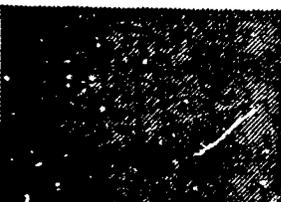


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Leptospirosis in Ceylon - Epidemiological & Laboratory Investigation

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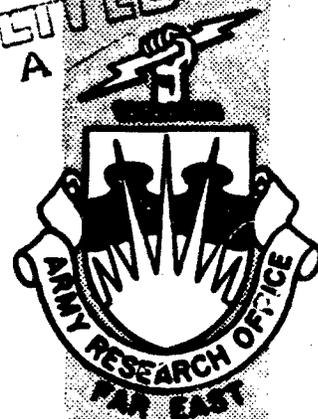
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October 1970



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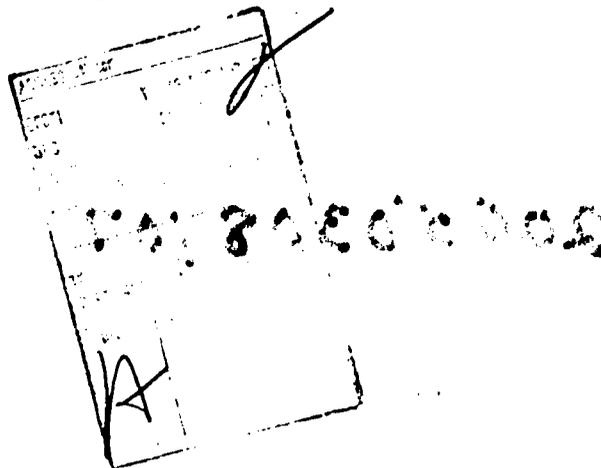
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ABSTRACT :

Serological confirmation using DIFCO commercial antigens, was not obtained in every case of leptospirosis even when the organism was isolated from the blood of the patient. One of the possible reasons could have been that the infecting strains in Ceylon were different from those used in the preparation of the DIFCO antigens. Therefore investigations were undertaken to establish the serotypes prevalent and the vectors for them in Ceylon. The methods adopted were i. human and animal tissues and body fluids were cultured direct; ii. tissues from some animals were passaged through hamsters; iii. surface waters from suspected endemic areas were passaged through hamsters; and iv. serological studies of blood of both humans and animals. Leptospira belonging to the following sero-groups were isolated and vectors for them established; L. javanica; L. icterohaemorrhagiae; L. grippityphosa; L. hebdomadis; L. pomona; and L. autumnalis. The vectors are rodents and dogs. Though large numbers of L. canicola were isolated from patients, no vectors have been established for this. Sera of cattle (30.0 %) and swine (22.1 %) were found to be seropositive for leptospira. The same cattle sera, except for two (2) were seronegative for brucella. These findings are proof that leptospirosis is more widespread among the dairy animals in Ceylon than hitherto recognised and the cause of abortion in the beef herds in our state farms should therefore be more diligently investigated. The isolation of a large number of L. autumnalis and the following hitherto unknown new serotypes, viz: L. icterohaemorrhagiae gem; L. autumnalis lanka; L. grippityphosa ratnapura; L. hebdomadis javawera; L. autumnalis alicia and L. javanica ceylonica: the first five only in the Ratnapura district is an indication of a possible sylvan reservoir which needs further investigation in addition to the wildlife of the rest of the island.

Introduction

Analysis of the indoor morbidity statistics in government hospitals in Ceylon reveals that amongst the largest groups of morbidity, Pyrexias of Unknown Origin ranks second to the big group of bowel diseases. The role of leptospirosis in the group of P.U.O. has not been evaluated. Awareness to this disease in Ceylon was created in 1959 (1). Since then Maretic, Z.; Arumainayagam, P.; Nityananda, K.; Wickremasinghe, R.L. and Ratnatunge, P.C.C., 1962 (2); Nityananda, K. 1962 (3); Rajasuriya, K.; Munasinghe, D.R.; Vitarane, U.T.; Wijesinghe C.P.de S.; Ratnaik, U.T. and Peiris, O.A. 1964 (4); Walloppillai, N.J.; Markhus, H.K.N.I. and Nityananda, K. 1966 (5); Nityananda, K. 1967 (6) and Silva, W.A.S.; Mendis, N.M.P. and Nityananda, K. (unpublished) by their studies have shown that leptospirosis is widely prevalent in this country. Yet the epidemiology of this disease has not been investigated, with the result that the public health importance of it is not appreciated. This is reflected very much in the attitude of most public health and veterinary personnel, who refuse to believe that leptospirosis may be prevalent amongst the livestock of this country to a significant degree as to adversely affect the agricultural economy. Tjalma, R.A. and Galton, M.M. in 1965 (7) while discussing the current concepts regarding the public health significance of this disease observed, "the phenomenal apparent spread and establishment of episootic leptospirosis among livestock has served to create an enormous reservoir and potential source of human infection." In 1954 (7), the United States Department of Agriculture estimated an annual loss to their agricultural economy of over 100 million dollars due to leptospirosis.

