A Rationale for Synchrony Strategies in Chemotherapy

Barry W. Brown and James R. Thompson

Introduction

A synchrony strategy in the treatment of a tumor consists of two parts. First is the introduction of an agent which blocks the tumor cells at some point in the cell cycle. Second is the release of this block until the partially synchronized tumor cell population arrives in a wave at a vulnerable part of the cell cycle where it is treated with a phase specific agent.

The effectiveness of a therapeutic regime against cancer is determined by the amount of damage produced in the neoplastic tissues versus the amount of damage produced in the limiting normal tissues. The limiting normal tissues are those which first express the damage done by the regime and cause a limitation of the treatment. The treatment-limiting cell populations in much of chemotherapy are those with a short cycle time (roughly twenty-four hours) in the bone marrow and the gastro-intestinal tract.

The basic reason for using a synchrony strategy is that it produces a greater kill of neoplastic cells compared to treatment-limiting normal cells than does chemotherapy which does not take account of cellular kinetics. This therapeutic gain is due to the greater average cell cycle time of the malignant cells and to random variation in cell cycle times.

We take the synchrony strategy described by Barranco et al. (1) which was used for the treatment of melanoma as a prototype. Sherakawa et al. (4)

* This work was supported in part by the Office of Naval Research under NR-042-283 and the U.S. Public Health Service under CA-11430.

† Department of Biomathematics, The University of Texas, M.D. Anderson Hospital and Tumor Institute, Houston, TX 77025.

‡ Department of Mathematical Sciences, William Marsh Rice University, Houston, TX 77001.
estimate that the cycle time of melanoma is 74 hours, with the mean time in
the G1, S and G2 phases being 48 hours, 21 hours, and 5 hours respectively.
The synchrony strategy employed was to infuse bleomycin for four days (slightly longer than the mean cycle time of the melanoma cells) thus blocking cycling melanoma as well as normal cells at the S/G2 junction. At the end of this four day period, infusion is ended and the bleomycin is rapidly eliminated from the body. The melanoma nodules are biopsied periodically in order to identify the time when the partially synchronized neoplastic cells enter S-phase. This should be the time required to traverse G2, M, and G1, or about two days. Peaks in labeling index were indeed found between two and three days, (1). These peak labeling indices were up to four times the pre-infusion value, although typically they were only twice the pre-treatment value. When the peak occurs, an S-phase specific agent such as hydroxy urea or Ara-C is given. Twice as many malignant cells were vulnerable as would have been had the synchrony treatment not been employed.

In the same two to three days that the malignant cells are progressing through the cycle to S-phase, the normal cells will have traversed their cell cycle two to three times. Due to random fluctuations in the cell cycle time, these cells will have completely desynchronized. Hence, the killing of these cells should be no more extreme than it would be had the synchrony procedure not been employed.

The therapeutic gain obtained by using synchrony over the phase specific agent without synchrony is of the order of two to one for a single treatment. The gain from several treatments should multiply so that if we had ten such treatments we would expect a gain of about one thousand to one and with twenty treatments, about one million to one.

This analysis ignores the fact that most of the tumor cells are not
cycling. In melanoma, the estimates of the growth fraction of the tumor seem to center around twenty percent (1,4). In the following sections of this paper, we present a simple model of the changes in growth fraction in the tumor with treatment and the consequences of these changes to the possible outcome of a series of treatment utilizing a synchrony strategy.

The model

In Figure 1 we exhibit a simple model of the cycle of a mammalian cell. There is abundant evidence (2,4,5) that as the tumor increases in size the growth fraction, i.e., fraction of cells traversing the cell cycle, decreases. One model discussed by Gilbert and Lajtha (3) proposes that cells which leave the cycle do so after mitosis and enter the cycle only by returning to G1. We refer to the non-cycling cells as being in the G0 phase.

The quantitative nature of the changes in growth fraction as a tumor either grows or recedes under treatment seems not to be completely known. Neither are the mechanisms which cause these changes. However, work by Tanock (5) suggests that the vascularization of the tumor may be an important factor and we base the following considerations on this concept.

We assume that vascularization of the tumor is sufficient to support a particular number of cells in cycle and that this is the number of cells which are cycling prior to treatment. We also assume that the number of cells which can be in cycle does not change appreciably during the course of treatment.

