received 100% oxygen, were allowed to awaken and resume spontaneous breathing, and were then extubated and returned to their cages. Sham rats underwent identical preparation and anesthesia, but no CCI (n = 6 per anesthetic group).

Motor function, including beam balance and beam walking tasks, was tested by an observer blinded to group assignment on days 1 - 5 after injury (Dixon et al., 1987, 1991). Briefly, in the beam balance task, the rat was placed on a suspended, narrow wooden beam (1.5-cm wide) and the time that the rat remained on the beam was recorded (up to 60 sec). For beam walking, the rat was placed at one end of the beam and a dark, quiet chamber was located at the other end. An adverse stimulus of loud white noise was applied and the time for the rat to escape across the beam into the chamber was recorded (up to 60 sec). Rats were
trained with three trials before CCI or sham injury, which also served as baseline values.

Morris Water Maze (MWM) testing was performed using an acquisition paradigm on days 14 - 20 after injury (Hamm et al., 1992). Briefly, on post-injury days 14 - 18, the rat was placed into a pool (2-m diameter) and required to locate a hidden platform in order to escape the water. On post-injury days 19 and 20, the platform was raised so that the surface was visible 5-cm above the water level. Latency to find the platform was used to compare performances. Swim speed was measured on post-injury day 20 to insure that rats in all experimental groups exhibited equal motivation and motor function.

Lesion volume and hippocampal neuronal survival were assessed on day 21 after injury (Clark et al., 2000; Kochanek et al., 1995). Rats were re-anesthetized and then perfused with heparinized saline followed by 4% paraformaldehyde. Brains were removed, post-fixed and cryoprotected. Serial coronal sections (10-μm) were made at 1-mm intervals through the entire brain. Sections were mounted on slides and stained with cresyl violet. The areas of both tissue loss and the entire uninjured hemisphere were determined by an observer blinded to experimental group using an image analysis system (MCID, Imaging Research, St. Catherines, Ontario, Canada). Lesion volume was reported in cubic mm and as a percentage of uninjured hemisphere. Surviving hippocampal
neurons were counted under 400X magnification in the entire anatomic CA1 and CA3 hippocampal regions in a coronal section taken 5-mm from the occiput by an observer blinded to experimental group (KS). Neuronal counts were reported as the mean number of surviving neurons per 400X field.

ICP Protocol

Based both on results of the outcome protocol showing that fentanyl-treated rats had higher MAP throughout the experiment and on a recent report that increased MAP may exacerbate injury after TBI (Kroppenstedt et al., 1999), ICP and brain water were monitored in a separate cohort of rats (n = 9 per anesthetic group) subjected to either fentanyl or isoflurane anesthesia and CCI in an identical paradigm to that used to assess functional outcome.

Surgical preparation, randomization, anesthetic administration, and CCI were identical to the outcome protocol, with minor exceptions. Specifically, an intraparenchymal ICP monitor (Codman microsensor transducer, outer diameter 1.0-mm, Johnson and Johnson, Raynham, MA) was inserted through a burr hole in the frontal bone into the contralateral (right) frontal cortex at the time of craniotomy. After CCI, anesthesia was continued and ICP was monitored for 4 h in both anesthetic groups. Cerebral perfusion pressure (CPP) was calculated as the difference between MAP and ICP. Rats were killed by decapitation at the end
of the anesthetic period. Brains were immediately removed and a 3-mm coronal slice was made through the center of the contusion using a rat brain slicer. Percent brain water was determined in the coronal slice using the wet-dry weight method (Kochanek et al., 1995). The section was weighed immediately, dried in an oven at 110°C for 48 hours and then reweighed. Brain water content was determined in both the injured and the homologous region of the uninjured hemispheres.

As an additional control, a separate cohort of rats \((n = 3)\) was subjected to CCI and allowed to recover without anesthesia. These rats were prepared for CCI under isoflurane anesthesia as described, allowed to recover a tail-pincha response and then subjected to CCI. MAP was monitored via an indwelling femoral arterial catheter for 4 h during recovery without anesthesia.

Statistical Analysis

Physiological parameters (\(\text{PaCO}_2\), \(\text{PaO}_2\), glucose, hematocrit, MAP, ICP, CPP) and beam balance, beam walking, and MWM latencies were assessed by two-way analysis of variance for repeated measures. Tukey’s test was used in all post-hoc comparisons involving repeated measures. Swim speed, brain water content, and hippocampal neuronal survival were compared by one-way analysis of variance. The Student-Newman-Keuls test was used for post-hoc comparisons
after one-way analysis of variance. Time to extubation and lesion volume were compared between treatment groups using unpaired student’s t-test. A p value < 0.05 was considered statistically significant. Values are expressed as mean ± SEM.

Results

Outcome Protocol

Average time to extubation did not differ after injury between isoflurane and fentanyl treatment groups (269 ± 21 min vs 275 ± 45 min, p = 0.29). Physiologic values, including PaCO₂, PaO₂, blood glucose and hematocrit did not differ between anesthetic groups. In contrast, MAP was higher in injured rats treated with fentanyl compared to their isoflurane counterparts (p < 0.05) during the entire post-trauma period (Figure 1). Similarly, MAP was higher in shams treated with fentanyl vs isoflurane (p < 0.05) during the entire duration of anesthesia (Figure 1). Fentanyl-treated rats had a MAP of approximately 150 mm Hg compared to approximately 105 mm Hg in the isoflurane groups.

Rats anesthetized with isoflurane performed better on beam balance and beam walking tasks after TBI compared to their fentanyl counterparts (p < 0.05, Figure 2). Following injury, isoflurane-anesthetized rats also performed better than their fentanyl-treated counterparts during MWM testing with a hidden
platform ($p < 0.05$, Figure 3). Motor and MWM performances did not differ between sham groups. All experimental groups showed improved MWM performance during visible (vs hidden) platform testing ($p < 0.05$ for all groups) and had similar swim speeds (Figure 3), indicating that all rats had similar motivation and motor ability during MWM testing. However, performance on the visible platform paradigm of the MWM was better in isoflurane vs fentanyl treated rats after TBI ($p < 0.05$, Figure 3). This suggests that the difference in MWM performance may not be solely attributable to cognitive deficits.

Lesion volume, expressed as mm$^3$ or as percent of uninjured hemisphere, at 21 days after TBI did not differ significantly between isoflurane and fentanyl treatment groups (Figure 4A). In contrast, neuronal counts in the injured CA1 hippocampus were markedly greater in isoflurane-treated rats ($p < 0.05$, Figure 4B). Neuronal counts in the injured CA3 hippocampus did not differ significantly between treatment groups (Figure 4C). In the uninjured hemisphere, neuronal counts in both CA1 and CA3 hippocampus did not differ between isoflurane- and fentanyl-treated rats, and were similar to shams (data not shown).

**ICP Protocol**

Physiologic values, including PaCO$_2$, PaO$_2$, glucose and hematocrit, did not differ between anesthetic groups. As in the outcome protocol, MAP was
higher in rats treated with fentanyl compared to their isoflurane counterparts ($p < 0.05$, Figure 5A). ICP was similar in both anesthetic groups; however, there was a trend toward higher ICP in rats anesthetized with isoflurane by 3 - 4 h after TBI (Figure 5B). This strongly suggests that the higher MAP in fentanyl vs isoflurane treated rats did not exacerbate intracranial hypertension. As expected from the difference in MAP, CPP was significantly higher in the fentanyl treatment group (Figure 5C). Brain water content, assessed at 4 h after injury, was higher in the injured vs uninjured hemisphere ($p < 0.05$) for both anesthetic groups (Figure 6). However, brain water in either the injured or uninjured hemisphere did not differ between isoflurane- and fentanyl-treated rats, indicating that edema was not exacerbated in the fentanyl group.