Dr. A. Bandaranayake, the Deputy Director, Animal Production and Health, in Ceylon states, the incidence of abortion among the livestock in this country is about 5-10% and this is normal in dairying; the cause according to him is brucellosis (personal communication). Reinhard in 1952 (8) stated that when abortion is the principal sign it may be necessary to obtain laboratory assistance to distinguish leptospirosis from brucellosis, vibriosis or physiological and toxic causes;

Since each leptospiral serotype is believed to have a primary host, it is essential to know the association between the serotypes and their animal hosts in Ceylon before the epidemiology of leptospirosis in this country could be fully understood. This paper describes the work that was conducted to study the vectors and the probable infecting strains of leptospira present in Ceylon.

History of Leptospirosis in Ceylon

The available earliest evidence in record of Weil's Disease having been diagnosed in Ceylon, is in 1953 (9). As the knowledge of the disease increased and with the availability of improved laboratory facilities more cases were diagnosed, confirmed in the laboratory and reported. The majority of the cases were from Ratnapura in the Sabaragamuwa Province, then Ragama, Colombo and Kalutura in the Western Province, Matara in the Southern Province, to a lesser degree from Kandy and Matale in the Central and Anuradhapura in the North Central Province. These reports varied from time to time and place to place depending on the clinician working in that place. In 1959 (3) the first leptospira L. icterohaemorrhagiae was isolated from the blood of a patient in Colombo and soon after from the kidney of a sewer rat trapped in the vicinity of that patient's home. Since then upto 19 leptospiral serotypes belonging to 7 serogroups have been isolated and incriminated as the causative agent of leptospirosis in man and/or animals in Ceylon.

L. autumnalis, the elusive and confounding agent of Fort-bragg fever, is described by some workers outside Ceylon as apparently rare as a human pathogen. In Ceylon however, 56 leptospiral isolates obtained from humans in Ratnapura have been identified and 26 (46.4%) of these belong to the L. autumnalis serogroup.

Source of Infection

For many years rats, dogs and pigs were believed to be the primary animal carriers of leptospira. Today as a result of extensive studies the host range (10) has broadened to such an extent that no living agent, both domestic and wild, can safely be excluded. In addition to animals water too has long been recognised as an important vehicle by which pathogenic leptospira are disseminated and infection transferred from animal carriers to human beings. Three classical examples of water being the source of infection are, the outbreak of Fort-bragg fever in 1942 (11) and those described by Schaeffer in 1951 (12) and Bordjoski in 1952 (13).

A few such episodes have been investigated by the author in Ceylon too. In 1964 (5) some members of a gang of labourers dredging a canal at Wattala were taken ill with leptospirosis. In 1967 at Panadure a party of volunteers composed of civilian and police personnel worked ankle to knee deep to reclaim marshy land, as part of a Shramadana-Campaign (community-

welfare). Out of these, 12 (37.5%) were subsequently taken ill with leptospirosis. Further the majority of the patients investigated at the Ratnapura hospital stated that bathing in the river Kaluganga or panning for gems along the jungle streams as the probable source of their infection.

Materials and Methods

a. Materials

Rodents were trapped alive wherever possible, brought to the leptospirosis laboratory at the Medical Research Institute, anaesthetised and opened up under strict aseptic conditions. Blood was obtained from the heart and kidney plugs taken with sterile pasteur pipettes. Heart blood and kidneys from cattle and swine were taken at the time of their slaughter in the municipal slaughter house in Colombo. Heart blood and kidneys from stray dogs were taken soon after these animals were gassed at the municipal dog pound.

