AWARD NUMBER: W81XWH-15-2-0046

TITLE: VIPER: Chronic Pain after Amputation: Inflammatory Mechanisms, Novel Analgesic Pathways, and Improved Patient Safety

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Chronic pain is a significant problem after nerve injury from trauma or surgery. Current therapies and attempts at prevention have proven largely ineffective. Through analysis of data obtained in the Molecular Signatures of Chronic Pain Subtypes study termed Veterans Integrated Pain Evaluation Research (VIPER) (W81XWH-11-2-0003) we have discovered two novel pain pathways with potential therapeutic relevance (Wnt and TGR5). In addition, we recognize that improving the safety and efficacy of existing therapies must continue to be a priority and plan to use the large pharmacogenomic database at Vanderbilt University to identify patients at risk for adverse opioid related events. The current proposal intends to study the contribution of non-neuronal immune cells (macrophages) to chronic pain while also evaluating novel analgesics in relevant animal models. The current proposal also attempts to determine the optimal patient population for opioid therapy while identifying those patients at greatest risk from opioids.

15. SUBJECT TERMS
Post-amputation pain, Phantom limb pain, Residual limb pain, neuropathic pain, novel analgesics, opioid related adverse events.
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INTRODUCTION:
Chronic pain is a significant problem after nerve injury from trauma or surgery. Current therapies and attempts at prevention have proven largely ineffective. Through analysis of data obtained in the Molecular Signatures of Chronic Pain Subtypes study termed Veterans Integrated Pain Evaluation Research (VIPER) (W81XWH-11-2-0003) we have discovered two novel pain pathways with potential therapeutic relevance (Wnt and TGR5). In addition, we recognize that improving the safety and efficacy of existing therapies must continue to be a priority and plan to use the large pharmacogenomic database at Vanderbilt University to identify patients at risk for adverse opioid related events. The current proposal intends to study the contribution of non-neuronal immune cells (macrophages) to chronic pain while also evaluating novel analgesics in relevant animal models. The current proposal also attempts to determine the optimal patient population for opioid therapy while identifying those patients at greatest risk from opioids.

KEYWORDS:
Post-amputation pain, Phantom limb pain, Residual limb pain, neuropathic pain, novel analgesics, opioid related adverse events.

ACCOMPLISHMENTS:
What were the major goals of the project?

Goal 1: Characterize the role of Wnt signaling in macrophage polarization, mouse nerve injury models and human neuroinflammation.

Major Task 1: Characterize macrophage polarization changes after Wnt signaling modification in mouse macrophage cell culture – 0% complete

Major Task 2: Determine the specific wnt pathway responsible for prevention of mechanical allodynia in a mouse model of peripheral nerve injury and correlate this with macrophage polarization state and IL-6 to IL-10 ratio – 20% complete. We have collected plasma samples from mice with SNI and will now perform ELISA for IL-6 and IL-10 both at Duke and UHSHS.

Major Task 3: Characterize wnt pathway expression and DNA methylation changes in humans before and after amputation and determine the role of cytokine ratio measurement in prediction of pain phenotype - 30% complete

We have completed initial qPCR analysis of VIPER patient plasma looking at a number of wnt pathway constituents. This work was done by the lab at UHSHS and the results are displayed in Table 1 below. In this patient population there was a significant difference in the CTNNB1 gene with upregulation apparent in patients with pain. Two other wnt pathway constituents had differences between case and control that almost reached significance at the time of enrollment which was 3-18 months after amputation. The presence of one significant and a number of almost significant changes in wnt pathway constituents is exciting and, frankly, somewhat unexpected since the phenotype is already present at the collection time point. We will continue this experiment with VIPER valproate patient plasma using presurgical samples (and the knowledge of who goes on to develop chronic pain) to determine if wnt pathway
expression at the time of injury correlates with propensity to develop chronic pain in the future.

Table 1: Expression analysis of selected wnt pathway constituents in patients with and without residual limb pain enrolled in the Veterans Integrated Pain Evaluation Research (VIPER) study.

<table>
<thead>
<tr>
<th>wnt pathway gene</th>
<th>Case</th>
<th>Control</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>APC</td>
<td>0.928</td>
<td>1.018</td>
<td>0.11</td>
</tr>
<tr>
<td>CTNNB1</td>
<td>0.9</td>
<td>1.018</td>
<td>0.05</td>
</tr>
<tr>
<td>FZD1</td>
<td>0.884</td>
<td>1.024</td>
<td>0.06</td>
</tr>
<tr>
<td>FZD3</td>
<td>1.183</td>
<td>1.081</td>
<td>0.25</td>
</tr>
<tr>
<td>LRP-3</td>
<td>1.152</td>
<td>1.118</td>
<td>0.4</td>
</tr>
<tr>
<td>LRP6-3</td>
<td>1.579</td>
<td>1.267</td>
<td>0.08</td>
</tr>
</tbody>
</table>

Goal 2: Determine the role of TGR5 in astrocyte activation and treatment of mechanical allodynia in a mouse model of neuropathic pain.

Major Task 1: Determine role of TGR5 signaling in astrocyte activation – 30% complete

Astrocyte culture conditions have been worked out. Initial experiment looking at astrocyte activation using the TGR5 agonists deoxycholate and OA did not show differences in activation as our initial experiments did. We will continue this Task and perform using our Roche agonist.

Major Task 2: Determine the role of TGR5 signaling in treating mechanical allodynia in a mouse peripheral nerve injury model - 100% complete

The TGR5 agonist (deoxycholate) reduces baseline mechanical sensitivity in C57Bl6 mice (Figure 1)
We also found that the TGR5 agonist deoxycholate reduces mechanical allodynia in a peripheral nerve injury mouse model most dramatically at 21 days after injury suggesting that the effect is occurring through an astrocyte activation pathway as astrocyte activation occurs in the late stages of neuropathic pain transition (Figure 2).

Most excitingly, the orally available TGR5 agonist obtained from Roche decreases mechanical allodynia after SNI peripheral nerve injury at 7 days after injury (Figure 3).
Goal 3: Use existing data from the Vanderbilt EMR and genotyping repositories to look for associations between genetic variants and pain phenotypes

Major Task 1: Preliminary analyses conducted to confirm the precise numbers of patients for whom there are sufficient data available. Validation of previously published genotype-phenotype associations – 30% complete

With the Vanderbilt team, we have begun to tackle the most difficult part of this aim which is to identify patients who have had opioid related adverse events in hospital. Once these patients are identified, SNPs associated with risk can be defined using the existing dataset. Below is a brief overview of our selection criteria. This will be expanded upon in the next two quarterly reports. I also have a meeting set up with Vanderbilt in January to discuss patient selection for opioid adverse events.

Outcome =

**PRIMARY OUTCOME = MAJOR Adverse events PROBABLE = A + B**

**SECONDARY OUTCOME = MAJOR Adverse events POSSIBLE = B**

\[ A = \text{MAJOR ADVERSE OUTCOME OCCURRED} = \{\text{DEATH OR CARDIAC/ RESPIRATORY ARREST OR ICU ADMISSION OR RESPIRATORY SUPPORT OR COMPLICATIONS OF SURGICAL AND MEDICAL CARE NOT ELSEWHERE}\} \]

Death => (based on UB-04 Discharge Status Code).