Average MAP during the 4 h post-injury observation period did not differ significantly between fentanyl-treated rats and those recovering from CCI without anesthesia (157 ± 18.6 mm Hg vs 147 ± 12.3 mm Hg). In contrast, isoflurane-anesthetized rats had lower MAP (105 ± 16.5 mm Hg) compared to both fentanyl-treated rats and rats recovering without anesthesia ($p < 0.05$ vs both groups, one-way ANOVA).
Discussion

Isoflurane anesthesia administered to rats subjected to CCI improved performance on both motor and MWM tasks and attenuated damage to CA1 hippocampus after TBI. Although fentanyl-treated rats had higher MAP and CPP than their isoflurane counterparts, this was not accompanied by increased ICP or brain water during the first 4 h after TBI. This suggests that the improved functional outcome in rats anesthetized with isoflurane may be a direct result of either beneficial actions of isoflurane, and/or detrimental effects of fentanyl.

The pathophysiology of TBI includes a primary injury caused by the mechanical disruption of tissue and various secondary injuries mediated, at least in part, by post-insult hypoperfusion, ischemia, and excitotoxicity (Bryan et al., 1995; Hendrich et al., 1999; Hovda et al., 1995; Kochanek et al., 1995; McIntosh, 1993). Isoflurane anesthesia may be neuroprotective vs fentanyl by decreasing excitotoxicity and/or augmenting cerebral blood flow.

Following TBI, interstitial levels of excitatory amino acids (EAAs), such as glutamate, are increased due to direct tissue injury and secondary ischemia. EAAs stimulate NMDA, AMPA/kainate, and metabotropic receptors, leading to neuronal membrane depolarization, cellular swelling, calcium influx, and ultimately, neuronal death (McIntosh et al., 1999). Although models of cerebral ischemia have shown conflicting effects of isoflurane on glutamate levels in blood
and brain interstitial fluid (Patel et al., 1995; Stover et al., 1999), isoflurane has been shown to inhibit glutamate receptors, reduce NMDA-mediated calcium influx and delay neuronal injury induced by cerebral ischemia (Bickler et al., 1994; Patel et al., 1998).

To attribute the potential neuromodulatory actions of isoflurane on neuronal injury to only glutamate toxicity (Bickler et al., 1994) and glutamate signaling transduction (Dildy-Mayfield et al., 1996), however, is an oversimplification since isoflurane has many neural actions that could contribute to neuroprotection. Some include the inhibition of some voltage sensitive potassium channels (Kamatchi et al., 1999), the activation of specific receptor-coupled and voltage sensitive potassium channels (Magyar and Szabo, 1996; Winegar et al., 1996; Zorn et al., 1993), the uncoupling of muscarinic receptors (Anthony et al., 1989; Minami et al., 1997; Nietgen et al., 1998), the enhancement of GABA A channels (Lin et al., 1992), and the reduction of intracellular calcium stores and inhibition of IP3 sensitive intracellular calcium release (Hossain et al., 1991). Potential detrimental actions such as the enhancement of NMDA linked nNOS activation have also been documented (Rengasamy et al., 1997); however, the potential beneficial actions of isoflurane on excitotoxic cascades far outweigh the potential detrimental actions.
Additionally, isoflurane is a potent cerebral vasodilator. Studies of CBF after experimental TBI have shown significant reductions in both local and global CBF early (0.5 – 4 h) after injury (Bryan et al., 1995; Hendrich et al., 1999). Effects are greatest near the impact site, but global reductions in CBF are seen as well (Hendrich et al., 1999). In clinical studies, early post-traumatic hypoperfusion has been strongly correlated with poor outcome (Bouma and Muizelaar, 1992; Marion et al., 1991). Although the effects of fentanyl on CBF have been subjected to limited study, Safo et al (1985), have reported that CBF was markedly reduced in rats treated with fentanyl (100 µg/kg iv) vs control rats anesthetized with N₂O. In addition, using perfusion MRI in normal rats anesthetized with doses identical to those used in this study, we have shown that CBF is 2 - 3 times higher in rats treated with isoflurane vs fentanyl (unpublished data). By promoting CBF, isoflurane may help attenuate post-traumatic hypoperfusion, reducing secondary injury and improving recovery. The selective neuroprotection of CA1, but not CA3, hippocampus in isoflurane- vs fentanyl-treated rats is consistent with this concept. Additionally, another CBF promoting strategy, L-arginine, has recently been shown to improve outcome after TBI (Cherian et al., 1998; DeWitt et al., 1997). Augmentation of CBF with an associated increase in cerebral blood volume therefore offers a potential explanation for both improved functional and histological outcome and the
tendency toward higher ICP and brain water seen in isoflurane- vs fentanyl-treated rats in this study. Indeed, the combination of CBF promotion and reduced excitotoxicity may be particularly beneficial.

Alternatively, fentanyl may be detrimental after TBI. Opioids generally suppress neuronal excitability, however, mu receptor agonists, such as fentanyl, may contribute to hippocampal neuron excitation (Bradley and Brookes, 1984; Neumaier and Chavkin, 1986; Neumaier et al., 1988). In fact, high-dose fentanyl (25 - 100 μg/kg in humans and 400 μg/kg in rats) has been associated with subcortical seizures (Faden et al., 1985; Kearse et al., 1993; McIntosh et al., 1986; Ohta et al., 1995; Safo et al., 1985; Tempelhoff et al., 1992). Although fentanyl exhibits low affinity for kappa receptors (Ohta et al., 1995), kappa receptor antagonists have been shown to improve both neurological outcome after spinal cord injury and CBF after fluid-percussion injury in cats (Faden et al., 1985; McIntosh et al., 1986).

Other factors may contribute to improved outcome in isoflurane- vs fentanyl-treated rats. These include the disparity in both MAP and CPP and possible differences in the depth of anesthesia between treatment groups. Although increased MAP can have detrimental effects after TBI, in our study, ICP and brain water were not significantly different in isoflurane vs fentanyl treatment groups at the end of the 4 h treatment period. This suggests that the higher MAP
associated with fentanyl vs isoflurane anesthesia had no acute detrimental effects on intracranial hypertension or brain edema. Additionally, MAP did not differ significantly between rats treated with fentanyl and those allowed to recover without anesthesia. The values for MAP observed in both isoflurane and fentanyl groups were within the reported range of cerebral autoregulation (50 - 170 mm Hg) for normotensive rats (Hernandez et al., 1978; Hoffman et al., 1991; Zaharchuk et al., 1999). The disparity in MAP is therefore unlikely to have contributed importantly to the observed difference in functional outcome. Although a recent study using the CCI model in rats suggests that increased MAP and CPP may exacerbate injury after TBI, in that study, systemic hypertension was induced by large doses of dopamine (Kroppenstedt et al., 1999). Higher doses of dopamine may have produced detrimental effects after injury which were independent of blood pressure (Beaumont et al., 1999). Additionally, our study does not address the important detrimental effect of hypotension following clinical TBI, particularly with multiple trauma. The blood pressure supporting effects of fentanyl or its derivatives (vs other sedative agents) may be beneficial in this section (Tipps et al., 2000).

Although comparison of anesthetic depth between inhalation and intravenous agents is difficult, differences in anesthetic depth are unlikely to explain the observed difference in outcome seen between isoflurane- and fentanyl-
treated rats. The dose of isoflurane used (1% by inhalation), in combination with
\[ \text{N}_2\text{O}:\text{O}_2 \ (2:1) \]
represents approximately 1.2 minimal alveolar concentration (Hansen et al., 1989). The dose of fentanyl falls in the range between the ED50 for purposeful movement and complete blockade of this response (Kissin et al., 1983) and is similar to standard doses used in rat and cat models of central nervous system injury (Bedell et al., 1998; Miura et al., 1998; Murr et al., 1993, 1995; Soonthon-Brant et al., 1999). Additionally, fentanyl-treated rats did not exhibit signs of increased stress vs isoflurane-treated rats, such as higher blood glucose, suggesting that anesthesia was adequate. However, there is a massive catecholamine surge immediately after TBI (Rosner et al., 1984), and it is likely that this response is more effectively blunted by isoflurane than fentanyl (Mackensen et al., 1999). Finally, rats in both treatment groups emerged from anesthesia similarly in our paradigm. After discontinuation of fentanyl, the rats were fully alert and exhibited similar activity to isoflurane-treated rats following extubation.