The rodents investigated were; *Rattus norvegicus* (R.n.) - the sewer rat; *Guncmys gracilis* (G.g.) - Ceylon mole rat, also known as the paddy field rat; *Rattus rattus rufescens* (R.r.r.) - Ceylon flat-country house rat; *R.r.kandyanus* (R.r.k.) - Ceylon hill-country house rat; *Bandicoota malabarica* (B.m.) - bandicoot and *Suncus caeruleus giganteus* (S.c.g.) - the shrew, which is a member of the carnivores family.

b. Methods

i. Kidney tissues of all animals were inoculated direct into Fletcher's semi-solid medium, and/or ground up, suspended in sterile normal saline and passaged through hamsters.

ii. Blood was examined for evidence of leptospiral agglutinins from, (a) rat pound workers who are of occupational risk; (b) daily river bathers at Ratnapura which is a suspected endemic area and, (c) cattle, dogs and pigs.

iii. Surface waters from suspected endemic areas (Ratnapura and Ragama) were passaged through hamsters. These hamsters which were inoculated, if they died between the 6th and 14th day after inoculation, were opened up and their liver tissue inoculated into Fletcher's semi-solid medium. No leptospira were isolated from these waters.

Findings

a. Isolations. Kidney tissues from a total of 1611 animals were processed. These included, 592 rodents of 5 species, 3 shrews, 359 cattle, 351 dogs, 293 swine, 5 rock-squirrel, 2 red-mongoose, 2 grey-mongoose, 1 mongoose, 1 hare, 1 civet cat, and 1 field rat. The number of leptospira isolated is shown in table I.

Table I. Leptospira isolated from rodents, shrews and dogs in Ceylon.

	rodent species						dogs
	R.n.	G.g.	R.r.r.	R.r.k.	B.m.	S.c.g.	
Number examined	18	154	117	98	100	3	351
number of leptospira isolated	7	49	0	0	2	1	10

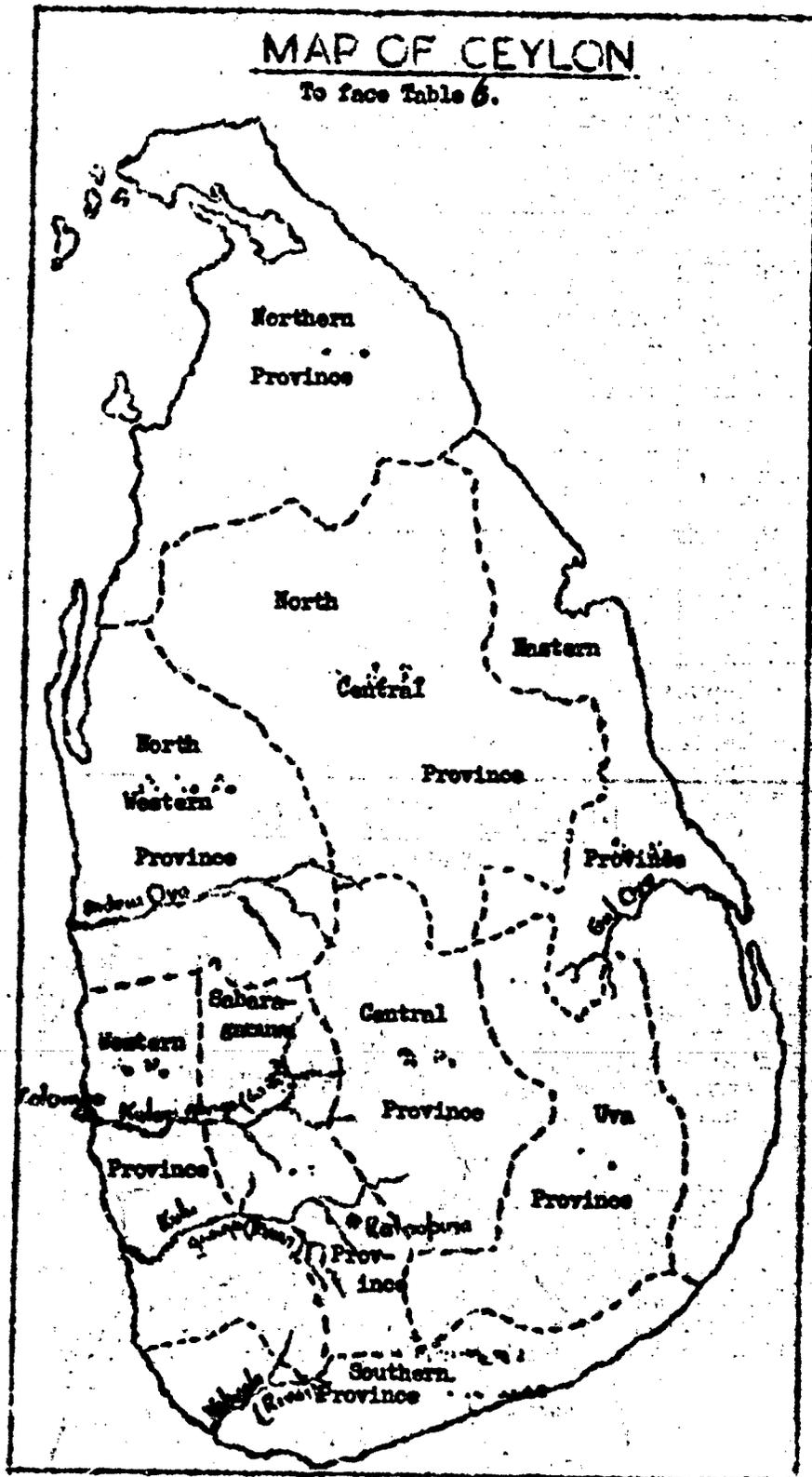
105 unidentified rodents were also examined and 8 isolates were obtained from these.