Cardiac/Respiratory Arrest => ICD-9 Diagnoses Codes 427.5 / 799.0 / 799.1 OR ICD-9 Procedure Codes 99.60/ 99.62 / 93.93 OR CPT Code 92950 (CPR) / or ICD9 V Code V 12.53

ICU Admission = Charges for ICU Admission
Respiratory Support = intubation (96.04) or non-invasive ventilation 93.9x or mechanical ventilation 96.7x

or complications not elsewhere classified (ICD-9 code 997.3 respiratory complications; 997.01 neurologic complications) or hypercapnia (786.09) or acute respiratory failure (518.81, 518.82, 518.83, 519.8) or apnea (786.03) or resp distress (786.09)

B = opioid overdose may have occurred = {use of opioid reversal or diagnosis of poisoning}

Use of opioid reversal = charge for naloxone or ICD-9 code for poisoning by sedatives and hypnotics 967.X or central nervous system depressants ICD-9 code 968.X or psychotropic agents 969.X

Major Task 2: Discovery and validation of novel exomic variants associated with opioid adverse drug events – 0% complete

What was accomplished under these goals?

Overall Progress

We are progressing as expected with the organization, experiments and logistics of this project. Following is a detailed list outlining accomplishments for this quarter.

- In late 2015, Dr. Van de Ven and Dr. Buchheit traveled to Vanderbilt University to meet with Dr. Shaw, Dr. Walsh and Dr. Bruehl over the course of two days to design the algorithms necessary to accurately identify opioid related adverse events from the Vanderbilt database. During our more recent visit to Vanderbilt, we also collaborated on a VIPER cytokine analysis paper that was accepted by the journal PAIN this month.
- In our Duke laboratory, we completed the Material Transfer Agreement and received the Roche TGR5 agonist necessary to complete specific aim 2. We have completed study of the effect of this agonist, with intraperitoneal injection, on the baseline paw sensitivity of treated mice.
- In our Duke laboratory, we have reproduced the decrease in baseline mechanical allodynia seen with deoxycholate (A constitutive TGR5 agonist) treatment in mouse paw. See figure above.
- We have found that TGR5 agonist given intraperitoneally reduced mechanical allodynia on day 7 after injury. See figure above.
- We have begun preliminary work with one of the Duke Flow Cytometry facilities to ensure the experimental parameters to complete flow sorting of macrophages are correct.
- UHSHS has been sent the cDNA samples from VIPER study patients for wnt pathway analysis. Because the amount of cDNA from these patients was limited, we decided to save the wnt arrays for the VIPER Valproate samples and to perform targeted wnt pathway analysis on the VIPER samples. They have completed initial qPCR analysis of a number of wnt pathway constituents,
What opportunities for training and professional development has the project provided?
Nothing to Report

How were the results disseminated to communities of interest?
Though we are still in the beginning stages of this work, we have had some interesting results that have been presented in poster and abstract form at our departmental research retreat.

What do you plan to do during the next reporting period to accomplish the goals?

**Description of work to be performed/completed during the next reporting period**
During the next three months, we expect:

- Final ACURO animal protocol approval
- Complete quantitative PCR analysis of the first round of human amputee blood samples to evaluate wnt pathway gene expression changes
- Begin ELISA based analysis of macrophage and astrocyte cultures to determine the effects of TGR5 agonists and wnt agonists on these important cell types.

**IMPACT:**
Describe distinctive contributions, major accomplishments, innovations, successes, or any change in practice or behavior that has come about as a result of the project relative to:

What was the impact on the development of the principal discipline(s) of the project?
Pain medicine is limited by the limited number of new analgesics and adverse effects of opioids. Over the past year we have confirmed that TGR5 is important in both inflammatory pain and neuropathic pain and that a TGR5 agonist reduces the sensitivity in an animal pain model. There is a long road before therapies like this can be used in humans but we are taking the first steps.

What was the impact on other disciplines?
Nothing to Report

What was the impact on technology transfer?
Nothing to Report

What was the impact on society beyond science and technology?
We are still early in this project and results have not been published although there is preliminary evidence that TGR5 agonists are able to treat mechanical allodynia in mice.

**CHANGES/PROBLEMS:**
Nothing to report

Changes in approach and reasons for change
There were no significant changes in approach. Minor changes include the method of delivery of TGR5 agonists. We have decided to use intraperitoneal dosing for two reasons: 1) mice treated intrathecally were behaving strangely and we were concerned the solvent needed to dissolve the TGR5 was causing neurotoxicity 2) the intraperitoneal approach (equivalent to IV in humans) will be a more convenient treatment site when these therapies are eventually tried in human trials.

Other minor changes include additional validation of flow cytometry results using qPCR of known targets that distinguish M1 from M2 phenotype in macrophages.

Actual or anticipated problems or delays and actions or plans to resolve them
ACURO approval was delayed, but this did not delay completion of animal work as we used other funding sources to continue our IACUC approved protocols.
Astrocyte culture was unsuccessful for the first 2 attempts. The problem turned out to be a simple error in that the growth media being used did not contain enough glucose. This problem has been resolved and astrocyte cultures are now growing, but ELISA work has not yet begun.

Changes that had a significant impact on expenditures
No significant changes on expenditures

Significant changes in use or care of human subjects, vertebrate animals, biohazards, and/or select agents
No changes or deviations

PRODUCTS:
Published


Books or other non-periodical, one-time publications.
Nothing to report

Other publications, conference papers, and presentations.
Submitted:

Website(s) or other Internet site(s)
Nothing to report

Technologies or techniques
Nothing to report

Other Products
Nothing to report

PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS
What individuals have worked on the project?

Name: Thomas Van de Ven
Project Role: Principal Investigator
Nearest person month worked: 4.58
Contribution to Project: Coordinates all aspects of the project and assumes overall responsibility for its success.

Name: Ru-Rong Ji
Project Role: Co Investigator
Contribution to Project: He is responsible for interpreting and troubleshooting the proposed animal behavioral testing and cell culture experiments and his lab provides deep expertise in all experimental procedures.

Name: Alexander Chamessian
Project Role: Graduate Student
Nearest person month worked: 12
Contribution to Project: He is responsible, along with Dr. Van de Ven, for completion of all animal behavior and cell culture experiments.

Name: Rachel Morales
Project Role: Program Manager
Nearest person month worked: 1.80
Contribution to Project: Overall project manager for all aspects of the proposal, including coordination of the biological samples, shipment of samples between sites and data organization, and ensures that the supplies are ordered and available.

Name: Thomas Buchheit
Project Role: Co Investigator
Nearest person month worked: 0.24
Contribution to Project: Works closely with Dr. Van de Ven on all aspects of the project
Funding Support: Other resources

Has there been a change in the active other support of the PD/PI(s) or senior/key personnel since the last reporting period?
Nothing to Report

What other organizations were involved as partners?
Organization Name: Vanderbilt University Medical Center
Location of Organization: 1161 21st Avenue South, Nashville, TN 37232-2520
Partner's contribution to the project: Collaborated in the research

Organization Name: Henry M. Jackson Foundation for the Advancement of Military Medicine Inc.
Location of Organization: 6720 A Rockledge Drive, Bethesda, MD 20817
Partner's contribution to the project: Collaborated in the research

SPECIAL REPORTING REQUIREMENTS
QUAD CHARTS:
Attached

APPENDICES:
Attachment 1- Quad Chart
Attachment 2- Manuscript
VIPER II: Chronic Pain After Amputation: Inflammatory Mechanisms, Novel Analgesic Pathways, and Improved Patient Safety.