The theoretical advantages of isoflurane vs fentanyl are compelling; however, explanations for the observed improvement in neurologic outcome after TBI in isoflurane-anesthetized rats remain to be determined. What has become clearer is that anesthetic agents may have considerable impact upon outcome following TBI.
The results of this study suggest two important potential ramifications. First, isoflurane may not represent the optimal anesthetic in experimental TBI since it may mask potential benefits of novel therapies. Second, despite common use, fentanyl may not be the optimal sedative/analgesic agent to administer to patients in the acute phase after severe head injury. Although we do not suggest that isoflurane represents a clinically applicable therapy for the initial stabilization and treatment of patients after TBI, further study of the mechanistic differences between isoflurane and fentanyl anesthesia is warranted. Defining the factors responsible for improved outcome with isoflurane may help to direct the clinical application of more optimal sedative/analgesic agents and possibly to identify novel therapies. Finally, more comprehensive comparisons of clinically relevant sedative/analgesic agents are needed in experimental TBI.

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References


**Figure 1:** MAP vs time after injury, outcome protocol. MAP in both fentanyl-treated injured (○) and sham (▲) rats was approximately 50 mm Hg higher than in isoflurane-treated injured (●) and sham (▲) rats at all time points. *p < 0.05, isoflurane vs fentanyl at each time point after injury, § p < 0.05, isoflurane vs fentanyl at all time points, including baseline, in shams.

**Figure 2:** A. Beam balance latency vs time in days after injury. Sham rats (▲ = fentanyl, ▲ = isoflurane) had similar latencies throughout the 5-day testing period. After injury, beam balance latency was shorter for both isoflurane (●) and fentanyl (○) treatment groups vs sham; however, isoflurane-anesthetized rats recovered more quickly (vs fentanyl). *p < 0.05, injured vs sham. B. Beam walking latency vs time in days after injury. Symbol designations are identical to those used in (A). Again, performance was impaired after injury in both anesthetic groups vs shams. Although isoflurane-treated rats recovered by post-injury day 3, fentanyl-treated rats failed to regain normal function by the end of the 5-day testing period. *p < 0.05, injured vs sham.

**Figure 3:** A. Latency to find a platform vs time after injury in an acquisition paradigm of the MWM. Sham rats anesthetized with isoflurane (▲) or fentanyl (▲) had similar performances throughout the testing period. During the first few
days of testing with a hidden platform, injured rats in both fentanyl (○) and isoflurane (●) treatment groups had impaired performance vs sham. By the third day of testing, latencies to find the hidden platform were similar in injured isoflurane-anesthetized rats and shams. In contrast, longer latencies persisted in injured fentanyl-treated rats throughout the 5-day hidden platform testing period. Latencies in all experimental groups improved during visible (vs hidden) platform testing; however, shams and injured isoflurane-anesthetized rats performed better than injured fentanyl-treated rats on both days of visible platform testing. *p < 0.05, injured vs sham; $p < 0.05$, isoflurane vs fentanyl. B. Swim speed, tested on day 20 after injury, did not differ between experimental groups.

**Figure 4:** A. Lesion volume, measured on post-injury day 21, did not differ significantly between isoflurane (■) and fentanyl (□) treatment groups. B. Neuronal counts in injured CA1 hippocampus were greater in isoflurane- (■) vs fentanyl-treated (□) rats. Neuronal counts in uninjured CA1 hippocampus were similar in both treatment groups. *p < 0.05, injured vs uninjured. C. CA3 hippocampal neuronal counts in either injured or uninjured hemisphere did not differ significantly between isoflurane- (■) and fentanyl-treated (□) rats.
**Figure 5:** A. MAP vs time after injury, ICP protocol. MAP was approximately 40 mm Hg higher in rats treated with fentanyl (□) vs isoflurane (■) throughout the observation period. * p < 0.05, fentanyl vs isoflurane vs fentanyl. B. ICP vs time after injury. Initial ICP was approximately 4 mm Hg in both isoflurane (■) and fentanyl (□) treatment groups. ICP progressively increased, reaching 10 - 18 mm Hg by 4 h after injury. Although ICP was similar between anesthetic groups, isoflurane-anesthetized rats exhibited a trend toward higher ICP after injury (vs fentanyl treated rats) that did not reach significance. C. CPP vs time after injury. CPP was increased in rats treated with fentanyl (□) vs isoflurane (■), * p < 0.05, isoflurane vs fentanyl at all time points except 3.5 and 4h.

**Figure 6:** Brain Water 4 h after TBI. Brain water in the injured hemisphere was increased compared to the respective non-injured hemisphere in both isoflurane- and fentanyl-anesthetized rats; however, brain water did not differ between anesthetic groups. * p < 0.05, isoflurane vs fentanyl.
CHANGES IN MITOCHONDRIAL MEMBRANE POTENTIAL IN STRETCH-INJURED ASTROCYTES AND NEURONS: S.M. Ahmed*, B.A. Rzalinski and E.P. Ellis. Dept. of Pharmacology and Toxicology, Medical College of Virginia, Virginia Commonwealth University, Richmond, VA.

The dynamics of energy failure in traumatically injured astrocytes and neurons are unclear. In order to better understand mitochondrial function and cell energetics following trauma we utilized the fluorescent dye Rhodamine 123, which is normally sequestered in mitochondria where its fluorescence is quenched. When mitochondrial membrane potential (MMP) decreases, such as with mitochondrial poisons, the dye moves to the cytoplasm and fluorescence is increased. Pure neuronal or astrocytic cultures were subjected to mild (5.7 mm), moderate (6.5 mm) or severe (7.5 mm) stretch-induced injury and the change in MMP measured. There were no significant changes in MMP in mildly to moderately injured neurons at 15 min, 24 or 48 hr post-injury. However severely injured neurons displayed an immediate 33% decrease in MMP that persisted to 48 hr. In contrast, mild and moderate astrocyte injury caused a dramatic, 39-52% drop in MMP at 15 min, with MMP returning to normal by 24 hr. Our results indicate that direct trauma-induced alterations in cell energetics vary greatly in neurons and astrocytes. We suggest that in vivo the deficit induced in astrocytes may alter astrocyte function, which in turn may produce dramatic effects on neuronal function. Supported by NS-27214 and NS-07288.

EVIDENCE FOR APOPTOTIC CELL DEATH FOLLOWING SUBDURAL HEMATOMA IN RATS. B. Alessandri*, X. Di, H. Chen, R. Bullock. Div. of Neurosurgery, Medical College of Virginia, Richmond, VA, 23298, USA.

Subdural hematoma (SDH) is a common and dangerous secondary event following traumatic brain injury. The mechanisms leading to neuronal death, even after SDH removal, are not fully understood. A mechanism which might contribute to cell death is apoptotic cell death (ACD), which has been shown to be involved in the development of traumatic and ischemic brain damage. The hematoma was produced by subdural injection of 250μL of autologous venous blood in Halothane anesthetized rats. Animals were allowed to survive 1 (n=3), 2 (n=3), 4 (n=3) or 7 days (n=4) after injection of SDH. Brain sections were stained by a commercially available apoptosis detection kit (TUNEL) for apoptotic cells by immunohistochemistry and light microscopy. The number of ACD-positive cells was counted in both hemispheres in cortical, subcortical and hippocampal areas. The ACD-pos. cell counts were 2.3±2.4, 5.4±4.8, 13.7±8.8, and 12.8±3.1 at 1, 2, 4 and 7 days after SDH, respectively. All ACD-positive cells were within the cortex, within and in the border zone of the SDH lesion. There were no ACD-pos. cells in the contralateral side. The number of ACD-pos. cells was highly correlated with the lesion area (r²=0.689, p<0.001).

The results indicate that ACD occurs following SDH, and is maximally seen at 2 days. ACD-pos. cells were only found within or in the border zone of the lesion. The correlation of ACD and lesion area underlines the importance of this type of cell death in SDH. The contribution of ACD to SDH-induced brain damage and its relevance for therapy needs further study.

HYPERTHERMIA ADVERSELY AFFECTS OUTCOME AFTER MODERATE HEAD INJURY. Philipp R. Aldana*, J. Marquez*, D.S. Petrin*, D. Johns*, W.D. Dietrich*, P.A. Villaneuva. Department of Neurological Surgery, Neurotrauma Research Center, University of Miami School of Medicine, Miami, Florida, 33101, USA.

Hypothermia has been shown to have beneficial effects after traumatic brain injury (TBI) in both human and animal studies. Conversely, hyperthermia after TBI has been shown to have deleterious effects in animals. No studies have addressed the effects of hyperthermia after moderate head injury in humans.