Table 2. Distribution of the leptospira isolated in Ceylon according to source and sero-group.

Source	sero-groups						
	ict.	can.	jav.	aut.	heb.	grip.	pon.
Human	+	+	±	+	+	+	o
R.n.	+	o	o	o	o	o	o
G.g.	+	o	+	o	o	o	o
B.m.	o	o	+	o	o	o	o
S.c.g.	o	o	±	o	o	o	o
Rodents unidentified	+	o	+	+	o	o	o
Dogs	+	o	+	o	+	o	+

MAP OF CEYLON

To face Table 6.



ict. = icterohaemorrhagiae: can. = canicola: jav. = javanica: aut. = autumnalis: heb. = hebdomadis: grip. = grippotyphosa: and pom. = pomona.
 + = leptospira of that serogroup isolated from that source.
 o = leptospira not isolated.
 ± = two different isolates, but belonging to the same new serotype - L. jav. ceylonica, (14).

Table 3. Distribution of the leptospira isolates according to the provinces in Ceylon.

Name of province	Serogroups							
	ict.	can.	jav.	aut.	heb.	grip.	pom.	wolf.
Western	1/s	1/s	1	1	o	o	1/s	o
Central	1	s	1	o	o	o	o	o
Sabara-gamuwa	1	o	o	ii	1	1	o	o
North - western	o	s	1	o	o	o	s	o
North - central	o	o	o	o	o	o	o	s

i = leptospira of that serogroup isolated in that area.
 ii. = leptospira of that serogroup isolated in that area in large numbers.
 s = serological evidence only.
 o = nil.

b. Serological survey

Sera were examined for the presence of leptospiral agglutinins by the microscopic agglutination (agglutination lysis) test employing live cultures belonging to 21 serotypes, as antigens. Agglutination reaction at a titre of 1:100 or more was considered significant. The employment of multiple screening antigens revealed a significantly high prevalence of sero-reactors.

1. Rat pound workers (rat catchers). The Colombo Municipal Council maintains a regular anti-plague campaign in which a permanent labour force distributes or lays rat cages in the city's sewers, surface drains and house gardens, collect these cages the following morning with the trapped live rodents, transport them in vans to the central rat depot and destroy

them. Most of the Ceylon mole rats, sewer rats, shrews and bandicoots from which there have been a high percentage of leptospira isolations by the author, were supplied alive from this campaign. These labourers walk bare-footed and work with their unprotected hands. They therefore are liable to injury and subsequent contamination directly with the urine of these rodents during collection and transport or indirectly from the polluted floor boards of the vans. Therefore one would expect a reasonable percentage of seropositivity among these labourers. The sera from 26 of these labourers who had been engaged in this work over several years were examined and all were seronegative. These findings are similar to those of Dr. J.C. Broom (15) and Dr. Jan W. Wolf (15), who investigated inhabitants of rat infested dumps. Both found that though the rats in those areas were carriers of leptospira, none of the human beings living there showed evidence of infection.

ii. Random samples Blood was examined from persons residing in Ratnapura bazaar area, who bathe regularly in the Kaluganga river. Of the 70 sera tested for leptospira agglutinins I9 (27.1%) were seropositive.

iii. Cattle Though no leptospira has been isolated from the kidneys of cattle and swine, there is strong serological evidence that both these are carriers of leptospira in Ceylon.