PI: Van de Ven, Thomas  
Org: Duke University  
Award Amount: $1,500,000

Study Aims

**Problem:** Current therapies for residual limb pain are ineffective or produce significant side effects.

**Hypotheses:** 1) Biomarkers found in the Veterans Integrated Pain Evaluation Research Study (VIPER) will lead to novel analgesics. 2) Pharmacogenomic profiling will improve the safety and effectiveness of current analgesics.

**Approach**
Convergent analysis of VIPER data show both the TGR5 and Wnt pathways to be important in chronic residual limb pain. We will: 1) Define the role of Wnt signaling in inflammation and mechanical allodynia after nerve injury using cell culture and animal models. 2) Test the effectiveness of TGR5 pathway agonists for the treatment of allodynia after nerve injury using animal models. 3) Use human pharmacogenomic predictors to improve the safety and effectiveness of current opioid treatments.

Goals/Milestones

**CY15 Goals**
- Begin macrophage polarization and astrocyte activation experiments
- Begin designing data capture for pharmacogenomic analyses
- Begin animal behavioral TGR5 experiments

**CY16 Goals**
- Begin wnt pathway human gene expression analysis
- Complete cell culture experiments
- Complete pharmacogenomic validation experiments
- Begin pharmacogenomic discovery experiments

**CY17 Goals**
- Complete ELISA and flow cytometry experiments
- Continue animal behavioral testing
- Continue pharmacogenomic discovery experiments
- Complete one manuscript

**CY18 Goals**
- Complete all experiments
- Complete two manuscripts
- Develop follow-on studies and apply for follow-on funding

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<th>16</th>
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<th>18</th>
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<td>Aim 1: Wnt - Cell culture, animal behavioral testing and cytokine measurement.</td>
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<td>Aim 2: TGR5 – Animal behavioral testing and cell culture experiments</td>
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<td>Aim 3: Pharmacogenomic analysis</td>
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PAIN

Differential Expression of Systemic Inflammatory Mediators in Amputees with Chronic Residual Limb Pain
--Manuscript Draft--

Manuscript Number:  
Article Type: Research Paper  
Section/Category: Clinical Science  
Keywords: residual limb pain, chronic post surgical pain, neuropathic pain, inflammatory markers, inflammation  
Corresponding Author: Thomas Van de Ven, M.D., Ph.D.  
Duke University Medical Center and Durham VA Medical Center  
Durham, NC UNITED STATES  
First Author: Alexander Chamessian, B.S.  
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Thomas Van de Ven, M.D., Ph.D.  
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Abstract: Chronic post-surgical pain impacts the majority of amputees, with more than half experiencing neuralgic residual limb pain. The transition from normal acute post-amputation pain to chronic residual limb pain likely involves both peripheral and central inflammatory mechanisms. As part of the Veterans Integrated Pain Evaluation Research (VIPER) study, we investigated links between systemic inflammatory mediator levels and chronic residual limb pain. Subjects included 36 recent active duty military traumatic amputees with chronic residual limb pain and 40 without clinically significant pain. Blood samples were obtained and plasma concentrations of an array of inflammatory mediators were analyzed. Residual limb pain intensity and pain catastrophizing were assessed to examine associations with inflammatory mediators. Pro-inflammatory mediators including TNF-α, TNF-β, IL-8, ICAM-1, Tie2, CRP, and SAA were elevated in patients with chronic residual limb pain. Across all patients, residual limb pain intensity was associated positively with levels of several pro-inflammatory mediators (IL-8, TNF-α, IL-12, TNF-β, PIGF, Tie2, SAA and ICAM-1), and inversely with concentrations of the anti-inflammatory mediator IL-13, as well as IL-2 and Eotaxin-3. Pain catastrophizing correlated positively with IL-8, IL-12, TNF-β, PIGF, and ICAM-1, and inversely with IL-13. Significant associations between catastrophizing and residual limb pain intensity were partially mediated by TNF-α, TNF-β, SAA, and ICAM-1 levels. Results suggest that chronic post-amputation residual limb pain is associated with excessive inflammatory response to injury or to inadequate resolution of the post-injury inflammatory state. Impact of pain catastrophizing on residual limb pain may be due in part to common underlying inflammatory mechanisms.
Cover Letter

Manuscript Title: Differential Expression of Systemic Inflammatory Mediators in Amputees with Chronic Residual Limb Pain

Dear editorial staff of Pain,

Thank you for considering publication of our study examining differences in plasma inflammatory markers in amputees with and without residual limb pain more than three months after amputation. We believe this data provides support for the hypothesis that chronic neuropathic pain is a result of a prolonged neuroinflammatory response by showing that amputees with chronic residual limb pain have an overall pro-inflammatory plasma signature even after wound healing is complete.

- This contribution represents original work that has not been previously published or submitted for publication elsewhere. It has been read and approved by all authors.
- There are no author conflicts of interest to report.
- This work was completed with funding support from
  - Congressionally Directed Medical Research Programs and the Department of Defense award# DM102142, W81XWH-12-2-0129 and W81XWH-15-2-0046
  - T32 NIH grant# 2T32GM008600
  - Reflex Sympathetic Dystrophy Syndrome Association grant

Sincerely,

Thomas Van de Ven, MD,PhD
Abstract

Chronic post-surgical pain impacts the majority of amputees, with more than half experiencing neuralgic residual limb pain. The transition from normal acute post-amputation pain to chronic residual limb pain likely involves both peripheral and central inflammatory mechanisms. As part of the Veterans Integrated Pain Evaluation Research (VIPER) study, we investigated links between systemic inflammatory mediator levels and chronic residual limb pain. Subjects included 36 recent active duty military traumatic amputees with chronic residual limb pain and 40 without clinically significant pain. Blood samples were obtained and plasma concentrations of an array of inflammatory mediators were analyzed. Residual limb pain intensity and pain catastrophizing were assessed to examine associations with inflammatory mediators. Pro-inflammatory mediators including TNF-α, TNF-β, IL-8, ICAM-1, Tie2, CRP, and SAA were elevated in patients with chronic residual limb pain. Across all patients, residual limb pain intensity was associated positively with levels of several pro-inflammatory mediators (IL-8, TNF-α, IL-12, TNF-β, PIGF, Tie2, SAA and ICAM-1), and inversely with concentrations of the anti-inflammatory mediator IL-13, as well as IL-2 and Eotaxin-3. Pain catastrophizing correlated positively with IL-8, IL-12, TNF-β, PIGF, and ICAM-1, and inversely with IL-13. Significant associations between catastrophizing and residual limb pain intensity were partially mediated by TNF-α, TNF-β, SAA, and ICAM-1 levels. Results suggest that chronic post-amputation residual limb pain is associated with excessive inflammatory response to injury or to inadequate resolution of the post-injury inflammatory state. The impact of pain catastrophizing on residual limb pain may be due in part to common underlying inflammatory mechanisms.
Differential Expression of Systemic Inflammatory Mediators in Amputees with Chronic Residual Limb Pain


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8Department of Medicine, Vanderbilt University Medical Center, Nashville, TN
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All the authors have no financial interests in this study.