The cells which are not cycling are not participating in the biochemical events necessary for reproduction and consequently are not subject to the disruption of these events by phase specific chemotherapeutic agents. Consequently, these cells are not damaged by the phase specific agents employed.
A RATIONALE FOR SYNCHRONY STRATEGIES IN CHEMOTHERAPY

There are three parameters in our model. The first is the pre-treatment growth fraction. This is held at 0.2 throughout. The second parameter is the proportion of cycling cells killed by each treatment. This parameter is denoted by $k$. Its value is systematically varied between 0.1 and 0.9 in the following. However, in the cited case of melanoma, S-phase constituted around .3 of the cycle time and synchrony strategies somewhat more than doubled the pre-treatment labeling index, so a $k$ of .7 would seem appropriate. The third parameter, $\tau$, is the proportion of additional cells which could be supported in cycle (due to killing of cycling cells by previous treatments) which do in fact transfer from $G_0$ to the cycling state between treatments. An example may make the meaning of this parameter clearer. Suppose that the vascularization is sufficient to support 1000 cells in cycle. Suppose that a treatment kills 700 of these cells. Then 700 additional cells could be supported in cycle, but if $\tau$ is 0.9, then only $0.9 \times 700 = 630$ will transfer from $G_0$ into the cycle before the next treatment. Chart 3 of (1) suggests that $\tau$ is quite high, probably even higher than the natural 1.0 limit suggested by our model. This chart represents data on only one patient, and we take here the more conservative position that $\tau$ is 0.9.

Using these parameters, the outcome of a course of treatment is easily calculated. For each treatment, the number of cycling cells is reduced to $1.0 - k$ of its pre-treatment value. The number of cells which will transfer from $G_0$ to the cycling state is computed using $\tau$ as described above. This sequence of calculations is repeated for each treatment.

Results

We look first at the effect of each of our parameters individually. The pre-treatment growth fraction is extremely important in determining the out-
come of a course of treatments. If each treatment produces a kill, \( k \) of 0.7, and if the transfer parameter is 0.9, then twenty treatments will produce the following surviving fractions of tumor cells: if the initial growth fraction is 0.1, then the surviving fraction is \( 6.2 \times 10^{-5} \); if the initial growth fraction is 0.2 then the surviving fraction is \( 1.2 \times 10^{-8} \); if the initial growth fraction is 0.3, then the surviving fraction is reduced to \( 8.8 \times 10^{-10} \). By our assumptions, a tumor with a growth fraction of 0.1 is about three times the size of a tumor with a growth fraction of 0.3, but this threefold difference in size make a difference of \( 10^5 \) in surviving fraction after a course of twenty treatments.

Figure 2 presents the surviving fraction following a course of twenty treatments as a function of the effectiveness, \( k \), of each treatment. Even in the most optimistic case presented, \( r \) of 0.9, we require that \( k \) be at least 0.6 before we get a therapeutically useful surviving fraction of \( 10^{-6} \). Killing rates of 0.3 or less would have virtually no effect on the tumor.

This points out the importance of a synchrony strategy. If phase specific drugs were used without synchrony, they would have to be applied quite frequently to achieve therapeutic effectiveness. This frequent application would do great damage to the rapidly cycling limiting normal tissues. By using a synchrony strategy in the case considered, we should be able to achieve a surviving fraction in the tumor of close to \( 10^{-8} \). If normal tissues have an S-phase which constitutes .3 of the total cycle, the surviving fraction after twenty treatments would be about \( 10^{-2} \). Synchrony has thus given a therapeutic advantage of \( 10^6 \).

Chart 3 illustrates the effect of the transfer parameter, \( r \), on the surviving fraction following a regime of twenty treatments. For low values of \( r \) the regime has little effect on the tumor and there is little change in
A RATIONALE FOR SYNCHRONY STRATEGIES IN CHEMOTHERAPY

surviving fraction with \( t \). For \( t \) slightly higher than 0.2, there is a distinct increase in treatment effectiveness with an increase in \( t \), although there is a leveling off of this change with still higher values of \( t \). It is difficult to make any inferences of use in therapy from this data although the value of \( t \) will undoubtedly be influenced by the period between treatments.

Figure 4 presents the surviving fraction following each treatment for \( k \) equals 0.7 and the same five values of \( t \) as figure 1. In each case, there is a shoulder where each treatment causes little incremental killing. Following the shoulder there is a rapid decrease in surviving fraction with each successive treatment. Figure 5 makes the reason for this shoulder effect clear: it presents the growth fraction following each treatment. A comparison of the two figures shows that the shoulder lasts until all of the cells are cycling. Before that point, many of the tumor cells are protected by being in the Go state.

It should be noted that chart 3 of (1) indicates that in the one experience in which several synchrony treatments were given to a melanoma patient, it seems that the growth fraction of the tumor rose to quite close to unity after only two treatments, thus the considerations here may be quite conservative.