Out of 359 cattle sera examined 108 (30.0%) were positive for leptospira agglutinins (Table 4). If so small a sample can yield so many positives the problem on a national basis must be one of tremendous proportions. The majority of the seropositives were against *L. wolffi*. All the cattle sera investigated for leptospirosis were also tested for brucellosis and only 2 specimens out of the 359 (0.5%) were positive. Therefore the cause of abortion in the cattle in Ceylon requires a more diligent investigation.

iv. Swine Of the 293 swine sera examined 65 (22.1%) were seropositive (Table 5). The majority of the positive reactors were against *L. pomona* and *L. canicola*. Apparently leptospirosis in swine is more widespread in this country than hitherto recognized. Though serum samples P-209 and P-212 were positive in very high titres, 1:25600, it was unfortunate that no leptospira were isolated from the kidneys of these two animals.

Table 4 Some of the results of serology of Cattle sera (ce

Specimen Nos.	C 4	C 8	C 11	C 13	C 17	C 28	C 33	C 48	C 95	C 108	C 111	C 119	C 120	C 150	C 151	C 157
Antigen		100														
ballum		100			100											
canicola		100														
ictero																
bataviae			100					100								
grippe																
pyrogenes																
autumnalis								400								
pomona	100			400								400				
wolffi		400	1600	100	100					100	100		100	400	100	400
australis																
tarassovi								400		100	100					
LT.117																
fort bragg	100														400	
hardjo																
Javanica						100	100						100	100		
scntot								100							100	
borineana						100	400						100	100		100
alexii																
djosiman												1600				
cynopteri																100
celladoni								100								

ictero = icterohaemorrhagiae. grippe = grippotyphosa

rology of Gattle sera (Ceylon)

	Q	C	C	S	C	Q	C	C	Q	Q	S	C	C	Q	Q	Q	C	C	C	Q
	111	119	120	150	151	157	164	201	206	213	214	216	217	219	226	227	228	236	247	259
								400												
								400												
								1600												
							100													
							100	100												
														100	100	400				
	400																			
00	100		100	400	100	400			200	1600	400	400	400	400	1600		100	400	100	
										400							100	100		
00	100						100	100												
									100						100					
					400														100	
			100	100																
					100															
			100	100		100			400	100	100		100	100	100	100	100	100	400	
										100		100								
	1600																			400
						100														

grippe = grippotyphosa

ga

Table 5 some of the results of serology of swine sera

Specimen Nos.	P 4	P 11	P 26	P 35	P 39	P 41	P 44	P 45	P 46	P 66	P 76	P 87	P 90	P 97	P 106
Antigen															
ballum					400						400				
canicola	400			100	6400			1600	1600	1600	1600				
ictero					1600		100								
bataviae															
grippe															
pyrogenes	100				6400	100									
autumnalis						100						400	100		
pomona		400	6400		400		6400						1600	400	1600
wolffi															
australis															
terassovi															
LT.117															
fort bragg															
hardjo															
javanica															
sentot															
borincana															
alexii															
djasiman						400									
cynopteri															400
celladoni				400											

ictero = icterohaemorrhagiae. grippe = gripotyphosa

ne gera.

P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P
7	106	115	125	151	161	164	172	194	209	212	214	221	223	232	234	239	253
		100															
		1600										400					
		100			100		100										
																	100
		1600															
			100	100	1600				1600			100					100
00	1600				1600		1600	1600	25600	25600	400	400	1600	100	1600	1600	
						400											100
					400				1600	400		400					400
									6400								100
				100	400	100	400			1600							
											100						
			100	100	400				1600	1600				400			
	400				400				100				100				

9a

otiphosa.

Table 6 Some of the results of serology of Dogs' sera.