Number of figures: 2
Number of Tables: 3
Number of words in text: 3428
Number of references: 36

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Disclaimer: The views expressed in this publication are those of the authors, and do not necessarily reflect the official policy of the Department of the Army, the Department of Defense, or the United States Government.
Abstract

Chronic post-surgical pain impacts the majority of amputees, with more than half experiencing neuralgic residual limb pain. The transition from normal acute post-amputation pain to chronic residual limb pain likely involves both peripheral and central inflammatory mechanisms. As part of the Veterans Integrated Pain Evaluation Research (VIPER) study, we investigated links between systemic inflammatory mediator levels and chronic residual limb pain. Subjects included 36 recent active duty military traumatic amputees with chronic residual limb pain and 40 without clinically significant pain. Blood samples were obtained and plasma concentrations of an array of inflammatory mediators were analyzed. Residual limb pain intensity and pain catastrophizing were assessed to examine associations with inflammatory mediators. Pro-inflammatory mediators including TNF-α, TNF-β, IL-8, ICAM-1, Tie2, CRP, and SAA were elevated in patients with chronic residual limb pain. Across all patients, residual limb pain intensity was associated positively with levels of several pro-inflammatory mediators (IL-8, TNF-α, IL-12, TNF-β, PIGF, Tie2, SAA and ICAM-1), and inversely with concentrations of the anti-inflammatory mediator IL-13, as well as IL-2 and Eotaxin-3. Pain catastrophizing correlated positively with IL-8, IL-12, TNF-β, PIGF, and ICAM-1, and inversely with IL-13. Significant associations between catastrophizing and residual limb pain intensity were partially mediated by TNF-α, TNF-β, SAA, and ICAM-1 levels. Results suggest that chronic post-amputation residual limb pain is associated with excessive inflammatory response to injury or to inadequate resolution of the post-injury inflammatory state. The impact of pain catastrophizing on residual limb pain may be due in part to common underlying inflammatory mechanisms.
Introduction

Chronic pain due to nerve trauma is a significant health problem. Among the various origins of such pain, limb amputation stands out as particularly important. In the United States alone, more than 100,000 patients per year undergo amputation due to trauma or medical conditions including diabetes and peripheral vascular disease and the incidence of long-term morbidity due to chronic pain in this population ranges from 50-80% [13,15,36].

Although the precise pathogenesis of chronic pain due to nerve trauma still remains elusive, great progress has been made in recent years in our understanding of this condition. [7,11,25]. There is now abundant evidence from preclinical animal models that the immune system plays a critical role in driving chronic pain[1,6,12,19]. Several human studies have also corroborated the important role of the immune system in various chronic pain states, with a particular focus on systemic inflammatory mediators such as cytokines, chemokines and related molecules. For example, in one study of patients with Complex Regional Pain Syndrome (CRPS), pro-inflammatory mediators such as tumor necrosis factor (TNF) and interleukin-(IL-)2 were found to be significantly elevated compared to controls, while anti-inflammatory mediators such as IL-4 and IL-10 exhibited the opposite trend[30]. Similarly, a study comparing patients with painful vs. non-painful peripheral neuropathies demonstrated that patients with painful neuropathy had elevated systemic TNF and IL-2 (both protein and mRNA) compared to their non-painful counterparts[31].

Given these past findings, we hypothesized that a systemic pro-inflammatory profile is also associated with chronic pain after nerve trauma due to amputation. To address this question, we examined blood samples collected from a cohort of recent active duty military post-traumatic amputees in the Veterans Integrated Pain Amputation Evaluation Research (VIPER) study [4] This study employed a case-control design, with amputees experiencing chronic residual limb pain classified as cases, and amputees with little or no pain designated as controls. Unique features of the VIPER study include the lack of significant co-morbidities in the otherwise young and healthy study cohort, as
well as the fact that both case and control groups experienced the same traumatic injury, minimizing the likelihood that injury status would confound results. The first aim of the present study was to investigate systemic inflammatory profiles in a subset of 76 patients from the VIPER cohort using multiplexed, high-sensitivity, electrochemiluminescent assays.

Pain catastrophizing shows ubiquitous associations with pain intensity across various chronic pain conditions, including phantom limb pain post-amputation [29,33]. However, little is known about links between catastrophizing and chronic post-amputation residual limb pain. While a few studies using evoked pain models have examined possible links between catastrophizing and inflammatory status[8,9]; this issue has received little study in the chronic pain setting [27] A second aim of this study was therefore to examine associations between catastrophizing, chronic post-amputation pain, and systemic inflammatory profiles.

Methods

Design

Data were obtained as part of a larger observational case-control study comparing young recent active duty military traumatic amputees with and without significant pain 3 to 18 months after injury. After enrollment, study subjects provided blood samples and psychometric data were collected. Patients were assigned case or control status based on average pain score over the week prior to enrollment (Figure 1). Blood plasma samples were then sent to the Duke Biomarker Core facility for inflammatory marker detection using the MesoScaleDiscovery System.

Subjects

All study procedures were approved by the Institutional Review Board of Walter Reed National Military Medical Center (WRNMMC). Subjects included 36 cases (as defined below) and 40 controls who had undergone post-traumatic amputations while on active duty. All potential subjects were being treated at WRNMMC and the clinical research was supervised through the Defense and Veterans Center for Integrative Pain Management (DVCIPM – DVCIPM.org), part of the Uniformed Services University.
Subjects were included if they were a military health care system beneficiary aged 18 years or older and undergoing treatment at WRNMMC with a diagnosis of post-injury amputation of all or part of one limb. Amputation injury must also have occurred between 3 and 18 months prior to enrollment. Patients were excluded if they were afflicted with severe traumatic brain injury, significant cognitive deficits, substantial hearing loss, spinal cord injury with permanent or persistent deficits, ongoing tissue damage that might cause pain, infection, heterotrophic ossification, poorly fitting prosthesis, or hip disarticulation.

We defined “Cases” as those with clinically significant pain, defined as an average pain score over the past week of greater than or equal to 3/10 on a numeric rating scale (NRS) (Figure 1). Those patients with clinically significant pain were further adjudicated into pain subtypes. Those subjects reporting no pain or pain less than 3/10 but greater than 0/10 were considered “Controls” (pain subtypes were not analyzed in the latter subgroup). This case/control methodology was chosen to facilitate the separate biomarker discovery and genomic analysis aims of the larger project.

Subject characteristics are summarized in Table 1. There were no significant differences between groups in subject age, BMI, ethnicity, smoking status, or time between injury and enrollment. Patients defined as cases reported significantly higher levels of pain catastrophizing. By study design, cases also had significantly higher average pain scores.

Procedures

After written informed consent was obtained, blood samples were obtained from each patient at one timepoint for subsequent analysis. For preparation of plasma, 6ml of blood was collected in EDTA-containing K2 tubes and inverted to mix. Tubes were then spun at 3,000g for 20 minutes at 4 degrees C. Plasma fraction was collected with a pipette and aliquoted into 1.5ml cryovials and stored at -20 degrees C for 24 hours and subsequently at -80 degrees C.

After blood sample collection, subjects completed the pain and psychometric measures described below.
Measures

Ratings of average pain intensity over the past week were provided by all subjects using the self-report version of the Leeds Assessment of Neuropathic Symptoms and Signs scale [2]. The S-LANSS is a validated measure of pain intensity and neuropathic pain characteristics. Pain intensity on the S-LANSS is rated on an 11-point numeric pain rating scale, anchored with “No Pain” and “Pain As Severe As It Could Be.” Given the current hypotheses and to minimize the number of analyses conducted, data regarding neuropathic pain characteristics from the S-LANSS are not reported here. All subjects also completed the Pain Catastrophizing Scale (PCS), a widely-used and validated measure of pain catastrophizing [20,28] Focus in the current study was on overall level of catastrophizing as reflected in total PCS scores.