The following formula may be used to give a comparison on the length of the shoulder using phase specific drugs with and without synchrony:

\[
\text{Shoulder length (without synchrony)} = \text{Shoulder length (with synchrony)} \times \frac{\text{Mean total tumor cycle time}}{\text{Mean time in sensitive phase}}.
\]

Thus in our case with \( k \) equals 0.7 and \( t \) equals 0.9, we see from charts 4 and 5 that the shoulder has a length of seven treatments. Without synchrony, according to the formula, the shoulder would be about three times as long or twenty-one treatments. Should the observation in (1) be true that the observed
shoulder with synchrony is only two treatments, without synchrony we would expect a shoulder of six treatments.

Chart 6 presents the therapeutic ratio obtained by the use of synchrony as a function of the number of treatments in the regime. The three lines were all computed assuming that each treatment killed 0.7 of the tumor cells and 0.3 of the limiting normal cells. The top line was computed under the (optimistic) assumption that all tumor and normal cells are cycling throughout. The middle line was computed assuming that both the tumor and the limiting normal cells behave according to our model. It does seem that bone marrow cells have kinetic behavior that protects from destruction in a way analogous to that discussed for tumor cells. We see that the therapeutic ratio can still rise to $10^6$ in twenty treatments. The shoulder effect is still noticeable -- the therapeutic ratio is small for the first few treatments and rises rapidly thereafter. Should the shoulder in fact be smaller than predicted by the model, the therapeutic ratio would be even higher than given by this curve. The bottom curve was calculated under the (pessimistic) assumption that the tumor behaved according to our model with $r$ of 0.9, but that the limiting normal cells were all cycling throughout treatment. From the graph we see that the first seven treatments yield a therapeutic ratio of less than one -- i.e., more damage is done to normal cells than to the tumor cells. The ratio does rise after that, however, and after twenty treatments reaches $3 \times 10^4$.

Conclusions

Several implications for chemotherapeutic strategies can be drawn:

(1) Induced synchrony seems almost a necessity if a large reduction in tumor size is to be effected by a reasonable number of treatments with phase-specific agents without unacceptable damage to normal tissues. This is emphatically
A RATIONALE FOR SYNCHRONY STRATEGIES IN CHEMOTHERAPY

so if the tumor has a large mass of non-cycling cells.

(2) A course of treatment is more effective on a small tumor in which most cells are cycling than on a large tumor in which most cells are in $G_0$. It is important to treat early.

(3) A synchrony strategy would be expected to be effective on disseminated disease with distant metastases provided that the kinetics in the metases are similar to each other and to those of the primary.

(4) Initial treatments are relatively ineffective in reducing the overall surviving fraction in the tumor compared to later treatments. It is important to have enough treatments.

BIBLIOGRAPHY


Figure 1. A model of the cycle of mammalian cells.

Figure 2. Logarithm to the base ten of the surviving fraction of tumor cells following a regime of twenty treatments, each of which kills a fraction of cycling cells given on the abscissa. The parameter $t$ is the proportion of cells transferred into cycle between treatments. In the case of melanoma, $t$ appears to be at least 0.9. For this case, note the difference in surviving fraction with synchrony where the kill is about 0.7 versus no synchrony where the kill is about 0.3.
A RATIONALE FOR SYNCHRONY STRATEGIES IN CHEMOTHERAPY

Figure 3. Logarithm to the base ten of the surviving fraction of cells following a regime of twenty treatments, each of which kills a proportion of the cycling cells given by $k$. The abscissa is the proportion of non-cycling cells which could be supported in cycle which do become cycling between treatments.

Figure 4. Logarithm to the base ten of the surviving fraction versus the number of treatments. Each treatment kills 0.7 of the cycling cells. The lines represent differing proportions of cells transferring into cycle between treatments. In the case of melanoma, $t$ seems to be at least 0.9. Note the shoulder effect: initial treatments have little effect whereas each subsequent treatment has a pronounced effect.
Figure 5. Growth fraction versus the number of treatments. The reason for the comparative lack of effectiveness of the initial treatments is that most cells are protected by being in a non-cycling state.

Figure 6. Logarithm to the base ten of the therapeutic ratio versus the number of treatments. The therapeutic ratio is the ratio of surviving fraction of treatment limiting normal tissue to that of tumor tissue. It is assumed that each synchrony treatment kills 0.7 of cycling tumor cells and 0.3 of cycling normal cells and that the parameter $t$ is 0.9 for both. The top curve (open circles) is calculated assuming that both tumor and normal tissues are cycling. The middle curve (closed circles) is calculated assuming that both types of tissue behave according to the model. The bottom curve (triangles) assumes that the tumor behaves according to the model whereas the normal cells are cycling. In the last case, seven treatments are required before there is any therapeutic gain but successive treatments greatly increase the gain.