104 patients admitted with a Glasgow Coma Score 9-12 due to blunt head trauma were studied. Demographics, comorbid factors and characteristics of the hyperthermic episodes (>38.6°C) were examined. The number of patients either dead, in a vegetative state or severely disabled during discharge was significantly larger for the hyperthermic group vs. the normothermic group (42.4% vs. 17.5%, respectively). A significantly larger percentage of the normothermic group had a good outcome compared to the hyperthermic group (50% vs. 20.3%, respectively). Among the hyperthermic patients, those with associated infections had significantly worse outcomes and a higher frequency of hyperthermic episodes than those without infections. We conclude that hyperthermia in the face of an associated infection may adversely affect the outcome of patients with moderate head injury. We advocate maintenance of at least normothermic conditions if moderate hypothermia cannot be achieved and treatment of any underlying infection after TBI.

VERTICAL VERSUS ANGLED CONTROLLED CORTICAL IMPACT IN RATS. H.L. Alexander*, C.L. Robertson, C.E. Dixon, R.S.B. Clark, S.H. Graham, P.J. Safar, P.M. Kochanek. Safar Center for Reanimation Research, Univ. of Pittsburgh, PA 15213

Although a variety of modifications of the controlled cortical impact (CCI) model exist, a comparison between the two most common variants, vertical and angled impact, has not been performed. Rats were subjected to vertical (n = 8), angled (n = 8) or sham (n = 8; 4 m/s, 2.5 mm) to the left parietal cortex, using a CCI model with hypoxia. Motor (beam balance, d1-5), cognitive (Morris Water Maze, d14-21) and histologic (lesion volume, CA1 and CA3 neuron counts, d21) outcomes were studied. Motor and MWM performance were impaired, but did not differ between injury groups. Lesion volumes also did not differ (vertical = 92.2±7.2 mm³, angled = 79.4±7.8, p = 0.25). CA1 neuron counts were decreased ipsilaterally to injury in both groups vs sham (vertical = 20.4±5.8 cells/hpf, angled = 32.7±15.8, sham = 55.5±3.9, p < .05). However, CA3 neuron counts were decreased ipsilaterally to injury in the vertical group vs sham (23.2±8.5 vs 32.1±6.6, respectively, p < .05), but the angled group (32.7±15.8) was not different from sham. We conclude that the vertical and angled variants of the CCI model produce similar functional deficits; however, the vertical impact appears to produce greater local damage, particularly in CA3 neurons.
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EFFECTS OF SEVEN ANESTHETICS ON OUTCOME IN EXPERIMENTAL TRAUMATIC BRAIN INJURY
Safar Center, University of Pittsburgh, Pittsburgh, PA 15260

We have shown improved outcome with isoflurane vs fentanyl in the rat controlled cortical impact (CCI) model. We now present the first comprehensive study of the effect of anesthetics on outcome in experimental TBI. Rats were prepared for CCI under isoflurane and allowed to recover to tail pinch. After CCI, rats were randomized to 1 h of either isoflurane, fentanyl, morphine, pentobarbital, diazepam, ketamine, or propofol treatment or recovery without additional anesthesia (n=9/group). Brain temperature, mean arterial blood pressure (MAP), blood gases, and glucose were monitored. Motor function was assessed on d 1-5 after CCI. MAP was higher in fentanyl vs isoflurane treated rats (p < 0.05), but did not differ between other groups. After CCI, beam balance and beam walking performances were worse for rats treated with morphine or propofol (p < 0.05 vs sham). Rats administered isoflurane only before injury did as well as those given isoflurane for 1h post-trauma. Taken in context of our prior study showing improved outcome with isoflurane vs fentanyl before TBI, these data suggest benefits of isoflurane near the time of impact. Considering their common clinical use, the detrimental effects of morphine or propofol deserve further characterization. Cognitive and histological assessment in these animals is ongoing.

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CHRONIC OVEREXPRESSION OF AMYLOID PRECURSOR PROTEIN (APP) AFTER TRAUMATIC BRAIN INJURY IN RATS. J. R. Ciallella, H. Q. Yan, X. Ma, D. W. Marion, S. T. DeKosky, and C. E. Dixon. Departments of Neurosurgery and Psychiatry, University of Pittsburgh Medical Center, Pittsburgh, PA USA. Traumatic brain injury (TBI) and Alzheimer’s disease (AD) produce cholinergic and metabolic deficits that may contribute to neurodegeneration. There is increasing evidence linking AD and TBI, including upregulation of APP in head injured patients (McKenzie et al. 1994 NeuroReg.6:161). To further investigate this linkage, we tested the hypothesis that controlled cortical impact (CCI) injury would produce chronic upregulation of APP protein levels at 4 weeks following injury. Our previous studies demonstrated significant changes in cholinergic proteins at this time point (Ciallella et al. 1998. Exp. Neurol. In Press). APP immunohistochemistry (n=3-5) and western blot (n=4) were performed on cortical and hippocampal regions from injured and sham animals. The same N-terminal antibody was used in all studies. A marked increase in cortical and hippocampal APP protein was demonstrated bilaterally by both immunohistochemistry and western blot in injured rats compared to sham controls. This demonstrates that a single TBI can lead to chronic upregulation of APP, concurrent with chronic alterations in cholinergic markers. Supported by AG05133, NINDS- T32NS07391, CDC-CCR312296, NIH-NS30313, and NIH-NS33150.

THE SUPPRESSION OF HIPPOCAMPAL NGF mRNA AFTER CEREBRAL ISCHEMIA IN RAT TREATED WITH ANTISENSE DNA TO C-fos, E.K. Cui, C.Y. Hsu, and P.K. Lin. Department of Neurosurgery, Baylor College of Medicine, Houston, TX 77030; Department of Neurology, Washington University, St Louis, MO 63110. The biological effects of Fos expression in the brain were examined using phosphorothioated oligodeoxynucleotides (s-ODNs) to c-fos, mcfosf15, Bisubinylated antisense mcfosf15 (bio-mcfosf15) plus lipofectin were delivered into the brain of male Long-Evans rats (225-250 gm) via intracerebroventricular infusion. The distribution of bio-mcfosf15 was detected using antibodies against biotin. Using dot blot analysis on the recovered bio-mcfosf15, the bio-mcfosf15 uptake in hippocampus peaked at 24-48 hrs, and the internalized bio-mcfosf15 was degraded within 72 hr of infusion. The s-ODN uptake in the brain was confirmed by 3'-end-labeling with digoxigenin-DUTP, using terminal transferase and anti-digoxigenin IgG-FITC. The presence of fluorescent aggregates in the brain cells near the vessel wall in animals treated with antisense mcfosf15 plus lipofectin suggests lipofectin mediated s-ODN transfer across the blood brain barrier. The uptake increased with time and with the dose delivered. The effectiveness of antisense mcfosf15 was shown by an inhibition of ischemia-induced Fos expression, and was accomplished by an inhibition of ischemia-induced hippocampal NGF mRNA expression in the brain of animals pretreated with antisense mcfosf15. The specificity of Fos suppression was suggested by a lack of antisense mcfosf15 effect on the expression of NT-3 and α-actin mRNA.