Inflammatory Mediator Assays
The Neuroinflammation Panel 1 by MesoScaleDiscovery (MSD #K15210D) was used to quantify 37 acute inflammatory and injury markers in human serum. These sandwich immunoassays consist of five microplates, each pre-coated with capture antibodies on 4 to 10 independent spots and are grouped based on optimal performance in a multiplex panel as follows: Proinflammatory Panel 1 (IFNγ, IL-1β, IL-2, IL-4, IL-6, IL-8, IL-10, IL-13, and TNF-α), Cytokine Panel 1 (IL-1α, IL-5, IL-7, IL-12/IL-23p40, IL-15, IL-16, IL-17a, TNF-β, and VEGF), Chemokine Panel 1 (Eotaxin, MIP-1β, Eotaxin-3, TARC, IP-10, MIP-1α, MCP-1, MDC, and MCP-4), Angiogenesis Panel 1 (VEGF-c, VEGF-D, Tie-2, Flt-1, PIGF, and bFGF), and Vascular Injury Panel 2 (SAA, CRP, VCAM-1, and ICAM-1). Each of these panels is a V-plex assay indicating it is fully validated according to fit-for-purpose principles and the FDA’s analytical validation guidelines, offering highly sensitivity and reproducible results from lot-to-lot. All assays were run according to the manufacturer and samples were run in duplicate. Values below LLOD were defined as ½ LLOD when determining significant differences in inflammatory mediator concentration between cases and controls.

Statistical Analysis
All analyses were conducted using IBM SPSS for Statistics version 23. Initial examination of the distributions of the inflammatory mediators indicated most were not normally distributed. Because of this, we used the nonparametric Mann-Whitney U test for evaluating differences in inflammatory mediators between groups (Case vs. Control) and used nonparametric correlations (Spearman’s rho) for examining associations between residual limb pain levels, pain catastrophizing (PCS scores), and inflammatory mediators.

Because of the unique data available in this study, we evaluated a mediation model in which the association of catastrophizing with chronic post-amputation residual limb pain intensity was conveyed in part via indirect effects of inflammatory mediators. To evaluate this mediation model, the approach of Preacher and Hayes (2004) was used to test the significance of the indirect effects[23]. Custom SPSS dialogue (the Indirect Procedure; http://www.afhayes.com/spss-sas-and-mplus-macros-and-code.html#sobel) was used to test the significance of indirect effects in these models using bootstrapping procedures[23]. This bootstrap methodology tested each mediation model in a series of 1000 random subsamples repeatedly drawn from the full sample, generating 95% confidence intervals (bias corrected) around the indirect effect test statistic. If the 95% confidence interval for the indirect effect generated by the model did not include zero, this indicated that the hypothesized indirect (mediated) effect was significant at the p<.05 level. To minimize the risk of bias in estimation of indirect effects, inflammatory mediator values were normalized via log-transformations prior to conducting mediation analyses.

In preliminary analyses, time (in months) since amputation was examined as it related to the primary outcomes to determine whether it might confound primary analyses. Correlational analyses indicated that pain duration was not associated with either average residual limb pain intensity (Spearman’s rho = -0.07, p = 0.572) or catastrophizing scores on the PCS (Spearman’s rho = -0.13, p = 0.274), nor with most inflammatory mediator values. Exceptions to the latter were: IL-12 (Spearman’s rho = -0.24, p = 0.035), IL-15 (Spearman’s rho = 0.26, p = 0.026), IL-16 (Spearman’s rho = -0.27, p = 0.022), IL-1alpha (Spearman’s rho = 0.27, p = 0.021), and VCAM-1
(Spearman’s rho = -0.23, p = 0.044). Because of the general absence of associations of pain duration with the key outcomes of interest, it was not included as a control variable in the analyses reported below.

Results

Multiple inflammatory mediators are upregulated in amputees with residual limb pain

To determine whether any differences in systemic inflammatory mediators were present between the case and control groups, we measured the levels of 37 inflammatory mediators in all 76 patients. Relative to patients defined as controls, patients defined as cases exhibited significantly higher levels of for TNF-α, TNF-β, IL-8, ICAM-1, Tie2, CRP, and SAA (Table 2). Each of the elevated markers have mainly pro-inflammatory properties. Descriptive statistics for all of the mediators tested are shown in Table 2.

Inflammatory mediators correlate with pain severity and catastrophizing in amputees with significant residual limb pain

A second aim of this study was to examine associations between systemic inflammatory mediators, post-amputation residual limb pain intensity, and pain catastrophizing. Ratings of average residual limb pain intensity and PCS scores were significantly correlated, r (74) = 0.62, p<.001. Table 3 summarizes associations between systemic inflammatory mediators and both residual limb pain intensity and PCS scores. Higher pain intensity was found to be associated with significantly higher levels of IL-8, IL-12, TNF-α, TNF-β, PIGF, Tie2, SAA, and ICAM-1, with inverse associations noted for IL-2, IL-13, and Eotaxin-3. Similarly, higher PCS scores were associated with significantly higher levels of IL-8, IL-12, TNF-β, PIGF, and ICAM-1, with an inverse association observed with IL-13. To address possible inflated type I error due to the number of inflammatory mediators examined, permutation testing (1000 permutations) was conducted to determine empirical probability values for the correlational analyses as a set. As indicated at the bottom of Table 3, set-wise associations with levels of
inflammatory mediators were highly significant for both average pain intensity and catastrophizing. These results indicate that the associations reported between these two variables and inflammatory status are unlikely to represent spurious findings.

Do Inflammatory Mediators Contribute to Associations between Pain Catastrophizing and Residual Limb Pain Intensity?

We considered the possibility that the positive association between pain catastrophizing levels (PCS) and residual limb pain intensity might be accounted for in part by indirect effects conveyed through systemic inflammatory mediators. While the only requirement for conducting analyses to test this model was that catastrophizing needed to be associated with pain intensity, we restricted our analyses to only those inflammatory mediators showing significant associations with the outcome of interest (residual limb pain intensity) to limit the number of analyses conducted.

Results using bootstrapped mediation tests indicated significant indirect effects between PCS scores and residual limb pain intensity via TNF-α (95% CI: 0.0004 – 0.0308), TNF-beta (95% CI: 0.0027 – 0.0309), SAA (95% CI: 0.0005 – 0.0351), and ICAM-1 (95% CI: 0.0011 – 0.0515). In each case, there were also significant direct effects of PCS scores on residual limb pain intensity independent of systemic inflammatory mediators (p’s < .001). Overall, these results indicated that the positive association between pain catastrophizing and residual limb pain intensity was partially mediated by TNF-α, TNF-beta, SAA, and ICAM-1 levels in plasma (model summarized in Figure 2). Tests of indirect effects for other systemic inflammatory mediators showing associations with residual limb pain intensity in correlational analyses were all nonsignificant (p’s > .05).

Discussion

In this study, we found elevated systemic levels of several pro-inflammatory mediators in amputees with residual limb pain (Cases) compared to those without clinically significant residual limb pain (Controls). Specifically, as might be expected given animal work and human findings in other chronic pain populations, the pro-
inflammatory mediators TNF-α, TNF-β, IL-8, CRP, SAA, Tie2 and ICAM-1 were significantly elevated in cases compared to controls.