Specimen Nos.	D 8	D 14	D 16	D 19	D 21	D 24	D 35	D 72	D 93	D 115	D 119	D 130	D 131	D 150	D 161	D 162
Antigen																
ballum														100		
canicola	400	100	100					1600						100	100	
ictero							1600		1600							
bataviae																
grippo.																
pyrogenes							400									
autumnalis			400	400					100							
pomona																
wolffi																
australis																
tarassovi																
LT. 117													100			
fort bragg			1600						100							
hardjo																
Jav. nice									400					100		
sentot																
borincana												100	1600			
alxi															100	
gasiman																
cynopteri																
celladoni					100	100										

ictero = icterohaemorrhagiae. grippo = grippotyphosa.

D	D	D	D	D	D	D	D	D	D	D	D			
193	194	246	287	292	294	303	304	308	316	327	328			
	100				100									
100			100		400		100	100						
		1600	400	600		100		1600		400	100			
100		100	100		100		100	400						
	100													
				100										
100					400			100	100					
		400												
100					100									

10a

v. Dogs In the case of dogs 351 specimens were processed. There were 10 isolations of leptospira and 31 (8.8%) seropositives (Table 6). Though the majority of the seroreactors were against L. canicola none of the isolates belong to the canicola serogroup.

Table 7. Results of serotyping of isolates and serology from 3 dogs: D-16, D-19 & D-32.

Specimen No.	Results of serology				isolate identified as
	can.	aut.	Pt.br.		
D-16	100	500	1600	negative against all others.	<u>L. hebdomadis</u>
D-29	negative against all serotypes.				<u>L. javanica</u>
D-32	- do -				<u>L. pomona</u>

The leptospira isolated from the kidney of dog -16 has been identified as L. hebdomadis whereas the serum from the same animal is negative against the homologous antigen, but positive against L. canicola, L. autumnalis and L. Fort bragg antigens. Further the leptospirae isolates from the kidneys of dog-29 and 32 have been identified as L. javanica and L. pomona respectively, but the sera from these animals are seronegative against the full range of antigens belonging to the 21 serotypes including the homologus.

Discussion

Leptospirosis is associated with a broad animal host spectrum and is transmitted from the animal carriers to other animals and man. In the investigation of possible reservoirs among wild and domestic animals, serological investigation alone may provide useful clues; at the same time they may provide an erroneous or misleading index of infectivity rates, since some seronegative animals may be carriers. This limitation is very clearly shown in table 7: Dogs - 29 and 32. The total absence of antibodies in carriers as in these dogs, though cannot be satisfactorily explained, has been met with. Mrs. Sulzer from the leptospirosis unit of the National Communicable Diseases Centre in Atlanta states, 'we once had a laboratory dog that had no antibodies but we isolated leptospira from the urine of this dog for once a month for as

long as one year after disease' (personal communication).

Almost all homes in Ceylon are rat infested, either with *R.r.rufescens* or *R.r.kandyanus*. From the findings of this study it is evident that the above two species of *rattus* are not vectors for *leptospira* in Ceylon; a finding of great relief indeed. The demonstration of *L.pomona* infection in the dog prompts the potential role of the dog in the infectious cycle of human and animal leptospirosis. In addition to *L.canicola*, *L.icterohaemorrhagiae* and *L.pomona*, dogs can be infected and can serve as a potential source of infection with numerous other serotypes. These include *L.hebdomadis*, *L.javanica* (Table 2); *L.australis*, *L.autumnalis*, *L.bataviae*, *L.medinensis* and *L.hyois* 1957 (16).

While *L.icterohaemorrhagiae*, *L.javanica*, *L.canicola*, *L.autumnalis*, *L.hebdomadis*, *L.grippotyphosa*, *L.pomona* and *L.wolffi* appear to be the primary serotypes involved in human and domestic animal infection in Ceylon, the detection of new serotypes hitherto unknown; viz: *L.ictero. gen.* (17), *L.jav. ceylonica*, *L.grippe. ratnapura* (18), *L.hebdo. jayawera* (18), *L.autum. alicia* (18) and *L.autum. lanka* (19), emphasizes the need for public health, medical and veterinary laboratories to maintain a greater awareness to this disease. Further study of the new feral mammals will be necessary to determine the prevalence of infection in other parts of the island and to determine their role in the epidemiology of this disease.