LOSS OF GLIAL POTASSIUM CURRENTS AND IMPAIRMENT OF POTASSIUM HOMEOUSTASIS, FOLLOWING FLUID PERCUSSION INJURY. R. D’Ambrosio, D.O. Matj, M.S. Brady, and D. Jurany. Dept. of Neurosurgery, Unlv. of Washington, Seattle, WA 98104. We compared the early effects of moderate in vivo fluid percussion injury (FPI) on the functional expression of potassium currents expressed in oligodendroglia and astrocytes from acutely isolated rat hippocampal slices. Whole cell recordings were performed from post-FPI and naive slices of 30 d.o.r. A C4 (1 mM) was used to block inward potassium currents. K+-selective electrodes were employed to measure K+ accumulation in radiatum CA3. GFAP immunostaining was enhanced in CA3 24-48 hrs following FPI, while immunostaining for oligodendroglia was reduced. A significant decrease in C4-sensitive potassium currents was observed following lesion in both oligodendroglia and astrocytes. Cells characterized by complex electrophysiological profiles as well as those characterized by inward rectification were equally affected (60% and 55% at -140 mV). Morphologically, complex cells visualized by biocytin staining could be classified as oligodendrocytes. Stimulation (1 Hz) of Schaffer collaterals induced K+ accumulation in radiatum CA3. Slices obtained from naive rats always showed a recovery of extracellular K+ basal levels within 10 seconds following stimulation (n=5). Slices obtained from post-FPI rats displayed recovery times ranging from 10 to 40 seconds (n=8). Additionally, 75% of the post-FPI slices generated multiple afterdischarges during stimulation, while only 20% of the control slices did. These results indicate that 1) post-FPI CA3 astrocytes are reactive or injured, 2) loss of C4-sensitive potassium current occurs in oligodendrocytes and astrocytes post-FPI, 3) neuronal-activity-induced elevation of [K+]out is more persistent at early time point post-FPI, 4) hyperexcitability is observed after trauma without detectable neuronal loss. We conclude that FPI may affect extracellular K+-homeostasis by impairing glial potassium currents. Supported by NIH-51614 and RO-1 NS3107.
EFFECT OF CALCIUM CHLORIDE ON REGIONAL CEREBRAL BLOOD FLOW DURING CARDIOPULMONARY RESUSCITATION IN PIGLETS

Molody Palmer Land, John Kuluz, Barry Gelman, Michael Narex, En Xu, and Charles Schlein. Pediatric Critical Care Medicine, University of Miami School of Medicine, Miami, FL 33101.

Introduction: The use of calcium chloride (CaCl₂) during CPR remains controversial. CaCl₂ may improve the effectiveness of CPR by increasing systemic vascular tone and vital organ perfusion. Alternatively, CaCl₂ may cause regional vasocostriction in the brain and heart, resulting in secondary ischemic injury. We hypothesized that administration of the standard dose of CaCl₂ during CPR decreases CBF.

Methods: Under pentobarbital anesthesia, 2-4 week old piglets underwent 6 min of cardiac arrest by ventilatory fibrillation, and 30 min of standard CPR. CBF was measured with microspheres at baseline and after 5, 15 and 30 min of CPR. CaCl₂, 20 mg/kg (n=7) or saline (n=5) was given after 1 and 19 min of CPR. Data (mean±SE) were analyzed by ANOVA and Student's t-test (*p<0.05).

Results: Ionized (i)Ca decreased from 1.40±0.03 at baseline to 1.16±0.05 iCa at 15 min and 1.18±0.05 iCa at 30 min CPR. After CaCl₂, iCa increased to 2.5±0.35 at 5 min and 2.04±0.21 iCa at 30 min, and was not different at baseline at 15 min CPR. Calcium increased aortic pressure (44±2 vs 38±2) and cerebral perfusion pressure at 5 min CPR. Total CBF was not different between groups at any time point; however, severe regional ischemia (CBF<1.5 ml/100g/min) was more common after 30 min CPR when CaCl₂ was given, particularly in subcortical regions (p<0.03).

Conclusion: These data show that CaCl₂ administration has adverse effects on CBF during prolonged CPR, and may worsen ischemic brain injury. Future studies will determine the effect of CaCl₂ on functional and neuropathologic outcomes.

50 NO LONG-TERM BENEFIT FROM HYPOTHERMIA AFTER SEVERE TRAUMATIC BRAIN INJURY WITH SECONDARY HYPOXEMIA IN RATS

Courtney L. Robertson, Robert Clark, C. Edward Dixon, Steven Graham, Henry Alexander, Stephen Winstead, Donald Marois, Peter Safer, Patrick Kochanek, Dept of Anesthesiology/CCCM, Pediatrics, Neurology, Epidemiology, and Neurosurgery, Salra Center for Resuscitation Research, University of Pennsylvania, PA 19123.

Introduction: Many reports have shown benefit from hypothermia in traumatic brain injury (TBI); but its effect in the setting of TBI with secondary insult remains undefined. Clinical studies show an increase in morbidity and mortality after severe TBI with secondary brain insult. In experimental rat models, outcomes were worse in brain injury with secondary hypoxia. Recently, we characterized a model of TBI with secondary hypoxemia and reported prominent neurologic apoptosis after injury. We hypothesized that hypothermia would improve outcome after controlled-cortical impact (CCI) with secondary hypoxia insult in rats.

Methods: Rats were subjected to severe CCI injury followed by 30 min of hypoxia (PaO₂=55±4 mm Hg). Rats were then randomized to normothermia (NT=37°C, n=19), immediate hypothermia (HT=32°C, after CCI, n=10), or delayed hypothermia (HT=32°C after hypoxia, n=14) for 4 h. Motor (beam balance/beam walking, d 1-3), cognitive (Morris Water Maze [MWM], d 14-21) and histologic outcome (lesion volume, hippocampal neuron counts, diI) were evaluated. Results: Motor and MWM performance were impaired, but did not differ between groups. Lesion volumes did not differ significantly between groups (NT=65.5±16.9, HT=56.2±8.2, HT=57.3±7.9). Hippocampal neuron counts (CA1-C3) were decreased on the injured side, but not between groups (NT=19.6±2.4, HT=18.8±2.6, HT=17.8±3.8, HT=18.5±3.3). Mortality rate did not significantly differ between groups. Conclusions: Immediate or delayed hypothermia did not improve long-term outcome after severe CCI with secondary hypoxemia in rats. The severity of the combined insult may be outside of the therapeutic window for hypothermia. Clinical studies have excluded patients with secondary insult, and have indicated that hypothermia is of limited efficacy in the subset of severely injured (GCS 3-4) patients. Novel therapeutic approaches or combination therapies may be necessary in this setting.


51 BRAIN NITRIC OXIDE CHANGES AFTER CONTROLLED CORTICAL IMPACT INJURY IN RATS

Leela Chelian, J. Clay, Goodman, Claudia S. Robertson. Departments of Neurology and Pathology, Baylor College of Medicine, Houston, TX 77030.

Introduction: The marked reduction in CBF that occurs after severe controlled cortical impact (CCI) injury in rats can be ameliorated by postinjury infusion of L-arginine. Since L-arg is a substrate for nitric oxide synthase, these studies suggest that a reduced production of nitric oxide (NO) may play a role in the CBF reduction that occurs after brain trauma. The purpose of this study was to measure brain tissue concentrations of NO after severe CCI.

Methods: 12 Long Evan rats were anesthetized with isoflurane and subjected to severe CCI (5 m/s, 3 m, 15 m/s) or sham. NO was directly measured using a NO electrode which was inserted at the site of the impact after calibration using 5-nitro-N-acetyl-D,L-penicillamine at 37°C. A microdialysis probe was inserted near the NO electrode and perfused with artificial CSF at 2 μl/min. The concentration of nitrate/nitrite was measured using a chemiluminescent method in serial 20 minute collections of dialysate. These measurements were obtained prior to injury, and for 3 hours after injury. Values were expressed as % of the pre-injury baseline values.

Results: Impact injury caused a transient increase in brain tissue NO concentrations to 178% of the baseline values in the CCI animals, compared to 98% in the sham injured animals (p=0.002). After the initial transient increase in NO, the concentrations of NO declined and reached significantly lower than that in the sham animals throughout the 3 hr study period. The results are summarized below as median (25th percentile, 75th percentile). A similar reduction in nitrate/nitrite was observed in the microdialysate.

Time after injury NO (% baseline) CCI injury (n=6) NO (% baseline) Sham injury (n=6) P value
2min 177.5 (158.97) 97.5 (94.15) 100 0.002
1hr 75.5 (50.7) 98.3 (96.11) 0.004
2hr 73 (65.79) 101 (95.14) 0.001
3hr 95.5 (92.19) 95.5 (92.19) 0.002

Conclusions: This study suggests that NO release is immediately after a severe brain injury and subsequently is found in decreased concentrations in the brain for at least 3 hours after injury. The reduction in CBF that occurs after severe CCI may be related to the reduced NO levels.