Across both patient groups, intensity of residual limb pain was associated positively with levels of several pro-inflammatory mediators (IL-8, TNF-α, IL-12, TNF-β, PIGF, Tie2, SAA and ICAM-1), and inversely with concentrations of the anti-inflammatory mediator IL-13, as well as IL-2 and Eotaxin-3. Eotaxin-3, initially thought to have mainly pro-inflammatory properties through agonism of CCR3, more recently was found to be an antagonist for multiple CCR receptors whose blockade prevents chemotaxis [21]. Similarly, IL-2 was initially thought to be mainly pro-inflammatory, stimulating cytotoxic T-cells and NK cells, but was later found to be an important stimulator of T<sub>reg</sub> cells[3]. Taken together, these findings demonstrate an overall pro-inflammatory signature in amputees with chronic pain. To the best of our knowledge, this is the first comprehensive study of systemic inflammatory mediators in human subjects with residual limb pain following amputation.

This study also appears to be the first to examine associations between post-amputation residual limb pain and levels of pain catastrophizing, a cognitive factor previously shown to exacerbate chronic pain intensity across a variety of other pain conditions[24,33]. Results, not surprisingly, indicated that elevated catastrophizing was strongly correlated with greater residual limb pain intensity. Although the causal direction of these effects cannot be ascertained due to the design of this study, the results nonetheless add to the existing literature by extending the apparent negative effects of pain catastrophizing into the post-amputation residual limb pain population.

Finally, the current findings to our knowledge are among the first to systematically examine possible links among catastrophizing, chronic pain intensity, and a comprehensive array of inflammatory mediators. While a limited number of studies have examined associations between catastrophizing and selected inflammatory mediators in the context of laboratory evoked pain stimuli, this issue has received little study in the chronic pain setting[8,9]. The present results revealed that in the context of post-amputation residual limb pain, elevated catastrophizing levels were associated with higher levels of IL-8, IL-12, TNF-β, PIGF, and ICAM-1, and lower levels of IL-13. These findings are consistent with generally pro-inflammatory influences of...
catastrophizing. Interestingly, the strong positive association between catastrophizing and residual limb pain intensity was statistically mediated by TNF-α, TNF-β, SAA, and ICAM-1 levels. In other words, this study suggests the possibility that catastrophizing might be linked with an elevated pro-inflammatory profile, which in turn produces elevated chronic pain intensity. Definitive conclusions regarding this causal model must await replication using a design with evaluation of pain catastrophizing, inflammatory mediator levels, and chronic pain intensity over time. Extending these results to other chronic pain conditions would also be worthwhile.

**Exaggerated pro-inflammatory response in amputees with chronic pain**

Our results demonstrate that there is an exaggerated and enduring pro-inflammatory response in amputees with chronic residual limb pain compared to amputees without pain. This finding is consistent with a substantial body of preclinical evidence suggesting that several of the mediators that were associated with pain in this study may be implicated in the pathogenesis of neuropathic pain. Of these, TNF and IL-6 are the most studied, with these pro-algesic cytokines having pleiotropic effects on neurons and immune cells throughout the neuraxis following nerve injury[16,17,34,35]. IL-8, while not examined directly in nerve injury, has been shown to cause hyperalgesia when administered exogenously to rodents, and has been associated with widespread tenderness to palpation in a large clinical study of TMD sufferers[18,26]. IL-2 and IL-12, have both been shown to be pro-algesic in animal models, and in clinical studies of CRPS and painful small fiber neuropathy, IL-2 was shown to be elevated at the protein and mRNA level from blood samples[30,31].

In further support of an overall pro-inflammatory signature in post-amputation residual limb pain, we report for the first time that IL-13, a cytokine with known anti-inflammatory properties, was negatively correlated with pain. This cytokine is closely related to the anti-inflammatory cytokine IL-4, which has been shown to attenuate neuropathic pain behaviors in animal models of nerve injury [14,32], and has been associated differentially with painless neuropathy in humans [31]. Indeed, the type II IL-4 receptor is the main functional receptor for IL-13, demonstrating the overlapping biological roles of these two cytokines[10]. Given these similarities, one might predict
that IL-13 would also play a role in the modulation of neuropathic pain. Our finding that IL-13 is inversely correlated with average pain scores with moderate effect size and high statistical significance suggests that this may indeed be the case.

Our unbiased analysis additionally allowed us to discover several other mediators not previously associated with post-amputation residual limb pain in clinical or preclinical models. For example, we found that the pro-inflammatory cytokine, TNF-β, which bears many biological similarities to TNF-α, was elevated in the cases and positively associated with limb pain intensity. Given the recognized importance of TNF-α in neuropathic pain, it is likely that TNF-b, which shares the same receptor with TNF-α, may also an important player in the pathogenesis of pain after nerve injury. Other novel associations revealed in our study include the cell adhesion molecule ICAM-1, the pro-angiogenic factors Tie2 and PIGF, the chemokine Eotaxin-3, and the acute phase reactants CRP and SAA, all of which further support the notion of a persistent pro-inflammatory state in amputees with pain.

Of particular interest is CRP, an acute phase reactant that is widely used in clinical practice as a general marker of inflammation. The elevation of CRP in amputees with chronic residual limb pain is consistent with a systemic pro-inflammatory state in this group. Recent work in a large community-based sample of women has reported small but significant positive associations between CRP levels and persistently elevated bodily pain, further supporting the pain relevance of CRP elevations [5]. Of note, CRP is an important risk factor for cardiovascular morbidity, with a value greater than 3 mg/L considered high risk [22]. The median CRP value of ~ 4 mg/L in the case group in the current study suggests these individuals could be at high risk for future cardiovascular disease.

**Limitations**

As a cross-sectional, observational study, this trial was not designed to determine causation with regards to any of the inflammatory mediators that were measured. Furthermore, since we only gathered information on this patient cohort at a single time point, several months after they had suffered injury and experienced chronic pain, we could not examine the temporal dynamics of the inflammatory mediators under study.
Thus, the possibility that the differential mediator profiles in these patients may have been present before injury cannot be excluded. Finally, the conclusions from this study arise from a modest sized cohort of primarily young and male military veterans (n=76), potentially limiting the generalizability of our findings. Given the modest sample size and the relatively large number of associations examined, concerns might be raised as to whether the effects reported might simply be due to inflated Type I error. Results of permutation testing indicating highly significant set-wise correlations between levels of all inflammatory mediators and both residual limb pain intensity and catastrophizing levels argues against our reported findings being spurious.

**Conclusion**

Amputees suffering from residual limb pain exhibit an overall pro-inflammatory signature when compared with amputees without significant pain. A pro-inflammatory profile is associated with both greater pain intensity and higher pain catastrophizing levels. These results generate intriguing hypotheses regarding the links between causation and resolution of the inflammatory state and chronic pain following nerve trauma. The mediators measured here may have utility as potential biomarkers of nerve injury-induced pain.
Acknowledgements

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References


Figure 1. VIPER study flow diagram defining inclusion/exclusion criteria and patient adjudication results.