Summary

The evidence established so far shows that in Ceylon, rodents, dogs, cattle and swine are carriers of *leptospira*. The infecting strains are *L.icterohaemorrhagiae*, *L.canicola*, *L.grippotyphosa*, *L.hebdomadis*, *L.javanica*, *L.autumnalis*, *L.pomona* and *L.wolffi*.

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without whose assistance this study could not have been attempted. Finally the author thanks the United States Army Research and Development Group (Far East) for their encouragement and aid which made this study possible.

References.

1. Falisevac, J. and Arumainayagam, P. 1959. Some Aspects of Pyrexias of Unknown Origin in Ceylon. Transactions of the Society of Medical Officers of Health, Ceylon. 18.
2. Maretic, Z.; Arumainayagam, P.; Nityananda, K.; Wickremasinghe, R.L. and Ratnatunge, P.C.C. 1962. Investigations of Pyrexias of Unknown Origin in Ceylon. A Preliminary Report. Cey. Med. J. 7, 2.
3. Nityananda, K. 1962. Isolation of Leptospira in Ceylon. Cey. Med. J. 7, 2.
4. Rajasuriya, K.; Munasinghe, D.R.; Vitarane, U.T.; Wijesinghe, C.P. de S.; Ratnaike, U.F. and Peiris, O.A. 1964. Leptospirosis in Ceylon-A Clinical Study. Cey. Med. J. 9, 2-3.
5. Walloppillai, N.J.; Marknis, H.K.N.I. and Nityananda, K. 1966. Leptospirosis in Ceylon. Cey. Med. J. 11, 1.
6. Nityananda, K. 1967. Leptospirosis-Serological Survey of Occupational Groups in Ceylon. J. Trop. Med. & Hyg. 70. 10.
7. Tjalma, R.A. and Galton, M.M. 1965. Human Leptospirosis in Iowa. Am. J. Trop. Med. & Hyg. 14.3
8. 15. Symposium on the Leptospirose. 1952. Medical Science Publication No. 1. Army Medical Service Graduate School, Walter Reed Army Medical Centre, Washington, D.C.
9. Administration Report of the Director of Health Services for the Year 1953. Ceylon Government Press.
10. Galton, M.M. 1966. Leptospiral Serotype Distribution Lists according to the hosts and countries. Geneva, World Health Organisation.
11. Gochanour, W.S. Jr., Smadel, J.E., Jackson, E.B., Evans, L.B. and Yager, R.H. 1952. Leptospiral Etiology of Fort Bragg Fever. Pub. Health Rep. 67. 8118.
12. Schaeffer, M. 1951. J. Clin. Invest. 30. 670.
13. Bordjovski, M. 1952. Vojnosanitotaki. Pregled. 9.
14. Nityananda, K. and Catherine R. Sulzer. 1969. A New Leptospiral Serotype in Javanica Serogroup from Ceylon. Trop. Geogr. Med. 21. 207.
15. Alexander, A.D., Gleiser, G.A., Malnati, P. and Yoder, M. 1957. Observations on the Prevalence of Leptospirosis in Canine Populations of the United States. Am. J. Hyg. 65. 43.

16. Nityananda, K. and Catherine R. Sulzer. Isolate confirmed by Dr. A.D. Alexander - WHO Leptospirosis Reference Laboratory, Walter Reed Army Medical Centre, Washington, D.C.
17. Cerzuba, Ju. G., Kokovin, I.L., Saharceva, T.F., Nityananda, K., Silva, V., Mendis, N., Pereira, N. 1968. Fifth Information Exchange in Leptospirosis, WHO.
18. Nityananda, K. and Catherine R. Sulzer. Paper submitted for publication to the Journal of Tropical Geographical Medicine.

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LEPTOSPIROSIS IN CEYLON-EPIDEMIOLOGICAL & LABORATORY INVESTIGATION (U)

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13. ABSTRACT
Investigations were undertaken to establish the serotypes prevalent and the vectors for them in Ceylon. The methods adopted were: i. human and animal tissues and body fluids were cultured direct; ii. tissues from some animals were passaged through hamsters; iii. surface waters from suspected endemic areas were passaged through hamsters; and iv. serological studies of blood of both humans and animals. Leptospira belonging to the following sero-groups were isolate and vectors for them established; L. javanica; L. icterohaemorrhagiae; L. grippityphosa; L. hebdomadis; L. pomona; and L. autumnalis. The vectors are rodents and dogs. (Author)

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