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MK-801 improves functional outcome in rats after controlled cortical impact. RA Ruppel, PM Kochnak, CE Dixon, HL Alexander, SH Graham, RSB Clark, SR Wisniewski, DW Marion, PJ Safar. Safar Center for Resuscitation Research, Univ of Pitt, Pgh, PA

Excitotoxicity is implicated as a key mechanism of secondary neuronal damage after traumatic brain injury (TBI). The NMDA receptor antagonist MK-801 has been shown to attenuate cerebral injury in focal ischemia and some models of TBI, but it has not been tested in controlled cortical impact (CCI). We hypothesized that MK-801 would improve functional and histopathologic outcomes in rats following CCI. Anesthetized Sprague-Dawley rats (n=8/gpr) were subjected to CCI (4 m/s, 2.5 mm depth), then randomized to immediate treatment with either MK-801 (1 mg/kg IP) or vehicle. Rats treated with MK-801 recovered motor function significantly earlier than vehicle controls, as shown by beam balance/walking performance (d 1-5). MK-801 treated rats also showed improvement in the probe trial of the Morris water maze (d 14-20) vs vehicle (p<0.05), but no differences were seen in latencies to target. Contusion volume and hippocampal cell counts (d 21) did not differ between the groups. These data demonstrate an important role for excitotoxicity early after cerebral contusion and support continued evaluation of anti-excitotoxic therapies for use in TBI.

Funding: DAMD17-97-1-7009

E49

PROTECTION WITH MK-801 AGAINST SUSCEPTIBILITY OF MICE EXPRESSING HUMAN APOLIPROTEIN E4 TO CA1 NEURONAL INJURY FROM TRAUMA AND OXIDATIVE STRESS. R.A. Wallis1,2, K.L. Panizzon,1,2 B. Teter,2,3 G.M. Cole,1,2,3 S.A. Frautschy,1,2,3 J.R. Gilbert,1 Dept. of Neuro., UCLA; 2Dept. of Geri., UCLA, LA, CA 90024; 3Sepulveda VAMC, Sepulveda, CA 91343; 3Dept. of Neuro., Duke University, NC 27710.

Susceptibility of CA1 neurons to trauma and oxidative stress was assessed in transgenic mice expressing the human apolipoprotein E4 gene and promoter in the ApoE knockout background. Mild trauma to hippocampal slices from ApoE knockout mice produced virtually full recovery of mean orthodromic (ortho.) and antidromic (anti.) population spike (PS) amplitude recovery of 94% ± 2 and 95% ± 1 one hr after trauma, while slices from ApoE4 mice showed ortho. and anti. PS recoveries of 16% ± 5 and 14% ± 3 (p<0.05). MK-801 32 μM treatment reversed ApoE4 susceptibility with CA1 ortho. and anti. PS recoveries of 66% ± 5 and 64% ± 3 (p<0.05). Oxidative stress (H2O2 5 μM, 6 min), to ApoE knockout mouse slices showed ortho. and anti. PS recovery after 1 hr of 89% ± 2 and 91% ± 2, while slices from ApoE4 mice showed ortho. and anti. PS recoveries of 23% ± 2 and 19% ± 2 (p<0.05). MK-801 reversed the susceptibility of ApoE4 mice to oxidative stress with ortho. and anti. PS recoveries of 83% ± 2 and 79% ± 2 (p<0.05). These findings suggest that the ApoE4 gene increases susceptibility of CA1 neurons to trauma and oxidative stress through excitotoxic mechanisms. Supported by the VA Research Service.

E50


GK-11 is a long-lasting blocker of NMDA receptors. Several studies have reported that NMDA receptor blockers reduce tissue damage. We assessed the effects of GK-11 on a well-standardized rat spinal cord contusion model, comparing 2.7 mg/kg, 0.9 mg/kg, and 0.3 mg/kg doses and vehicle started at 15 minutes or 60 minutes after 12.5 mm or 25.0 mm contusions with the NYU weight drop impactor. A total of 134 adult Long-Evan's hooded rats were studied in this study. The rats were anesthetized with intraperitoneal pentobarbital (male 60 mg/kg and female 45 mg/kg) and injured at T9-10 cord exposed with laminectomy. At 24 hours after injury, the spinal cords were rapidly removed and frozen. Six cord samples were removed from each rat and each sample was approximately 5 mm in length. To quantify tissue damage, we measured spinal cord lesion volumes, cell volume fractions (CVF), tissue Na, K concentrations and edema at and around the impact site. The results show that GK11 did not have any beneficial effects in tissue Na, K, water concentrations, cell volume fractions and lesion volumes compared to vehicle groups (p>0.05). The analyses also rule out a possible effect of GK-11 only on the surrounding cord because repeated measures analyses did not reveal any consistent treatment-related Na, K, CVF or lesion volume differences at specific sample sites. We therefore conclude that GK-11 in the dose ranges of 0.3-2.7 mg/kg given at 15 and 60 minutes after injury does not alter the cell loss in spinal cords contusion injury in the presence of pentobarbital anesthesia.

E51


Acute activation of Group I metabotropic glutamate receptors (mGluR5) contributes to traumatic brain injury (TBI) pathophysiology (J Neurosci 16:6012, 1996). We infused over 1 hr, the selective mGluR Group 1 antagonist, (RS)-1-aminooctan-1,5-dicarboxylic acid (AIDA) (0, 4, 2, 10 nmol) (n=6/group) into the hippocampus beginning at 5 min after parasagittal fluid percussion TBI. At 24 hr after TBI, coronal sections were stained with Fluoro-Jade, a fluorescent marker for neuronal degeneration. Positive staining cells were counted in 4 sections/rat. Significantly fewer (p<0.05) CA2-3 Fluoro-Jade positive neurons were detected in the 10 nmol AIDA group (184 ±32) compared to the vehicle group (310 ±47). In a second experiment, rats were administered 10 nmol AIDA or vehicle (n=10/group) after TBI as above and tested in the Morris water maze for acquisition of a spatial learning/memory task on days 11-15 post-injury. The mean swim distance for the 10 nmol AIDA-treated group (1973 ±146cm) was significantly (p<0.04) shorter than vehicle controls (1493 ±142cm). Post-injury blockade of Group I mGluR appears to reduce neuronal degeneration and improve functional outcome after TBI. Supported by NIH NS29995
216.1


Calcium influx is an important factor in the pathophysiology of traumatic brain injury. Our previous work has shown diffuse "calcium (Ca2+ ) accumulation" in the cortex immediately after lateral fluid percussion (LFP) injury lasting at least 2 days in P17 animals. In this study, in the first acute period of ischemia accumulation these studies suggested a second, delayed period of "Ca2+ accumulation" in the thalamus which correlated with the development of neurological deficits of the delayed "Ca2+ accumulation" seen at P36 P17, P34 P28, and 17 adult rats were subjected to a moderate-severe (2.75 atm) LFP. Immediately, 6 hours, 1 day, 2 days, 4 days, 7 days after injury the rats were injected with "Ca2+ (^34P) (^82P) and processed for autoradiography. Optical densities were measured in 16 regions of interest, including 6 thalamic nuclei. "Ca2+ accumulation" was evident within the first 4 days in the bilateral thalamus (immediately to 60% decrease), returning to sham levels within 4 days in all three age groups (p<0.003). In contrast, differences existed between the age groups for delayed "Ca2+ accumulation" in the thalamus. While P17 showed no delayed "Ca2+ accumulation", P34 and adult rats showed significant accumulation in the thalamus starting at 2 days and increasing out to 14 days (p<0.05 increase, p<0.03). Since histological analyses indicated cell death at the time of the delayed "Ca2+ accumulation", this delayed accumulation may represent cerebrovascular damage due to diffuse axonal injury and Ca2+ -induced apoptosis. The age-dependency of delayed "Ca2+ accumulation" in the thalamus may be attributed to differential biomechanical consequences. AP and LF injury greatly calcium buffering capacities of younger animals. The results suggest that two temporal patterns of "Ca2+ accumulation" exist following LFP: acute calcium flux associated with the injury-induced ion leak cascade and delayed calcium accumulation associated with secondary cell death. Supported by NS30538, NS27534.