Figure 2. Model in which effects of catastrophizing on residual limb pain intensity are conveyed through indirect effects of systemic inflammatory mediators.
Examination of plasma inflammatory markers reveals that patients with residual limb pain after suffering post-traumatic amputation while on active military duty have a significant increase in pro-inflammatory markers that correlates positively with pain intensity and pain catastrophizing. In addition, the association between pain catastrophizing and pain intensity was mediated by TNF-α, TNF-beta, SAA, and ICAM-1 levels in plasma.
Veterans Integrated Pain Evaluation Research (VIPER) Study

Patients enrolled 3-18 months after amputation

No Drop-outs

Blood samples obtained, Objective pain scoring performed

Adjudication

Average pain score ≥ 3
Case (n=36)

High sensitivity, quantitative ELISA performed for cytokines and soluble biomarkers

Average pain score < 3
Control (n=40)

Inclusion Criteria:
1. Males and Females > 18 years
2. 1 limb amputation from 3-18 months after injury/surgery

Exclusion criteria:
1. Traumatic Brain Injury
2. Hearing loss
3. Fibromyalgia
4. Cognitive impairment
5. Ongoing tissue damage, infection, poorly fitting prosthesis
6. Spinal cord injury
**Figure 2**

The diagram illustrates a model of pain intensity and chronic residual limb pain through the involvement of TNF-α, TNF-β, SAA, ICAM1, and catastrophizing.

1. **Indirect Effect:**
   - TNF-α
   - TNF-β
   - SAA
   - ICAM1

2. **Direct Effect:**
   - Catastrophizing

3. **Chronic Residual Limb Pain Intensity:**

The arrows indicate the direction of influence between these factors.
Table 1. Characteristics of sample participants from the Veterans Integrated Pain Evaluation Research (VIPER) study.

<table>
<thead>
<tr>
<th></th>
<th>Control (N=40)</th>
<th>Case (N=36)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean (SD)</td>
<td>Mean (SD)</td>
<td></td>
</tr>
<tr>
<td>Age</td>
<td>25.2 (4.8)</td>
<td>27.7 (9.1)</td>
<td>0.1526</td>
</tr>
<tr>
<td>Body Mass Index</td>
<td>25.7 (2.8)</td>
<td>26.8 (3.6)</td>
<td>0.1427</td>
</tr>
<tr>
<td>Time since amputation (months)</td>
<td>7.6 (3.5)</td>
<td>8.9 (5.9)</td>
<td>0.2243</td>
</tr>
<tr>
<td>Smoking (ppd)</td>
<td>0.6 (0.5)</td>
<td>0.6 (0.54)</td>
<td>0.8168</td>
</tr>
<tr>
<td>Average Pain Intensity (0-10)</td>
<td>1.2 (0.73)</td>
<td>5.6 (1.38)</td>
<td>0.000</td>
</tr>
<tr>
<td>Pain Catastrophizing Scale</td>
<td>2.6 (4.53)</td>
<td>11.8 (11.09)</td>
<td>0.000</td>
</tr>
<tr>
<td>Male</td>
<td>40 (100)</td>
<td>35 (97)</td>
<td>0.9577</td>
</tr>
<tr>
<td>Smokers</td>
<td>25 (63)</td>
<td>21 (58)</td>
<td>0.8918</td>
</tr>
<tr>
<td>Ethnicity</td>
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<td></td>
</tr>
<tr>
<td>American Indian/Alaska Native</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>1.0000</td>
</tr>
<tr>
<td>Asian</td>
<td>2 (5)</td>
<td>1 (3)</td>
<td>1.0000</td>
</tr>
<tr>
<td>Native Hawaiian or Other Pacific Islander</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>1.0000</td>
</tr>
<tr>
<td>Black or African American</td>
<td>3 (7)</td>
<td>3 (8)</td>
<td>1.0000</td>
</tr>
<tr>
<td>White</td>
<td>37 (93)</td>
<td>32 (89)</td>
<td>0.8836</td>
</tr>
</tbody>
</table>

Note: Continuous variables were analyzed using t-tests whereas categorical values were examined using a chi-squared test.
Table 2 – Systemic Inflammatory Mediator Concentrations in Cases vs. Controls.

<table>
<thead>
<tr>
<th>Mediator</th>
<th>Case (n=36) Median (Range)</th>
<th>Control (n=40) Median (Range)</th>
<th>Mann-Whitney U Test (p value)</th>
</tr>
</thead>
<tbody>
<tr>
<td>IFN-γ</td>
<td>4.1 (1.5-34.4)</td>
<td>3.5 (1.5-19.1)</td>
<td>0.252</td>
</tr>
<tr>
<td>IL-10</td>
<td>0.3 (0.1-0.9)</td>
<td>0.3 (0.0-3.0)</td>
<td>0.113</td>
</tr>
<tr>
<td>IL-13</td>
<td>0.8 (0.8-0.8)</td>
<td>0.8 (0.8-4.0)</td>
<td>0.099</td>
</tr>
<tr>
<td>IL-1β</td>
<td>0.0 (0.0-0.2)</td>
<td>0.0 (0.0-0.1)</td>
<td>0.859</td>
</tr>
<tr>
<td>IL-2</td>
<td>0.1 (0.1-0.9)</td>
<td>0.1 (0.1-0.8)</td>
<td>0.713</td>
</tr>
<tr>
<td>IL-4</td>
<td>0.0 (0.0-0.1)</td>
<td>0.0 (0.0-0.1)</td>
<td>0.970</td>
</tr>
<tr>
<td>IL-6</td>
<td>0.8 (0.1-10.3)</td>
<td>0.5 (0.1 - 2.6)</td>
<td>0.138</td>
</tr>
<tr>
<td><strong>IL-8</strong></td>
<td><strong>4.7 (2.3-25.2)</strong></td>
<td><strong>4.0 (1.9-9.9)</strong></td>
<td><strong>0.041</strong></td>
</tr>
<tr>
<td>TNF-α</td>
<td>2.2 (1.2-5.0)</td>
<td>1.9 (1.1-3.5)</td>
<td>0.031</td>
</tr>
<tr>
<td>IL-12</td>
<td>140.1 (23.9-289.5)</td>
<td>129.8 (48.5-240.5)</td>
<td>0.131</td>
</tr>
<tr>
<td>IL-17</td>
<td>1.1 (1.1-72.5)</td>
<td>1.1 (1.1-6.5)</td>
<td>0.546</td>
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<tr>
<td>IL-5</td>
<td>0.3 (0.1-3.1)</td>
<td>0.3 (0.1-7.8)</td>
<td>0.983</td>
</tr>
<tr>
<td>IL-7</td>
<td>3.4 (0.8-18.7)</td>
<td>3.5 (0.5-25.5)</td>
<td>0.768</td>
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<tr>
<td><strong>TNF-β</strong></td>
<td><strong>0.2 (0.1-1.1)</strong></td>
<td><strong>0.1 (0.1-0.3)</strong></td>
<td><strong>0.037</strong></td>
</tr>
<tr>
<td>VEGF</td>
<td>43.4 (16.0-309.3)</td>
<td>35.1 (16.5-82.3)</td>
<td>0.065</td>
</tr>
<tr>
<td>IL-15</td>
<td>1.9 (1.2-5.0)</td>
<td>1.8 (1.2-2.6)</td>
<td>0.914</td>
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<tr>
<td>IL-16</td>
<td>183.4 (114.2 - 292.9)</td>
<td>189.8 (93.8-444.2)</td>
<td>0.349</td>
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<tr>
<td>IL-1α</td>
<td>0.9 (0.1-111.5)</td>
<td>1.2 (0.2-12.4)</td>
<td>0.971</td>
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<tr>
<td>Eotaxin</td>
<td>74.7 (24.1-218.9)</td>
<td>86.4 (37.6-211.3)</td>
<td>0.938</td>
</tr>
<tr>
<td>Eotaxin-3</td>
<td>18.8 (11.4-1563.3)</td>
<td>22.3 (4.0-182.6)</td>
<td>0.399</td>
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<tr>
<td>IP-10</td>
<td>252.6 (99.7-11413.7)</td>
<td>209.8 (50.5-561.5)</td>
<td>0.293</td>
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<tr>
<td>MCP-4</td>
<td>45 (14.4-122.4)</td>
<td>52.4 (20.7-107.6)</td>
<td>0.182</td>
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<tr>
<td>MDC</td>
<td>660.4 (296.4-1647.4)</td>
<td>657 (427.7-1362.6)</td>
<td>0.979</td>
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<tr>
<td>MIP-1α</td>
<td>5.5 (5.5-300.4)</td>
<td>5.5 (5.5-625.0)</td>
<td>0.494</td>
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<td>MIP-1β</td>
<td>53.8 (28.5-119.1)</td>
<td>45.0 (22.0-118.0)</td>
<td>0.302</td>
</tr>
<tr>
<td>TARC</td>
<td>58.5 (7.6-247.9)</td>
<td>71.9 (19.3-297.3)</td>
<td>0.163</td>
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<tr>
<td>Flt1</td>
<td>47.5 (22.8-145.0)</td>
<td>47.5 (23.9-171.2)</td>
<td>0.881</td>
</tr>
<tr>
<td></td>
<td>Mean (95% CI)</td>
<td>Mean (95% CI)</td>
<td>p-value</td>
</tr>
<tr>
<td>-------</td>
<td>------------------------</td>
<td>------------------------</td>
<td>---------</td>
</tr>
<tr>
<td>PIGF</td>
<td>28.2 (16.0-39.6)</td>
<td>25.0 (8.6-47.7)</td>
<td>0.056</td>
</tr>
<tr>
<td>Tie2</td>
<td>6384.7 (1602.6-9030.0)</td>
<td>5546.9 (1093.2-8823.6)</td>
<td>0.008</td>
</tr>
<tr>
<td>VEGF-C</td>
<td>43.5 (6.9-157.2)</td>
<td>61.0 (6.9-232.7)</td>
<td>0.112</td>
</tr>
<tr>
<td>VEGF-D</td>
<td>540.4 (124.3-3474.5)</td>
<td>749.6 (120.7-6020.1)</td>
<td>0.172</td>
</tr>
<tr>
<td>bFGF</td>
<td>7.7 (0.4-158.1)</td>
<td>7.2 (1.0-68.6)</td>
<td>0.298</td>
</tr>
<tr>
<td>CRP</td>
<td>4011.5 (62.9-53937.9)</td>
<td>2147.0 (79.7-24416.1)</td>
<td>0.034</td>
</tr>
<tr>
<td>SAA</td>
<td>3878.7 (787.8-316972.6)</td>
<td>1981.9 (371.8-22428.1)</td>
<td>0.002</td>
</tr>
<tr>
<td>ICAM-1</td>
<td>421.2 (264.2-810.3)</td>
<td>379.3 (266.6-607.7)</td>
<td>0.007</td>
</tr>
<tr>
<td>VCAM-1</td>
<td>414.8 (282.0-660.0)</td>
<td>414.9 (257.0-683.8)</td>
<td>0.873</td>
</tr>
</tbody>
</table>