216.3


A marker used to identify peroxynitrite activity following CNS injury is the 3-nitrotypoxyrine residue of proteins. Recently, a number of studies have purported measurement of 3-nitrotypoxyrine (3-NT) in brain proteins measured by HPLC. These assays vary substantially in processing, chromatographic and detection methodologies. Halliwell and collaborators (J. Neurochem. 70:2220-2222, 1998) reported an artifact of an artificial substrate in brain tissues which exhibited chromatographic, electrochemical and chemical properties nearly identical to 3-NT. It was suggested that this artifact might confound the detection of 3-NT in brain tissue. We have developed an HPLC assay for the measurement of 3-NT that circumvents the problem of artifact detection. This was accomplished by using gradient elution, ion pairing and multi-channel electrochemical detection methodology. We were able to measure, in injured brain protein digests, 3-NT as a percentage of tyrosine (3-NT/tyr) at levels much lower (0.004%) than purported (J. Cereb. Blood Flow Metab. 18:123-129, 1998, in rat, at 24 hrs after impact/acceleration head injury in rat, hippocampal 3-NT/tyr was used not different from sham animals. However, in the same model, another paper that exist very close to 3-NT increased significantly after injury. This same peak was found to increase in midline/study with rat head injury as well, with no apparent change in 3-NT. The former response was blocked by L-NAME, the non-selective inhibitor of nitric oxide synthase. We suggest that this be the same artifact reported by Halliwell and collaborators. Isolation of this peak material followed by LC-MS, LC-NMR, LC-EC and LC-UV confirms the identity as guanosine. We recommend including guanosine as an HPLC standard to avoid misidentification with 3-NT. (Supported by Warner-Lambert/Parke-Davis)

216.5


In mice, anoxia or cerebral ischemia is associated with a marked delay in neuronal death. Such delayed death is characterized by (CCl)1/4-induced traumatic brain injury (TBI) in mice, CA1 and CA3 hippocampus both exhibit delayed neuronal death, with DNA damage at 24 hrs, morphological death at 72 hrs, and complete loss of hippocampal parenchyma by 216. We hypothesized that Ad-mediated expression of B-Gal in hippocampus would be attenuated after CCl in mice. Isolation of the adenovirus construct was performed in rabbits, with either CCI to left perihemical cortex or sham injury (Lesion only). Ad-CMV-B-Gal (3 x 10^7 PFU) was then immediately injected into left dorsal hippocampus. At 24 or 72 hrs, mice were sacrificed perfused and B-Gal expression was quantified (enriching protein). Separate mice (n=8) were used to confirm the production of B-Gal staining, in vibratome sections. Robust B-Gal expression in left hippocampus was detected in sham and was similar at 24h (1 8 A (4.4)) vs 72h (1 8 A 9.4). CCI did not reduce B-Gal expression in ipsilateral hippocampus, CA1 or CA3 (n=5), but did reduce B-Gal expression in contralateral hippocampus (1 8 A 2.5) vs 72h (1 8 A 0.6, p<0.05 sham vs CCI). B-Gal was seen in many cell types in ipsilateral hippocampus. Contralateral expression was restricted to ventricle cells and CA3 neurons. Despite the eventual nearly complete loss of ipsilateral hippocampus by 21 days in this model, Ad-mediated gene transfer is robust in this structure early after TBI. This supports the use of this approach to test novel genes targeting hippocampal neuronal death in this model. Inhibition of gene expression in contralateral hippocampus by injury may reflect reduced CSF circulation or failure of neuronal transfer of Ad after CCI. J.She et al. 1997. Watan et al. 1999. Support: NS38181 and GM64490.

216.6


Despite the routine use of fentanyl for initial sedation of patients after severe traumatic brain injury (TBI), it remains to be determined if it is the optimal sedative agent. Isoflurane is the most commonly used anesthetic in experimental models of TBI. Recent studies in experimental cerebral ischemia suggest that isoflurane is neuroprotective (versus fentanyl) in part by increasing cerebral blood flow (CBF) and reducing metabolic demands. To our knowledge, fentanyl has not been directly compared to isoflurane in experimental TBI. We hypothesized that isoflurane would be neuroprotective versus fentanyl when administered early after TBI in rats. Male Sprague-Dawley rats (n=8/group) were subjected to controlled cortical impact to the left perihemical cortex and randomized to receive either fentanyl (100 mcg/kg, bolus followed by a 25 mcg/kg/hr infusion) or isoflurane (1% by inhalation) for 4 hrs. Motor (beam balance, beam walking, 1-5) and cognitive (Morris water maze performance, 14-26) function were used to assess functional outcome and rats were perfused for the assessment of lesion and hippocampal volumes on day 21. Rats treated with isoflurane had marked behavioral and cognitive function compared to those treated with fentanyl (both p<0.05 on multiple days); there were no differences in either control or hippocampal volumes between treatment groups. We speculate that the increase in CBF in concert with metabolic suppression produced by isoflurane may be neuroprotective after TBI. Therefore the use of isoflurane may mask the beneficial effects of novel treatments tested in experimental models of TBI. In addition, fentanyl may mask CBF changes and may even be detrimental in the acute phase after severe TBI. Support: USArmy/DAMD17-97-1-7009

Despite routine use of fentanyl in patients after traumatic brain injury (TBI), it is unclear if it is the optimal analgesic. Isoflurane is routinely used in TBI models. Studies in cold lesion and ischemia suggest isoflurane is neuroprotective vs. fentanyl. To our knowledge, fentanyl and isoflurane have not been compared in TBI. We hypothesize that isoflurane is neuroprotective vs. fentanyl early after TBI. Male Sprague-Dawley rats (n=18) underwent controlled cortical impact and received 4 h of fentanyl (10 mcg/kg bolus, 50 mcg/kg/h infusion) or isoflurane (1% inhalation). Functional outcome (beam balance, beam walking and Morris water maze [MWM] tasks) and lesion volumes were assessed. Motor and MWM performances were better in rats treated with isoflurane vs. fentanyl (p<0.05). Lesion volumes were not different between groups. We speculate that isoflurane may be neuroprotective after TBI by increasing CBF, suppressing metabolism, and/or modulating excitotoxicity. Isoflurane may mask beneficial effects of novel treatments in experimental TBI. Finally, fentanyl may not be the optimal analgesic agent early after TBI in humans. Support: USArmy#DAMD17-97-1-7009.


The objective of this study was to determine whether an episode of hypoxia 24 hr after brain trauma augments histologic injury. Male C57BL/6 mice (n=10) were subjected to controlled impact injury using a deformation depth of 1 mm and impact velocity of 5 m/sec. After recovery for 24 hr, hypoxia was produced by lowering the percentage O2 to 9% for 5 min and 7% for an additional 30 min. After an additional recovery period of 5 days, the animals were perfusion-fixed with FAM, and the brains were embedded in paraffin, sectioned, and stained with acid fuchsin/thionin. The stained sections were examined for histologic alteration and the volume of cortical infarction was measured.

Histopathologic alteration was not detected in any region of the contralateral hemisphere. Hypoxia significantly increased the size of the cortical lesion: Sham-hypoxia = 1.95 ± 0.42 mm3 (mean ± SD, n=5) vs. Hypoxia = 3.15 ± 0.48 (p<0.01). The other only histologic alteration detected was in the dentate granule cell layer of the ipsilateral hippocampus. There was both loss of neurons and acidophilic transformation of neurons in this layer. However, the number of acidophilic dentate granule cells was not altered by hypoxia (Sham-hypoxia = 74 ± 4 vs. Hypoxia = 70 ± 19). These results indicate that the traumatized cortex remains vulnerable for 24 hr to a level of hypoxia which does not cause histologic injury in the contralateral hemisphere.

Supported by NIH Grant NS-08893.

HYPOXIA EXACERBATES CA3 HIPPOCAMPAL NEURONAL DAMAGE AFTER FLUID PERCUSSION BRAIN INJURY IN RATS. Namiko Nomura*, Kojiro Wada, Yoshitomo Matsushita, Hiroshi Nawashiro, Katsui Shima. Dept. of Neurosurgery, National Defense Medical College, Tokorozawa, Saitama, Japan.

We have reported that increased vulnerability of hippocampal CA3 neurons to hypoxia after mild concussion. The present study was designed to determine if a model of moderate fluid-percussion (F-P) brain injury with hypoxia exacerbates hippocampal CA3 lesions, if those lesions are associated with apoptosis using the terminal deoxynucleotidyl transferase-mediated biotin-dUTP nick-end labeling method (TUNEL). Anesthetized Sprague-Dawley rats were injured with a moderate severity fluid percussion pulse (3.5-4.0 atmospheres) administered over the right parietal cortex. The experimental animals were divided into 2 groups, traumatic brain injury (TBI) group (n=6), which was subjected to TBI alone, and TBI + hypoxia group (n=6), which was subjected to TBI followed by 20 min of moderate hypoxia (FiO2: 10%). Three days following TBI, % neuronal density per 1 mm length of CA3 neurons in the ipsilateral hippocampus was significantly decreased in the TBI + hypoxia group (45.2 ± 29.6; p < 0.05) compared to the TBI alone group (90.8 ± 24.1 %). No significant difference in the number of TUNEL positive cells was observed at 6-h, 24-h and 3-day (n=2) in both groups. These results suggest that TBI with moderate hypoxia induced more hippocampal damage due to not only apoptosis but also necrosis.
SOLUBLE FAS IS INCREASED IN CSF FROM INFANTS AND CHILDREN AFTER HEAD INJURY

Neal A. Seidberg, Acad Hosp of Pittsburgh, Pittsburgh, PA; Robert B. Clark, Univ Pittsburgh and SCAF for Resuscitation Res, Pittsburgh, PA; Patrick M Kochanek, SCAF for Resuscitation Res, Pittsburgh, PA; P David Adelson, Margaret A Satchell, Randall R Ruppel, Stephen Wisniewski, Zaichkin Mi, Keri L Janesko, SCAF for Resuscitation Res, Pittsburgh, PA; Robert B Clark, Univ Pittsburgh and SCAF for Resuscitation Res, Pittsburgh, PA; Edwin K Jackson, SCAF for Resuscitation Res, Pittsburgh, PA

Introduction: Fas, a member of the TNF receptor family, and its ligand FasL, provide a system for regulating intercellular programmed cell death (PCD), where binding of FasL to Fas receptor triggers apoptosis. Fas has been identified in neurons and astrocytes, and FasL is present on microglia and inflammatory cells, thus, PCD in brain after injury may be regulated in part by Fas/FasL interactions. Accordingly, increased levels of CSF infant and children after traumatic brain injury (TBI) for alterations in Fas and Fas-L. Methods: CSF was obtained from 20 patients with severe TBI who required neurintensive care including intraventricular catheter placement. Samples (n = 60) were collected on d 1 - 10 and were immediately centrifuged to remove cells. Control CSF was obtained from 14 children without TBI or meningitis. Fas and FasL were measured by ELISA. Results: TBI patients ranged in age from 1 mo - 11 y, 18 survived and 2 died. CSF Fas was increased 3-fold in TBI patients vs control (see table). Post hoc analysis also revealed an association between Fas and age and suspected child abuse. Therapies targeting cell death receptors, such as Fas, may represent effective strategies aimed at reducing PCD after TBI. Support: ROI NS0850, K08 NS01456, & P09 NS02018

<table>
<thead>
<tr>
<th>Group</th>
<th>Fas (pmol/L)</th>
<th>FasL (pmol/L)</th>
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<tbody>
<tr>
<td>Control (n = 14)</td>
<td>258 ± 59</td>
<td>359 ± 97</td>
</tr>
<tr>
<td>TBI (n = 20)</td>
<td>188 ± 43</td>
<td>125 ± 43</td>
</tr>
<tr>
<td>Accidental TBI (n = 16)</td>
<td>660 ± 154</td>
<td>648 ± 45</td>
</tr>
<tr>
<td>Suspected Child Abuse (n = 4)</td>
<td>1370 ± 341</td>
<td>1641 ± 37</td>
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</tbody>
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*n* = SEM; p < 0.05 vs control; t = 0.5 vs accidental

**SYSTEMIC TREATMENT WITH A PAN-CASPASE INHIBITOR IMPROVES HIPPOCAMPAL NEURON SURVIVAL AFTER TRAUMATIC BRAIN INJURY IN MICE**

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Introduction: Traumatic brain injury (TBI) induces cell death both immediately after injury as a result of direct mechanical insult, and in a delayed fashion as a result of secondary injury. Programmed cell death (PCD), or apoptosis, contributes to secondary cell death. The caspase family of cysteine proteases serve as effectors and executors of PCD, and caspase inhibitors reduce cell death in vitro and in vivo. We hypothesized that systemic administration of the pan-caspase inhibitor hepcaspin (N-000 methyl ketone) (BAP) would reduce hippocampal cell death after controlled cortical impact (CCI) in mice. Methods: Anesthesia was induced in subjects to subjective CCI to the left parietal cortex. Immediately after CCI mice were given 100 nmol BAP or vehicle (DMSO) i.p. in a randomized fashion. In one squadron of mice Caspase-3 activity was measured in injured brain at 24 h (n = 3/group). Separate mice underwent motor function tests (beam and round tube balance) at baseline and 24 h after CCI, then were killed for assessment of hippocampal neuron survival and DNA fragmentation using TUNEL (n = 6/group). Results: BAP treatment prevented the increase in relative caspase activity typically produced by CCI vs vehicle (86 vs 174% of uninjured hemispheres, p = 0.04). The number of surviving CA1 hippocampal neurons, cells vulnerable to PCD in this model, were increased in BAP treated mice vs vehicle (247 ± 28 vs 149 ± 13, p = 0.02). TUNEL-positive cells in hippocampi were similar between groups. Motor function was worse in BAP treated mice vs vehicle (p < 0.01). Conclusions: Pan-caspase inhibition systemic treatment with BAP completely inhibits caspase-3 activity and enhances survival of vulnerable neurons 24 h after TBI in mice. Surprisingly, motor function at 24 h after TBI is worsened. This may be due to preservation of dysfunctional neurons, some specific effects of BAP. Additional studies evaluating the long-term effects of pan-caspase inhibition after TBI are ongoing. Further investigation to determine the optimal treatment paradigm targeting caspase inhibition after TBI is warranted. Support: ROI NS0850, and P09 NS02018.

**ISOFLURANE IMPROVES LONG-TERM NEUROLOGIC OUTCOME COMPARED TO FENTANYL AFTER TRAUMATIC BRAIN INJURY IN RATS**

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Introduction: Isoflurane, an inhaled anesthetic, improves cerebral blood flow in patients after traumatic brain injury (TBI), it is unclear if it is the optimal sedative/analgesic agent. Isoflurane is commonly used in models of TBI. Recent studies in cerebral ischemia and focal ischemic lesion suggest that isoflurane may be neuroprotective vs fentanyl when given early after TBI in rats. Methods: Adult rats (n=18) underwent controlled cortical impact (CCI) with physiologic monitoring and then received 4h of NaNO (2:1) and either fentanyl (10 mg/kg bolus, 500 mg/kg/h infusion) or isoflurane (1% inhalation). Sham (n=8) underwent identical preparation and anesthesia but no CCI. Functional outcome (beam balance, beam walking, Morris water maze (MMW) tasks) was assessed on 104 in injured and sham rats. Lesion volume was quantified on d11. Additional rats (n=14) underwent CCI and anesthesia as described above with intracranial pressure (ICP) monitoring (Codman intra- cranial transducer) for 4h. Brain water (wet dry weight method) was assessed at the end of the anesthetic period. Results: After injury, motor and MWM performance were better in isoflurane vs fentanyl treated rats (p<0.05, ANOVA) but did not differ between groups. Lesion volume between groups. There was increased frequency of ICP > 20 mm Hg and higher brain water in rats treated with isoflurane vs fentanyl (p<0.05, ANOVA). Conclusions: Isoflurane treated with isoflurane has improved long-term functional outcomes compared to those treated with fentanyl, despite increases in ICP and brain water. We speculate that isoflurane may mediate improved long-term functional outcome after CCI in rats, by the preservation of cerebral blood flow, suppression of metabolism, and/or modulation of excitotoxicity. Fentanyl may not be the optimal sedative/analgesic agent early after TBI in humans. Support: USA, NIH (DA15725), 1. Mira, et al. Anesthesiology 1996; 89:400. 2. Murr, et al. Anesth Analg 1995; 80: 1108-11.

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