* All concentrations in pg/ml except with the exception of CRP, SAA, ICAM-1 and VCAM-1, which are in ng/ml
Table 3. Spearman correlations between inflammatory mediators, average residual limb pain intensity, and pain catastrophizing in the full sample (n=76).

<table>
<thead>
<tr>
<th>Systemic Mediator</th>
<th>Average Pain</th>
<th>P value</th>
<th>PCS</th>
<th>P value</th>
</tr>
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<tbody>
<tr>
<td>IFN-γ</td>
<td>0.11</td>
<td>0.339</td>
<td>0.18</td>
<td>0.119</td>
</tr>
<tr>
<td>IL-10</td>
<td>0.15</td>
<td>0.191</td>
<td>0.03</td>
<td>0.804</td>
</tr>
<tr>
<td>IL-13</td>
<td>-0.45</td>
<td>0.000</td>
<td>-0.29</td>
<td>0.010</td>
</tr>
<tr>
<td>IL-1β</td>
<td>0.06</td>
<td>0.604</td>
<td>0.13</td>
<td>0.280</td>
</tr>
<tr>
<td>IL-2</td>
<td>-0.24</td>
<td>0.038</td>
<td>-0.15</td>
<td>0.184</td>
</tr>
<tr>
<td>IL-4</td>
<td>-0.22</td>
<td>0.054</td>
<td>-0.06</td>
<td>0.612</td>
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<td>IL-6</td>
<td>0.17</td>
<td>0.137</td>
<td>0.10</td>
<td>0.374</td>
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<td>IL-8</td>
<td>0.26</td>
<td>0.024</td>
<td>0.25</td>
<td>0.030</td>
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<tr>
<td>TNF-α</td>
<td>0.30</td>
<td>0.008</td>
<td>0.18</td>
<td>0.129</td>
</tr>
<tr>
<td>IL-12</td>
<td>0.31</td>
<td>0.006</td>
<td>0.24</td>
<td>0.037</td>
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<tr>
<td>IL-17</td>
<td>0.21</td>
<td>0.065</td>
<td>0.10</td>
<td>0.369</td>
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<tr>
<td>IL-5</td>
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<td>0.706</td>
<td>0.08</td>
<td>0.517</td>
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<tr>
<td>IL-7</td>
<td>-0.03</td>
<td>0.774</td>
<td>0.12</td>
<td>0.293</td>
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<tr>
<td>TNF-β</td>
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<td>0.000</td>
<td>0.39</td>
<td>0.001</td>
</tr>
<tr>
<td>VEGF</td>
<td>0.19</td>
<td>0.104</td>
<td>0.17</td>
<td>0.139</td>
</tr>
<tr>
<td>IL-15</td>
<td>-0.01</td>
<td>0.906</td>
<td>0.02</td>
<td>0.855</td>
</tr>
<tr>
<td>IL-16</td>
<td>0.09</td>
<td>0.439</td>
<td>0.21</td>
<td>0.077</td>
</tr>
<tr>
<td>IL-1a</td>
<td>0.11</td>
<td>0.334</td>
<td>0.03</td>
<td>0.773</td>
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<tr>
<td>Eotaxin</td>
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<td>0.856</td>
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<td>0.017</td>
<td>-0.10</td>
<td>0.385</td>
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<tr>
<td>IP-10</td>
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<td>0.557</td>
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<td>MCP-1</td>
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<td>0.01</td>
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<tr>
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<td>0.358</td>
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<td>0.15</td>
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<td>PIGF</td>
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<td>0.34</td>
<td>0.003</td>
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<td>0.002</td>
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<td>bFGF</td>
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<td>0.270</td>
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<td>Value1</td>
<td>Value2</td>
<td>Value3</td>
<td>Value4</td>
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<td>--------</td>
<td>--------</td>
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<tr>
<td>CRP</td>
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<tr>
<td>SAA</td>
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<td>0.004</td>
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<tr>
<td>ICAM-1</td>
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<td>0.44</td>
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<tr>
<td>VCAM-1</td>
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<td>0.346</td>
<td>0.21</td>
<td>0.074</td>
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<tr>
<td>Set-wise p value</td>
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<td>0.008</td>
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<td>0.005</td>
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